



## A review of micronutrients in sepsis: the role of thiamine, L-carnitine, vitamin C, selenium and vitamin D

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### Abstract

Sepsis is defined as the dysregulated host response to an infection resulting in life-threatening organ dysfunction. The metabolic demand from inefficiencies in anaerobic metabolism, mitochondrial and cellular dysfunction, increased cellular turnover, and free-radical damage result in the increased focus of micronutrients in sepsis as they play a pivotal role in these processes. In the present review, we will evaluate the potential role of micronutrients in sepsis, specifically, thiamine, L-carnitine, vitamin C, Se and vitamin D. Each micronutrient will be reviewed in a similar fashion, discussing its major role in normal physiology, suspected role in sepsis, use as a biomarker, discussion of the major basic science and human studies, and conclusion statement. Based on the current available data, we conclude that thiamine may be considered in all septic patients at risk for thiamine deficiency and L-carnitine and vitamin C to those in septic shock. Clinical trials are currently underway which may provide greater insight into the role of micronutrients in sepsis and validate standard utilisation.

**Key words:** Sepsis: Micronutrients: Thiamine: L-Carnitine: Selenium: Vitamin C: Vitamin D

### Introduction

Sepsis is the result of a complex interaction between the host response and infecting organism resulting in life-threatening organ dysfunction<sup>(1)</sup>. The host response results in a systemic inflammatory syndrome that manifests as hyper/hypothermia, tachycardia, tachypnea and alterations in the leucocyte count<sup>(2)</sup>. When this response becomes pathological, it leads to micro-circulatory derangements<sup>(3)</sup>, hypotension, cellular hypoxia/dysoxia<sup>(4)</sup> and ultimately death. It is estimated that 1.7 million adults were hospitalised in the USA in 2014 with sepsis<sup>(5)</sup>, which results in a major economic burden given that it is the most expensive condition to treat in US hospitals<sup>(6)</sup>. Sepsis is the leading cause of mortality in hospitalised patients, contributing to 1 in every 2–3 deaths<sup>(7)</sup>. Despite its invasive nature and high mortality rates, every targeted, large pharmaceutical trial has failed<sup>(8)</sup>.

Sepsis results in a high energy expenditure due to its elevated metabolic demand as well as inefficiencies in normal biochemical processes<sup>(9)</sup>. The hypermetabolic state induced by the host response to the infection as well as the need to replenish damaged cells contributes to micronutrient deficiencies. The resulting deficiencies lead to alterations in normal energy homeostasis and inefficiencies in energy production. The standard of care to treat sepsis is early intravenous fluids, prompt broad-spectrum antibiotics, source control of the infecting agent, low tidal volume ventilation and

haemodynamic optimisation to maintain an adequate blood pressure to perfuse end-organs<sup>(10)</sup>. However, a successful adjunctive treatment to curtail the inflammatory response, diminish free-radical damage and/or improve metabolic derangements would result in a much-needed weapon in the fight against sepsis. Micronutrient therapy has the potential to aid in such processes making it an interesting subject to study. We present the current state of knowledge on micronutrients in sepsis as it pertains to thiamine, L-carnitine, vitamin C, Se and vitamin D, as these have been extensively studied in the field of sepsis (Table 1).

### Thiamine

Thiamine serves as a cofactor for multiple cellular enzymes that are essential in aerobic carbohydrate metabolism, maintenance of cellular redox status, mitochondrial oxidative phosphorylation and synthesis of adenosine triphosphate<sup>(11)</sup>. Despite its importance, the body does not produce thiamine and can only store up to 30 mg at a time in tissues such as the skeletal muscle, heart, kidney and brain<sup>(12)</sup>. Due to its quick turnover, thiamine deficiency can develop within 2 weeks without adequate supplementation. Many well-described syndromes are known to result from deficiencies of thiamine, including cardiac beriberi and Wernicke's encephalopathy<sup>(13,14)</sup>. Thiamine is found in raw

**Abbreviations:** ICU, intensive care unit; LPS, lipopolysaccharide.

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**Table 1.** Summary table of micronutrient literature in sepsis

