

## Nucleotides as semiessential nutritional components

A. Sánchez-Pozo\* and A. Gil

Department of Biochemistry and Molecular Biology, Faculty of Pharmacy, University of Granada, 18071 Granada, Spain

Dietary nucleotides are required nutrients for some tissues under certain circumstances. A lack of dietary nucleotides negatively influences protein synthesis in both the liver and the small intestine of rats. Ribosome degradation has been observed as being among the mechanisms responsible for this effect. Dietary nucleotides can also modulate gene expression by interaction with specific transcription factors, in both the liver and the small intestine.

### Dietary nucleotides: Protein synthesis: Gene regulation

#### Introduction

Nucleotides are normal components of the human diet and the body provides mechanisms for their absorption and incorporation into tissues (for a review see Sánchez-Pozo *et al.* 1998). These compounds are not considered to be essential nutrients because they can be synthesized endogenously. In fact, no particular disease has been related to a nucleotide deficiency. However, in certain circumstances, and for some tissues, a lack of dietary nucleotides may impair important functions, suggesting a key nutritional role.

Cellular proliferation, among other biological processes, requires significant amounts of nucleotides. Synthesis of nucleotides from amino acids and phosphoribosylpyrophosphate is an energy consuming process and therefore the utilization of exogenous nucleotides may be beneficial from a bioenergetic point of view. In the absence of exogenous nucleotides the *de novo* synthesis is thought to be activated (Yamaoka *et al.* 1997). Some tissues such as the lymphoid tissue (Perignon *et al.* 1987) or the intestine (Leleiko *et al.* 1983) have a low biosynthetic capacity, probably being dependent on an exogenous supply (Uauy, 1989; Van Buren & Rudolph, 1997).

Dietary nucleotides may also be conditionally essential nutrients in a variety of clinical situations and during development. Thus, it has been shown that they accelerate the recovery of the liver and small intestine after a variety of insults (Núñez *et al.* 1990; Bueno *et al.* 1994; Uauy *et al.* 1994; Jackson *et al.* 1997; Torres *et al.* 1997; Tsujinaka *et al.* 1997; Yamamoto *et al.* 1997). They also promote maturation in both the liver and the small intestine (Uauy, 1989; Carver, 1994; Ortega *et al.* 1995a). Maturation of the small intestine is particularly important in neonates because of their rapid growth, especially in low-birth-weight infants

due to their intrauterine malnutrition. Among other actions, dietary nucleotides have a significant effect in immunity (Carver, 1994; Yamamoto *et al.* 1997; Carver, 1999; Rueda & Gil, 2000). Therefore, nucleotide supplementation of formulas for infant nutrition or for parenteral nutrition is considered beneficial.

The mechanisms by which dietary nucleotides exert their effects are not fully understood. However, there are data to suggest that they affect some biosynthetic processes such as the synthesis of proteins. Furthermore, there is evidence that they can modulate gene expression.

#### Dietary nucleotides and biosynthetic processes

When the rat diet does not contain nucleotides there is a transient decrease in the RNA content of the liver (López-Navarro *et al.* 1995). The lack of dietary nucleotides slightly affects the total concentration of soluble nucleotides, whereas the decrease in RNA is significant. We observed a reduction of the number of ribosomes associated with the endoplasmic reticulum as well as a reduction in the size of the nucleolus of the cells (López-Navarro *et al.* 1996a). Additional experiments showed that these changes are proportional to the nucleotide content of the diet. These findings indicate that ribosome formation is reduced and, what is more remarkable, pre-existing ribosomes are degraded in response to a lack of nucleotides in the diet. We think that these observations indicate a buffering role of RNA, which can protect the cell from nucleotide depletion while the biosynthesis responds to the lack of nucleotides from the diet. This buffering effect is conceivable, as there are no cellular stores of nucleotides. As a consequence, protein synthesis is decreased (López-Navarro *et al.* 1996b).

In the small intestine of rats, we have also found a

**Abbreviations:** CNT1, concentrative nucleoside transporter; IEC, intestinal epithelial cell; HPRT, hypoxanthine phosphoribosyl transferase; mRNA, messenger RNA; SPNT, sodium-dependent purine nucleoside transporter.

