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REFERENCES

1. Haun N, Hooper-Lane C, Safdar N. Healthcare personnel attire and devices as fomites: a systematic review. *Infect Control Hosp Epidemiol* 2016;37:1367–1373.
2. Arnaud I, Maugat S, Jarlier V, Astagneau P, National Early Warning, Investigation and Surveillance of Healthcare-Associated Infections Network (RAISIN)/Multidrug Resistance Study Group. Ongoing increasing temporal and geographical trends of the incidence of extended-spectrum beta-lactamase-producing Enterobacteriaceae infections in France, 2009 to 2013. *Euro Surveill* 2015;19: pii = 20804.
3. Cardoso T, Almeida M, Carratalà J, et al. Microbiology of healthcare-associated infections and the definition accuracy to predict infection by potentially drug resistant pathogens: a systematic review. *BMC Infect Dis* 2015;11:15–565.
4. Huskins WC, Huckabee CM, O'Grady NP, et al; STAR*ICU Trial Investigators. Intervention to reduce transmission of resistant bacteria in intensive care. *N Engl J Med* 2011;364:1407–1418.
5. Tschudin-Sutter S, Sepulcri D, Dangel M, Schuhmacher H, Widmer AF. Compliance with the World Health Organization hand hygiene technique: a prospective observational study. *Infect Control Hosp Epidemiol* 2015;36:482–483.

Multidrug-Resistant Organisms in the Rooms of Patients in Healthcare Facilities

To the Editor—The study by Shams et al¹ adds to the existing body of literature demonstrating the frequent environmental presence of multidrug-resistant organisms (MDROs), including *Clostridium difficile*, in the rooms of patients in healthcare facilities. I commend the authors for their work and wish to make a few comments.

First, it might have been helpful to analyze the data from long-term care facilities and acute-care hospitals separately, given the potential differences in infection control protocols, MDRO prevalence, patient mix, and the variable impact of infection control interventions, including environmental cleaning, on healthcare-associated infections between these 2 types of facilities.^{2,3} Second, it would have been useful to report the breakdown of hospital rooms by specialized units (eg, burn or intensive care units) versus general wards to help the reader determine the generalizability of the results to their specific hospital units. Third, given the varied methods of terminal cleaning of patient rooms (including the adoption of “no-touch” technologies) in response to the ongoing transmission of MDROs in healthcare facilities,^{4,5} further clarification of the methodology and type of cleaning products used

(reported to have been recorded as stated in the Methods section) by participating facilities would have been welcome. Lastly, with 30% of rooms remaining culture positive for MDROs after terminal cleaning—with their attendant risk of transmission to the new occupants⁶—the results of the study by Shams et al support those of prior works demonstrating similarly high rates of MDRO-positive rooms despite seemingly adequate terminal cleaning.^{7–9} Although, as stated by the authors, the relationship between the levels of microbial contamination in the environment and patient acquisition of MDROs remains unclear, one could argue that under the right circumstances in a susceptible host (eg, immunosuppressed or with open wounds), no level of environmental contamination with MDROs in terminally cleaned rooms may be considered safe, and that more effort should be directed now toward devising safe and cost-effective means of eliminating them from the surfaces of all newly vacated patient rooms.

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REFERENCES

1. Shams AM, Rose LJ, Edwards JR, et al. Assessment of the overall and multidrug-resistant organism bioburden on environmental surfaces in healthcare facilities. *Infect Control Hosp Epidemiol* 2017;37:1426–1432.
2. Nicolle LE. Infection control in long-term care facilities. *Clin Infect Dis* 2000;31:752–756.
3. Makris AT, Louise M, Gaber DJ, Richter A, Rubino JR. Effect of a comprehensive infection control program on the incidence of infections in long-term facilities. *Am J Infect Control* 2000; 28:3–7.
4. Rutala WA, Weber DJ. Disinfectants used for environmental disinfection and new room decontamination technology. *Am J Infect Control* 2013;41:S36–S41.
5. Manian FA, Griesnauer S, Bryant A. Implementation of hospital-wide enhanced terminal cleaning of targeted patient rooms and its impact on *Clostridium difficile* infection rates. *Am J Infect Control* 2013;41:537–541.
6. Otter JA, Yezli S, Salkeld JAG, French GL. Evidence that contaminated surfaces contribute to the transmission of

hospital pathogens and an overview of strategies to address contaminated surfaces in hospital settings. *Am J Infect Control* 2013; 41:S6–S11.

7. Manian FA, Griesnauer S, Senkel D, et al. Isolation of *Acinetobacter baumannii* complex and methicillin-resistant *Staphylococcus aureus* from hospital rooms following terminal cleaning and disinfection: Can we do better? *Infect Control Hosp Epidemiol* 2011; 32:667–672.
8. Manian FA, Griesnauer S, Senkel D. Impact of terminal cleaning and disinfection on isolation of *Acinetobacter baumannii* complex from inanimate surfaces of hospital rooms by quantitative and qualitative methods. *Am J Infect Control* 2013; 38:384–385.
9. Eckstein BC, Adams DA, Eckstein EC, et al. Reduction of *Clostridium difficile* and vancomycin-resistant contamination of environmental surfaces after an intervention to improve cleaning methods. *BMC Infect Dis* 2007;7:61.