	Thiamine	L-Carnitine	Se	Vitamin C	Vitamin D
Mechanism of action in sepsis	Needed to convert pyruvate to acetyl-coA, allowing entry into the citric acid cycle and aerobic metabolism. Antioxidant <sup>(20)</sup>	Energy production: involved in the rate-limiting step of $\beta$ -oxidation to transport long-chain fatty acids from the cytoplasm to mitochondria	Incorporated into glycoproteins for antioxidant effects <sup>(43,45–47)</sup>	Important in biosynthetic and metabolic processes, potent antioxidant, improves vasopressor synthesis <sup>(56–60,72)</sup>	Regulation of immune system's ability to modulate systemic inflammatory response <sup>(96,97)</sup>
Levels in sepsis	Deficient in 20–71 % of patients with septic shock. Unclear if levels correlate with mortality <sup>(16–18,22)</sup>	Lack of quality human studies	Non-survivors had lower admission levels and declined more rapidly <i>v.</i> survivors <sup>(48)</sup>	Levels decreased in patients with sepsis <sup>(19,86)</sup>	Deficiency associated with higher sepsis severity in patients hospitalised for infection <sup>(105,106)</sup>
Animal studies	Improved pH, mean arterial pressure and cardiac index in septic dogs <sup>(25)</sup>	Mortality benefit in pre-treated rats exposed to lipopolysaccharide <sup>(36)</sup>	Decreased myeloperoxidase activity and lung inflammation <sup>(52)</sup>	Reverses microcirculatory derangements <sup>(82)</sup> , attenuates sepsis-induced organ injury <sup>(83,84)</sup>	Reduces coagulation derangements, alveolar inflammation, cellular damage and hypoxia in septic mice <sup>(104,108)</sup>
Human studies	Improved survival in subgroup of patients with deficiency <sup>(26)</sup> . Decreased mortality in combination with vitamin C and hydrocortisone <sup>(29)</sup>	Improves early haemodynamic parameters <sup>(37)</sup> and mortality in patients with septic shock <sup>(38)</sup>	Mixed results with most recent meta-analysis showing no effect on mortality <sup>(53–55)</sup>	Decreased mortality in combination with vitamin C and hydrocortisone <sup>(29)</sup> . Decreased vasopressor and mortality <sup>(89)</sup>	No effect on mortality; larger trials are needed <sup>(110–112)</sup>
Recommended dosages from clinical trials	200 mg intravenous bolus every 12 h for 4 d or until ICU discharge <sup>(29)</sup>	4 g bolus injection over 2–3 min followed by an 8 g infusion over the following 12 h <sup>(38)</sup>	1000 $\mu$ g of sodium selenite bolus followed by a 14 d infusion of 1000 $\mu$ g <sup>(53)</sup>	1.5 g intravenous infusion over 30 min every 6 h for 4 d or until ICU discharge <sup>(29)</sup>	A single dosage of calcitriol 2 $\mu$ g intravenously <sup>(110)</sup>
Recommendation	Consider in those at risk for deficiency	Consider in those with septic shock	Not currently supported by trials	Consider in those with septic shock	Not currently supported by trials

ICU, intensive care unit.

foods including green vegetables and nuts as well as certain processed foods such as fortified cereals<sup>(15)</sup>.

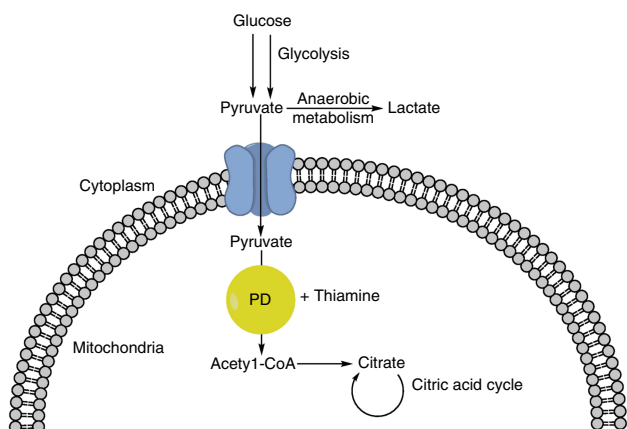
Thiamine deficiency is commonly found among septic patients, with a range in prevalence between 20 and 71%, depending on measurement techniques and inclusion criteria<sup>(16–18)</sup>. Donnino *et al.*<sup>(16)</sup> found that 20% of septic patients exhibited thiamine deficiency, where sepsis was defined as suspected infection and evidence of tissue hypoperfusion (lactate >4 mmol/l or hypotension requiring vasopressor support); an absolute thiamine deficiency was noted at a level <9 nmol/l. In another study, Costa *et al.*<sup>(17)</sup> showed that the incidence of thiamine deficiency was 71.3% in septic shock patients, defined as hypotension requiring vasopressor support. A serum thiamine below 16 ng/ml was set as the threshold below which thiamine deficiency was defined. Lastly, Dizdar *et al.*<sup>(18)</sup> found that 56% of septic patients exhibited thiamine deficiency, with an average serum concentration of 28.3 ng/ml (normal range: 33–99 ng/ml). In this work, sepsis was again defined as an infection-induced inflammatory response resulting in hypotension requiring vasopressor support. Although these studies were mostly small, uni-centre undertakings, they demonstrate that, despite large variability, thiamine deficiency is prevalent in the sepsis population.