\* **Corresponding author:** Dr A. Sánchez-Pozo, fax +34 958 248 960; e-mail sanchezp@platon.ugr.es

decrease in RNA and soluble nucleotides, and consequently in protein synthesis when a nucleotide-free diet is fed. In addition, restriction of dietary nucleotides caused a decrease in DNA content (López-Navarro *et al.* 1996b). The changes were not transient as in the case of the liver and a degree of mucosal atrophy was observed morphologically and enzymatically (Ortega *et al.* 1995a,b).

These findings are consistent with the many studies in which it has been shown that tissue recovery or maturity are positively affected by dietary nucleotides. It is noteworthy that the contribution of dietary nucleotides to maintain nucleic acid levels is dependent on the tissue growth rate, affecting RNA in resting cells such the liver and also DNA in proliferating cells such the intestine.

In conditions of dietary nucleotide restriction, nucleotide diphosphates are significantly reduced in both the liver and the small intestine. These findings point to other relevant biosynthetic effects, as they are involved in many biosynthetic processes such as glycogen synthesis through uridine diphosphate derivatives, phospholipids through cytidine diphosphate derivatives and protein glycosylation. With regard to proteins, we believe that not only protein synthesis but also secretion of proteins may be influenced by dietary nucleotides. This is consistent with many studies showing a reduction of secreted proteins such as apolipoproteins (Morillas *et al.* 1994; Sánchez-Pozo *et al.* 1994, 1995) or immunoglobulins (Navarro *et al.* 1996; Martínez-Augustin *et al.* 1997; Navarro *et al.* 1999) and may explain why dietary nucleotides promote an optimal immune response.

### Dietary nucleotides and gene expression

Two lines of evidence support the idea that dietary nucleotides may exert a direct effect on gene expression. Using isolated nuclei from the small intestine or from an intestinal epithelial cell line (IEC-18) we observed a significant effect of nucleotide availability on the hypoxanthine phosphoribosyl transferase (HPRT) gene transcription rate (Walsh *et al.* 1990). A down regulation of HPRT expression was observed when there were no nucleotides available. Thus, the enzyme responsible for nucleotide salvage is not expressed when there are no nucleotides available. Further characterization of the effect of nucleotides on HPRT expression was conducted by transfecting IEC-18 with several constructs containing deletions of the HPRT promoter and 5' flanking sequences and placing the cells in media containing or lacking nucleotides. A region of 35 bp upstream from the HPRT gene was characterized as the specific responsible *cis*-acting element, which confers sensitivity to nucleotides (Walsh *et al.* 1990). Experiments performed afterwards resulted in the purification of a sequence-specific DNA binding protein of 66 kD, the *trans*-acting element (Walsh *et al.* 1992) with the characteristics of the type of enhancers present in other class II genes. Footprint analysis has mapped the protection from DNAase hydrolysis to a sequence of GTCTGGGT by using both affinity-purified protein and crude nuclear extracts (Walsh *et al.* 1992). Database searches have identified similar sequences of this DNA motif in other genes related to cell growth and proliferation, such as the

ornithine decarboxylase gene. Thus, it is conceivable that many genes that respond to dietary nucleotides may influence cell division.

Experiments with diets containing or lacking nucleotides identical to those used for RNA studies described before, have shown that the genes for both the sodium-dependent purine nucleoside transporter (SPNT) and concentrative nucleoside transporter (CNT1) are modulated by the nucleotide content of the diet (Valdés *et al.* 2000). In the case of the purine-preferring carrier SPNT, mRNA and protein amounts, in both the liver and the small intestine, decreased when no nucleotides were in the diet. A result expected considering the lower need for purine uptake. Interestingly, it has been reported that the expression of this carrier is linked to the cell cycle and in regenerating conditions such as partial hepatectomy (Felipe *et al.* 1997). In the case of the pyrimidine-preferring carrier CNT1, we found additional levels of regulation. Thus, whereas in the liver CNT1 is regulated in a similar way to SPNT, in the intestine, a reduction in the mRNA is observed together with higher amounts of protein. These findings may be explained as a result of post-transcriptional regulation. The opposite behavior of intestine and liver regarding CNT1 expression may be a consequence of the high nucleotide biosynthetic capacity of the liver and the low capacity of the small intestine. Thus, the down-regulation in the liver occurs when no nucleotides are available, whereas there is a higher uptake in the intestine in order to compensate for the low biosynthetic capacity. The regulation of carrier expression by the diet is relevant, as nucleotide carriers participate in the uptake of drugs used in antiviral and cancer therapies.