An Observational Study to Compare Oral Hygiene Care With Chlorhexidine Gluconate Gel Versus Mouthwash to Prevent Ventilator-Associated Pneumonia

To the Editor—Ventilator-associated pneumonia (VAP) is defined as pneumonia that developed 48 hours or longer after the use of mechanical ventilator. Most importantly, it can significantly prolong length of hospital stay and increase mortality of critically ill patients.^{1,2} To prevent this fatal disease, several interventions were initially constituted into the ventilator care bundles by the Institute for Healthcare Improvement (IHI): elevation of head, daily sedation vacation, and assessment of readiness to extubate, daily oral hygiene care, and assessment of stress ulcer and deep venous thrombosis prophylaxis.³ Oral hygiene care using chlorhexidine gluconate (CHG) as an element of the ventilator bundle is supposed to decontaminate the mouth, avoid aspiration of contaminated secretions into the respiratory tract, and prevent VAP.^{4–6} CHG is provided in different formulas such as mouthwash or gel; however, studies comparing the usefulness of these 2 CHG formulas in preventing VAP are lacking. At our institution, we implemented a bundle that included oral hygiene care using CHG mouthwash, and we then changed to CHG gel. We comparatively assessed the effect of CHG gel versus CHG mouthwash on reducing the risk of VAP in the ICU.

This study was conducted in 2 surgical ICUs at a regional teaching hospital that has a total of 26 adult ICU beds and 1 intensivist bed. In these 2 ICUs, a ventilator bundle was implemented in 2015 that included (1) 30°–45° elevation of the head, (2) daily interruption of sedation, (3) daily assessment of readiness to extubate, (4) performance of oral hygiene

care with 0.2% CHG mouthwash 3 times a day, and (5) discharging excess water from the pipeline. In June 2016, the oral antiseptic agent was changed to 0.2% CHG gel, and other care bundles were maintained without change. To evaluate the effect of CHG gel on the reducing risk of VAP, we collected from the infection-control practitioner the numbers of ventilator days and VAP cases monthly between June and December 2016 (ie, gel phase). The rate of VAP was defined as the number of cases of VAP per 1,000 ventilator days. As a baseline measurement for comparison of the effect of oral care of CHG gel versus CHG mouthwash in relation to VAP incidence, we retrospectively collected the same data for January to May 2016 (ie, mouthwash phase).

During the gel period, 5 cases of VAP were recorded, and the total number of ventilator days was 2,724. Overall, the rate of VAP was 1.84 per 1,000 ventilator days. During the mouthwash phase, a total of 5 episodes of VAP were recorded in 1,939 ventilator days, for a VAP rate of 2.58 per 1,000 ventilator days. In the ICU with 16 beds, the rate of VAP declined from 3.08 per 1,000 ventilator days during the mouthwash phase to 2.81 per 1,000 ventilator days during the gel phase. In the other ICU with 10 beds, the rate of VAP declined from 1.55 per 1,000 ventilator days during the mouthwash phase to 0 per 1,000 ventilator days during the gel phase. Additionally, we observed that oral care using the CHG gel took the nurse 15 minutes each time, but oral care required 20 minutes when CHG mouthwash was used. Moreover, the average cost of CHG gel for 1 month is US\$285.28 (8,938 New Taiwan Dollars [NTD]), which is less than CHG mouthwash at US\$622.40 (19,500 NTD).

Although oral hygiene care using CHG can effectively reduce the risk of VAP in critically ill patients from 25% to ~19%,⁷ until now, there has been no evidence regarding which CHG formula, gel or mouthwash, is more cost-effective in the ICU. In this survey, we found that the VAP rate could be reduced after CHG mouthwash was replaced with CHG gel for oral hygiene care. This finding may be explained by the effectiveness of CHG gel for performing oral hygiene in previous studies.^{8,9} A double-blind placebo-controlled multicenter study in ICUs showed that antiseptic decontamination of gingival and dental plaque with a CHG gel significantly decreased the oropharyngeal colonization by aerobic pathogens in ventilated patients.⁸ Another study with handicapped children further confirmed that CHG gel was significantly more effective than either the mouthwash or spray in controlling dental plaque.⁹ Therefore, in line with our finding, oral hygiene care using CHG gel seems to be more effective at reducing VAP than CHG mouthwash.

In addition to the clinical benefit of CHG gel, we observed that the use of CHG gel required less time than CHG mouthwash in oral hygiene care provided by critical care nurses. Therefore, the use of CHG gel is a better choice than mouthwash in clinical nursing practice. Finally, regarding medical cost, we also found the cost of CHG gel to be less than