There are multiple possible mechanisms to explain the association between thiamine deficiency and sepsis, though whether the deficiency is a cause or consequence of sepsis is not evident. Decreased appetite, diarrhoea, impaired absorption, and/or increased metabolic demand could deplete thiamine reserves and contribute to thiamine deficiency during sepsis. By decreasing the activity of the pyruvate dehydrogenase complex (needed to convert pyruvate to acetyl-coA in order to enter the citric acid cycle), thiamine deficiency can increase lactic acid production by favouring the anaerobic metabolism of pyruvate and shunting it away from the citric acid cycle (Fig. 1). Further, the profound oxidative stress that is a hallmark of sepsis can result in consumption of endogenous antioxidants<sup>(19)</sup>. Thiamine, with its role in preventing lipid peroxidation and oleic acid oxidation, is one such antioxidant<sup>(20)</sup>. Animal studies may also provide clues regarding

this association. In an experimental model of sepsis, thiamine deficiency was associated with oxidative stress and inflammatory changes<sup>(21)</sup>. In this paper, mice in which sepsis was induced via caecal ligation and puncture were fed a diet deficient in thiamine; these mice displayed increases in inflammatory and oxidative stress markers compared with controls. This suggests that thiamine deficiency can compound the body's inherent stress response to sepsis.

Despite the prevalence of thiamine deficiency in sepsis, studies have found conflicting relationships between thiamine levels and clinical outcomes. One study found that those with lower levels of thiamine had an increased mortality rate<sup>(22)</sup> while others failed to find any such link<sup>(18)</sup>. A potential drawback of the first study was that it was retrospective, conducted only in intensive care unit (ICU) patients requiring parenteral nutrition support, and used an indirect measurement of thiamine activity. In a prospective observational study of 125 patients admitted to the ICU, Corcoran *et al.*<sup>(23)</sup> examined the association of vitamin deficiency with mortality and found no significant association. A potential drawback of this study was that only nine of the 125 patients had a measurable thiamine deficiency. Similarly, Dizdar *et al.*<sup>(18)</sup> found that while thiamine levels were reduced in patients who died, the levels were not independently associated with mortality. Therefore, more studies with larger sample sizes are needed to draw a definite conclusion.

Due to the prevalence of thiamine deficiency and its possible effect on clinical outcomes in septic patients, studies have been undertaken to determine the therapeutic relevance of thiamine administration. Using cultured rat cardiomyocytes, Shin *et al.*<sup>(24)</sup> demonstrated that thiamine can be cytoprotective in cells undergoing hypoxic stress by preventing apoptosis. An early animal study corroborated the idea that thiamine administration was beneficial in septic shock. Thiamine pyrophosphate administration improved pH, mean arterial pressure and cardiac index in a dog model of endotoxic shock<sup>(25)</sup>. More recent clinical research has shown mixed results. In the largest human randomised, double-blind clinical study investigating the therapeutic potential of thiamine to date, Donnino *et al.*<sup>(26)</sup> found that administration of thiamine did not improve lactate levels, mortality or ICU length of stay in patients with septic shock and elevated lactate. In this study, a plasma thiamine level  $\leq 7$  nmol/l was considered deficient and a dose of 200 mg of thiamine intravenously was administered twice daily for 7 d or until hospital discharge. However, when baseline thiamine deficiency was taken into account, patients had a significantly lower lactate level at 24 h and longer survival rates. A secondary analysis of the study indicated that those randomised to the thiamine group had lower creatinine levels and a lower rate of progression to renal replacement therapy<sup>(27)</sup>. This study suffers from some limitations, including the fact that the subgroup of thiamine-deficient patients was very small (fifteen in treatment, thirteen in control) and was no longer randomised. In support of Donnino's findings that thiamine-deficient septic patients could benefit from thiamine supplementation was a retrospective study in which patients with alcohol use disorder (known to be associated with lower levels of thiamine) who were given thiamine during their hospital stay had a decreased