In conclusion, it is clear that dietary nucleotides influence biosynthetic processes and modulate gene expression, at least of those genes involved in nucleotide metabolism. This high degree of regulation suggests that the uptake and metabolism of nucleotides are of great importance to a number of cell types.

### Acknowledgements

We are indebted to Dr López Navarro, Dr Ortega de la Torre, Dr Pastor-Anglada and Dr Walsh for their significant contributions. This work has been supported by grants of the Spanish Ministry of Education.

### References

- Bueno J, Torres M, Almendros A, Carmona R, Núñez MC, Ríos A & Gil A (1994) Effect of dietary nucleotides on small intestinal repair after diarrhea. Histological and ultrastructural changes. *Gut* **35**, 926–933.
- Carver JD (1994) Dietary nucleotides: cellular immune, intestinal and hepatic system effects. *Journal of Nutrition* **124**, 144S–148S.
- Carver JD (1999) Dietary nucleotides: effects on the immune and gastrointestinal systems. *Acta Paediatrica* **88**, 83S–88S.
- Felipe A, Ferrer-Martínez A, Casado FJ & Pastor-Anglada M (1997) Expression of the sodium-dependent purine nucleoside carrier (SPNT) mRNA correlates with nucleoside transport activity in rat liver. *Biochemical and Biophysical Research Communications* **233**, 572–575.
- Jackson CD, Weis C, Miller BJ & James SJ (1997) Dietary