**Fig. 1.** Glucose undergoes glycolysis, forming pyruvate. Pyruvate can then be converted to acetyl-CoA via pyruvate dehydrogenase (PD) and thiamine, entering the citric acid cycle. Alternatively, pyruvate can undergo anaerobic metabolism and form lactic acid.

mortality rate from septic shock<sup>(28)</sup>. Additionally, Marik *et al.*<sup>(29)</sup> showed that the combination of hydrocortisone (50 mg every 6 h for 4 d or until ICU discharge), vitamin C (1.5 mg every 6 h for 4 d or until ICU discharge) and thiamine (200 mg every 12 h for 4 d or ICU discharge) had a significant effect on patients with sepsis and septic shock. Patients receiving the intervention had a marked reduction in mortality (8.5% (four out of forty-seven) compared with 40% (nineteen out of forty-seven) of controls), where all four of the patients in the treatment group were deemed to have died from non-sepsis-related causes. Further, there was a significant decrease in time to weaning of vasopressor support in the treatment group compared with controls. The authors purport that thiamine was added since it is a co-enzyme for vitamin C metabolism and that the encouraging results can be attributed to the synergistic effects of hydrocortisone and vitamin C.

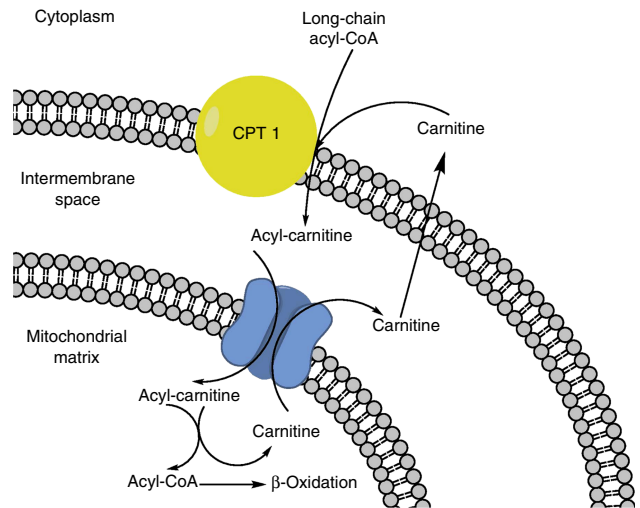
The above studies together indicate that the administration of thiamine, probably in conjunction with other micronutrients, may be beneficial in the septic patient with risk factors for thiamine deficiency, as intravenous thiamine is cheap and has relatively few side effects. Such risk factors for thiamine deficiency include malnutrition, alcoholism, chronic wasting diseases, renal replacement therapy, hyperemesis gravidarum, anorexia nervosa, gastric bypass surgery and refeeding<sup>(30)</sup>. Due to the small sample sizes and drawbacks of the aforementioned studies, we instead feel that the clinician may consider thiamine supplementation in the septic shock patient with risk factors for thiamine deficiency. Future studies may benefit from elucidating whether altered thiamine utilisation and uptake by various tissues during sepsis may be a critical determinant of whether patients benefit from thiamine administration.

### L-Carnitine

Fatty acids and TAG are utilised as energy in a process known as  $\beta$ -oxidation. The rate-limiting step in this process requires an L-carnitine-mediated transport of long-chain fatty acids from the cytoplasm into the mitochondria (Fig. 2). Carnitine palmitoyl transferase 1 is the enzyme responsible for this transport and its activity is inhibited during sepsis<sup>(31)</sup>.

L-Carnitine is primarily obtained from food, with meat being the largest source; however, it can be endogenously synthesised from the amino acids lysine and methionine<sup>(32,33)</sup>. Systemic primary carnitine deficiency is characterised by episodes of metabolic crisis characterised by hypoketotic hypoglycaemia events, hyperammonaemia, hypertransaminasaemia, hepatomegaly and hepatic encephalopathy. In addition, skeletal muscle weakness, cardiomyopathy and elevated creatinine kinase levels can be seen<sup>(34,35)</sup>.

The first animal study investigating the link between L-carnitine and sepsis was published in 1989 and showed a mortality benefit in pre-treated L-carnitine rats exposed to a lipopolysaccharide (LPS) sepsis model<sup>(36)</sup>. Two human clinical trials have evaluated the use of L-carnitine in sepsis and both showed promising results. The first, published in 1991, demonstrated early improvements in haemodynamic parameters in patients with septic shock<sup>(37)</sup>. The second, a small phase 1 randomised



**Fig. 2.** Long-chain acyl-CoA is shuttled into the mitochondrial matrix for  $\beta$ -oxidation by combining with carnitine via carnitine palmitoyl transferase 1 (CPT 1).

control trial of thirty-one patients, analysed the safety and efficacy of L-carnitine infusion in patients undergoing vasopressor-dependent septic shock. This study showed a lower 28 d mortality rate of 25 *v.* 60% favouring L-carnitine with no differences in significant adverse events<sup>(38)</sup>. The dosage for L-carnitine in the later study was a 4 g bolus injection (20 ml) over 2–3 min followed by an 8 g infusion (8 g in 1000 ml of 0.9% normal saline) over the following 12 h (83 ml/h). A phase 2 trial with an estimated 250-patient enrollment is currently underway<sup>(39)</sup>.

In conclusion, preliminary results merit consideration for further research in this area and clinicians may consider L-carnitine supplementation for patients in septic shock.

### Selenium

Se is an essential nutrient required for life<sup>(40)</sup>. The recommended daily allowance is 55  $\mu$ g/d, with the median American ingesting 81  $\mu$ g/d. It is widely available and found in dairy products, meats, seafood, vegetables and grains<sup>(41)</sup>, with regional differences in the soil resulting in variations in consumption across geographical areas<sup>(42)</sup>. The majority of Se is incorporated into glycoproteins<sup>(43)</sup>, with the liver being the major source of secretion<sup>(44)</sup>. For example, the glycoprotein glutathione peroxidase requires Se at its active site to detoxify reactive oxygen species such as  $H_2O_2$  and phospholipid hydroperoxide<sup>(45,46)</sup>. Se is frequently referred to as an antioxidant due to its role in reversing the effects of oxidised lipids and methionine residues, and detoxifying hydrogen peroxidase<sup>(47)</sup>.

Sakr *et al.*<sup>(48)</sup> studied the time course of Se levels in sixty surgical ICU patients with four equal subgroups consisting of ICU-controls, systemic inflammatory response syndrome (SIRS), severe sepsis, and septic shock ICU patients. He found that 92% of patients had Se levels below the standard for healthy subjects (74  $\mu$ g/l) and that all but the ICU-control cohort had declining

levels of Se during their ICU stay. Se levels were lowest on admission and decreased more markedly in non-survivors. Lower Se levels were associated with greater tissue damage and the presence of infection and organ dysfunction. This decline in Se levels during sepsis is consistent with a mouse study in which selenoprotein P declined after administration of LPS<sup>(49)</sup>. A subsequent study found that ICU patients with low admission levels of Se were at increased risk of nosocomial pneumonia, organ system failure and mortality<sup>(50)</sup>. During times of Se deficiency, the thyroid and brain maintain adequate levels at the expense of the immune system, which undergoes a rapid decline in availability<sup>(51)</sup>. This is problematic, given the decline in Se levels seen in patients with sepsis.

Se has been studied in mouse and *in vitro* models of sepsis. Se-treated mice had decreased myeloperoxidase activity (a marker of neutrophil accumulation) and decreased lung inflammation<sup>(52)</sup>. Human trials have been met with conflicting results. A multi-centred, randomised control trial involving eleven ICU in Germany in which severe sepsis or septic shock patients were randomised to either 1000 µg of sodium selenite as a 30-min bolus followed by a 14 d continuous infusion of 1000 µg *v. placebo* showed a mortality benefit in those receiving sodium selenite<sup>(53)</sup>. However, a more recent multi-centred randomised controlled trial involving thirty-three ICU in Germany failed to show an effect of high-dosage sodium selenite on the 28 d mortality rate of patients with severe sepsis or septic shock<sup>(54)</sup>. Both studies used the same initial bolus loading dose and daily continuous infusion. The most recent meta-analysis analysing twenty-one randomised control trials found no effect of Se on mortality<sup>(55)</sup>.

Although lower levels of Se are seen in patients with sepsis and are associated with worsening outcomes, human trials have shown mixed results. It is thus not recommended at the current time that Se be given to those patients with sepsis.