- nucleotides: effects on cell proliferation following partial hepatectomy in rats fed NIH-31, AIN-76A or folate/methyl-deficient diets. *Journal of Nutrition* **127**, 834S–837S.
- Leleiko NS, Bronstein AD, Baliga BS & Munro HN (1983) De novo purine synthesis in the rat small and large intestine: effect of dietary protein and purines. *Journal of Pediatric Gastroenterology and Nutrition* **2**, 313–319.
- López-Navarro AT, Gil A & Sánchez-Pozo A (1995) Deprivation of dietary nucleotides results in a transient decrease in acid-soluble nucleotides and RNA concentration in rat liver. *Journal of Nutrition* **125**, 2090–2095.
- López-Navarro AT, Bueno JD, Gil A & Sánchez-Pozo A (1996a) Morphological changes in hepatocytes of rats deprived of dietary nucleotides. *British Journal of Nutrition* **76**, 579–589.
- López-Navarro AT, Ortega MA, Peragón J, Bueno JD, Gil A & Sánchez-Pozo A (1996b) Deprivation of dietary nucleotides decreases protein synthesis in the liver and small intestine of rats. *Gastroenterology* **110**, 1760–1769.
- Martínez-Augustín O, Boza JJ, del Pino JL, Lucena J, Martínez-Valverde A & Gil A (1997) Dietary nucleotides might influence immune response against cow's milk proteins in preterm neonates. *Biology of the Neonate* **71**, 215–223.
- Morillas J, Moltó L, Robles R, Gil A & Sánchez-Pozo A (1994) Lipoprotein changes in small-for gestational-age infants fed nucleotide supplemented milk formula. *Acta Paediatrica* **83**, 481–485.
- Navarro J, Ruiz-Bravo A, Jiménez-Valera M & Gil A (1996) Modulation of antibody-forming cell and mitogen-driven lymphoproliferative responses by dietary nucleotides in mice. *Immunology Letters* **53**, 141–145.
- Navarro J, Maldonado J, Ruiz-Bravo A, García-Salmerón JL, Molina JA & Gil A (1999) Influence of dietary nucleotides on plasma immunoglobulin levels and lymphocyte subsets of preterm infants. *Biofactors* **10**, 67–76.
- Núñez MC, Ayudarte MV, Morales D, Suárez MD & Gil A (1990) Effect of dietary nucleotides on intestinal repair in rats with experimental chronic diarrhea. *Journal of Parenteral and Enteral Nutrition* **14**, 598–604.
- Ortega MA, Gil A & Sánchez-Pozo A (1995a) Maturation status of small intestine epithelium in rats deprived of dietary nucleotides. *Life Sciences* **56**, 1623–1630.
- Ortega MA, Núñez MC, Gil A & Sánchez-Pozo A (1995b) Dietary nucleotides accelerate intestinal recovery after food deprivation in old rats. *Journal of Nutrition* **125**, 2090–2095.
- Perignon JL, Bories DM, Houllier AM, Thuillier L & Cartier PH (1987) Metabolism of pyrimidine bases and nucleosides by pyrimidine-nucleoside phosphorilases in cultured human lymphoid cells. *Biochimica et Biophysica Acta* **928**, 130–136.
- Rueda R & Gil A (2000) Influence of dietary compounds on intestinal immunity. *Microbial Ecology Health Diseases* **2**, 146S–156S.
- Sánchez-Pozo A, Morillas J, Moltó L, Robles R & Gil A (1994) Dietary nucleotides influence lipoprotein metabolism in newborn infants. *Pediatric Research* **35**, 112–116.
- Sánchez-Pozo A, Ramírez M, Maldonado J, Gil A, Van Vierlviert JP & Rosseneu M (1995) Dietary nucleotides enhance plasma lecithin cholesterol acyl transferase activity and apolipoprotein A-IV concentration in preterm newborn infants. *Pediatric Research* **37**, 328–333.
- Sánchez-Pozo A, Rueda R, Fontana L & Gil A (1998) Dietary Nucleotides and cell growth. *Trends in Comparative Biochemistry and Physiology* **5**, 99–111.
- Torres MI, Fernández MI, Gil A & Rios A (1997) Effect of dietary nucleotides on degree of fibrosis and steatosis induced by oral intake of thioacetamide. *Digestive Disease Sciences* **42**, 1322–1328.
- Tsujinaka T, Kishibuchi M, Iijima S, Yano M & Monden M (1997) Role of supplementation of a nucleic acid solution on the intestinal mucosa under total parenteral nutrition. *Nutrition* **13**, 369–371.
- Uauy R (1989) Dietary nucleotides and requirements in early life. In *Textbook of Gastroenterology and Nutrition in Infancy*, pp. 265–280 [E Leibel, editor]. New York, NY: Raven Press.
- Uauy R, Quan R & Gil A (1994) Role of nucleotides in intestinal development and repair: implications for infant nutrition. *Journal of Nutrition* **124**, 1436S–1441S.
- Valdés R, Ortega MA, Casado FJ, Felipe A, Gil A, Sánchez-Pozo A & Pastor-Anglada M (2000) Nutritional regulation of nucleoside transporter expression in rat small intestine. *Gastroenterology* **119**, 1623–1630.
- Van Buren CT & Rudolph F (1997) Dietary nucleotides: a conditional requirement. *Nutrition* **13**, 470–472.
- Walsh MJ, Sánchez-Pozo A & Leleiko NS (1990) A regulatory element is characterized by purine-mediated and cell-type-specific gene transcription. *Molecular Cell Biology* **10**, 4356–4364.
- Walsh MJ, Tsao KL & Leleiko NS (1992) Characterization of DNA-protein interactions within a distal regulatory element upstream of a mammalian housekeeping gene promoter. *Journal of Biological Chemistry* **267**, 7026–7035.
- Yamamoto S, Wang MF, Adjei AA & Ameho CK (1997) Role of nucleosides and nucleotides in the immune system, gut reparation after injury, and brain function. *Nutrition* **13**, 372–374.
- Yamaoka T, Kondo M, Honda S, Iwahana H, Moritani M, Ii S, Yoshimoto K & Itakura M (1997) Amidophosphoribosyltransferase limits the rate of cell growth-linked *de novo* purine biosynthesis in the presence of constant capacity of salvage purine biosynthesis. *Journal of Biological Chemistry* **272**, 17719–17725.