## Vitamin C

Vitamin C is water-soluble and involved in numerous biosynthetic and metabolic processes. It is integral for collagen<sup>(56)</sup>, carnitine<sup>(57)</sup> and neurotransmitter biosynthesis<sup>(58)</sup>, serves as a potent antioxidant scavenging against excess reactive oxygen species<sup>(59)</sup>, restores other cellular antioxidants, acts as an immunomodulatory agent<sup>(60)</sup>, and is an essential cofactor for Fe-containing enzymes<sup>(61)</sup>. Most plants and animals synthesise ascorbic acid endogenously; however, mammalian species lack the enzyme gulonolactone oxidase, thus requiring exogenous supplementation via dietary intake and supplements<sup>(62)</sup>. At physiological pH, ascorbate is bioavailable equally as either dehydro-L-ascorbic acid (DHA) or L-ascorbic acid (AsCA). Specialised cells can take up reduced vitamin C (AsCA) through Na-dependent ascorbate co-transporters (SVCT1 and SVCT2)<sup>(63)</sup>. Most other cells take up vitamin C in its oxidised form (DHA) via facilitative GLUT<sup>(64)</sup>.

In septic hosts, the immunomodulation and antioxidant activity of vitamin C complements other host responses in the multifaceted inflammatory processes, leading to sepsis-mediated multiple organ dysfunction. Vitamin C is involved in

attenuating the proinflammatory and procoagulant state which induces vascular-ischaemic-induced organ injury<sup>(65,66)</sup>. It contributes towards the recovery of endothelial dysfunction by preventing formation of reactive oxygen species and via reductive recycling which uncouples endothelial NO synthase<sup>(67)</sup>. Additionally, laboratory research suggests that vitamin C reduces platelet aggregation of surface P-selectin expression<sup>(68)</sup>, attenuates hypothalamic neuronal damage<sup>(69)</sup>, may prevent cellular immunosuppression<sup>(70)</sup>, impedes phagocyte adhesion to endothelial cells preventing phagocyte oxidative damage<sup>(71)</sup> and improves endogenous vasopressor synthesis<sup>(72)</sup>.

In critically ill patients, several investigations have demonstrated low circulating levels of vitamin C, particularly in sepsis<sup>(71)</sup>. Ascorbate may become oxidised to ascorbyl free radical and dehydroascorbic acid, thus making ascorbate concentrations subnormal in plasma and leucocytes while plasma ascorbyl free radical concentration is elevated<sup>(73)</sup>. Deficiency may have serious consequences and be further compounded by diminished endothelial cell uptake due to inflammatory cascade activation given that inflammatory cytokines (TNF- $\alpha$ , IL-1 $\beta$ ) inhibit ascorbate uptake via down-regulation of ascorbate-specific transporters<sup>(74)</sup>. Plasma concentrations may become low within 24 h of onset of illness, and concentrations of <10 µmol/l are described in critically ill patients<sup>(75,76)</sup>. Low plasma concentrations are associated with inflammation, severity of organ failure, and mortality<sup>(19,75,77–79)</sup>.

Animal studies have supported a scientific basis for ascorbate having a beneficial and therapeutic effect in the host response to infection. Pre-existing ascorbate deficiency may decrease survival in a mouse model injected with pathogenic bacteria as animals were three times more likely to succumb to infection with introduction of *Klebsiella pneumoniae*<sup>(80)</sup>. In mouse influenza models, ascorbate deficiency has been shown to worsen lung pathology following induction of infection<sup>(81)</sup>. Additionally, animal sepsis models have demonstrated that ascorbate infusions protect against impairment of microvascular blood flow<sup>(82)</sup>, attenuate sepsis-induced organ injury<sup>(83,84)</sup> and decrease the escalation of severity of disease in combination with antimicrobials<sup>(85)</sup>.

While very preliminary and exploratory, human observational studies have revealed that ascorbate levels are significantly lower in septic patients and those with other aetiologies of critical illness such as severe trauma and acute respiratory distress syndrome<sup>(19,75,77,78,86)</sup>. Low plasma concentrations correlate inversely with the incidence of organ failure and markers of inflammation<sup>(78)</sup>. For dosing considerations, such low serum levels are not corrected by parenteral nutrition containing the daily recommended dose of ascorbate (200 mg/d)<sup>(75,76)</sup>. This is perhaps due to the accelerated destruction of ascorbate or a depletion of total body stores during the inflammatory cascade. High doses (3 g/d) given intravenously for 3 d or more appear necessary in order to achieve normal baseline serum levels<sup>(87)</sup>.

Preliminary-phase safety studies of high-dose ascorbate have not demonstrated any untoward pro-oxidative effects in healthy volunteers<sup>(88)</sup>. Case reports, early-phase clinical trials and retrospective human studies in septic patients have suggested that intravenous ascorbic acid infusions (either independently

or in combination with thiamine and hydrocortisone) are safe, well tolerated, may reduce multi-organ failure and biomarkers of inflammation, may reduce vasopressor requirements in septic shock, and may reduce mortality in patients with severe sepsis in septic shock<sup>(29,89–91)</sup>. However, these studies are very preliminary and limited by design and small sample size. Vitamin C may prove to be an important therapeutic option and can be considered for patients in septic shock; however, further studies are needed before it should be deemed standard of care.

### Vitamin D

Vitamin D is well known for its impact on bone health through regulation of Ca–phosphate homeostasis. However, it is often forgotten that the prototypical disease of vitamin D deficiency, nutritional rickets, was originally postulated to be infectious in origin, so great was its association with pneumonia and tuberculosis<sup>(92)</sup>. As the biochemical relationship between vitamin D and UV exposure was elucidated in the early decades of the 20th century<sup>(93)</sup>, rachitic children with infections of the respiratory system were often treated with cod oil and sun exposure. Interest in the role of vitamin D in the human response to infection was reinvigorated in the 1980s as a greater understanding of its role in the innate and adaptive immune systems emerged.

The hormonally active form of vitamin D is calcitriol (1,25-dihydroxyvitamin D<sub>3</sub>), a metabolite of vitamin D<sub>3</sub> derived from two sequential reactions. The primary sources of vitamin D<sub>3</sub> are: (1) the solar UVB conversion of 7-dehydrocholesterol present in the skin; and (2) vitamin D<sub>3</sub> directly absorbed from the gastrointestinal tract. Vitamin D<sub>3</sub> is converted to 25-hydroxyvitamin D by vitamin D-25-hydroxylase in the liver and finally to calcitriol in the kidney and other tissues. Once converted, calcitriol binds to the vitamin D receptor (VDR), which is expressed in a wide range of cells and plays a crucial role in the optimal functioning of many organ systems and their physiological response to illness. Highlighting the importance of vitamin D status in health and response to illness, the 2011 Institute of Medicine and the Endocrine Society established minimal concentrations of >20–30 ng/dl to optimise health benefits<sup>(94)</sup>. It is estimated that 1 billion individuals are vitamin D deficient, including 40–100% of the US geriatric population<sup>(95)</sup>.

The discovery of VDR in activated CD4<sup>+</sup>, CD8<sup>+</sup> T cells, B cells, neutrophils, macrophages and dendritic cells highlights the integral role of vitamin D in the regulation of the innate and adaptive immune system's ability to modulate the systemic inflammatory response syndrome<sup>(96,97)</sup>. Cathelicidin-related antimicrobial peptides (LL-37, hCAP-18) and toll-like receptors (TLR) are key components of this system and play a significant role in antimicrobial activity. Adequate concentrations of circulating calcitriol have been demonstrated to play a crucial role in LL-37 production by macrophages in bronchial epithelial cells<sup>(98)</sup>. Similarly, co-stimulation of human macrophages by calcitriol and TLR ligands appear to up-regulate the expression of VDR and result in the induction of LL-37<sup>(99)</sup>. *In vitro* studies have also demonstrated the anti-inflammatory effects of vitamin D by inhibiting the CD4<sup>+</sup> Th1 cells production of

cytokines IL-2, interferon and TNF<sup>(100)</sup>. Vitamin D also enhances IL-4, IL-5 and IL-10 production of Th2 cells, in theory mitigating the deleterious effects of the pro-inflammatory state<sup>(101)</sup>. Human monocytes stimulated with LPS showed dose-dependent reductions in TNF- $\alpha$  and tissue factor, key inflammatory molecules in sepsis<sup>(102)</sup>.

Given the vital role that vitamin D appears to play in the innate immune response to infection, it is plausible that vitamin D levels are a determinant of risk for sepsis severity and clinical outcomes in septic shock. Indeed, vitamin D deficiency has been demonstrated in a high percentage of critically ill patients with sepsis and found to correlate with low levels of LL-37<sup>(103,104)</sup>. Patients presenting to an emergency department with suspected infection and a baseline 25-hydroxyvitamin D insufficiency were found to be more likely to have severe sepsis and a higher sepsis severity (sequential organ failure assessment (SOFA) > 2) than infected patients with normal vitamin D levels<sup>(105)</sup>. In a large study involving more than 3000 critically ill patients, vitamin D deficiency was a significant predictor of sepsis and carried a 1.6-fold increase in mortality<sup>(106)</sup>.

Given the association with vitamin D deficiency and risk for sepsis severity and mortality, vitamin D repletion is a promising target for therapeutic intervention. *In vitro* models of sepsis seem to support this strategy. In murine models, pretreatment of human endothelial cells with vitamin D reduced LPS-induced production of proinflammatory cytokines<sup>(107)</sup>. Similarly, sepsis-induced coagulation derangements were attenuated by the administration of 1,25-dihydroxyvitamin D<sub>3</sub> in one study and cholecalciferol treatment 6 h post-injury reduced alveolar inflammation, cellular damage and hypoxia in another, both involving a murine model of caecal ligation and puncture<sup>(104,108)</sup>. Clinical trials have demonstrated promise but appear to be inconclusive. Quraishi *et al.*<sup>(109)</sup> reported an association between high-dose cholecalciferol administration and increased 25-hydroxyvitamin D and LL-37 in a cohort of septic shock ICU patients. However, a double-blind randomised trial comparing calcitriol *v.* placebo in sixty-seven critically ill patients with severe sepsis or septic shock failed to demonstrate an increase in plasma cathelicidin or a clear impact on immunomodulatory markers with calcitriol administration<sup>(110)</sup>. While not a primary outcome, supplemental calcitriol did not have an effect on mortality, or ICU or hospital length of stay. Systematic reviews of interventional trials reflect this inconsistency in results<sup>(111,112)</sup>.

Research over the last decade has established the pluripotent regulatory role that vitamin D plays in the human response to infection<sup>(113)</sup>. *In vitro* studies demonstrate the promise that interventions aimed at optimising vitamin D homeostasis can attenuate the pathophysiological cascade in septic shock models. The data are robust regarding the high prevalence of vitamin D insufficiency in the critically ill population and its relationship to critical illness outcomes including sepsis. Vitamin D supplementation is generally regarded as safe. While the demonstration of clinical efficacy for vitamin D strategies in clinical practice has proved more challenging, future investigation is warranted and forthcoming. Optimising vitamin D levels would appear to have a role in primary infection prevention for high-risk critical care populations and as an

acute adjuvant therapy to mitigate the inflammatory and coagulation cascade associated with sepsis. Further clinical studies are required, and clinicians should not consider vitamin D therapy for sepsis standard of care until these trials are concluded.

## Conclusion

In this modern era, mortality rates of septic shock remain unacceptably high and micronutrient resuscitative strategies may complement and enhance traditional resuscitative strategies. Additionally, with the advent of 'Precision Health' the identification of strategies to identify patient populations with absolute or relative micronutrient deficiency or functional pathway impediment may yield insights which could halt sepsis cascade escalation in specific hosts. For investigators, there remains an abundance of opportunities for further investigation at the molecular and biochemical levels and in human observational (i.e. biomarker) and clinical trials. If the current body of evidence is validated by higher-phase human clinical trials, sepsis micronutrient resuscitative strategies could revolutionise the current standard of care. Future trials should focus on early treatment, as inflammatory biomarkers are elevated at the most proximal part of hospital presentation<sup>(114)</sup>. Based on all the current literature, clinicians may consider the use of thiamine in all septic patients at risk for thiamine deficiency and L-carnitine and vitamin C to those in septic shock. Future ongoing studies will lead to a clearer picture on the use of micronutrients in sepsis.

## Acknowledgements

The present review received no specific grant from any funding agency, commercial or not-for-profit sectors.

J. B. B. conceived of the paper; all authors were involved in the final editing of the manuscript. Each section had a main author who wrote the first draft: J. B. B., abstract, introduction, L-carnitine, Se; C. R. W., vitamin C, conclusion; V. J., thiamine; J. E. S., vitamin D.

There are no conflicts of interest.

## References

1. Singer M, Deutschman CS, Seymour CW, *et al.* (2016) The Third International Consensus Definitions for Sepsis and Septic Shock (Sepsis-3). *JAMA* **315**, 801–810.
2. Bone RC, Balk RA, Cerra FB, *et al.* (1992) American College of Chest Physicians/Society of Critical Care Medicine Consensus Conference: definitions for sepsis and organ failure and guidelines for the use of innovative therapies in sepsis. *Crit Care Med* **20**, 864–874.
3. Trzeciak S, Dellinger RP, Parrillo JE, *et al.* (2007) Early microcirculatory perfusion derangements in patients with severe sepsis and septic shock; relationship to hemodynamics, oxygen transport, and survival. *Ann Emerg Med* **49**, 88–98.
4. Loiacono L & Shapiro D (2010) Detection of hypoxia at the cellular level. *Crit Care Clin* **26**, 409–421.
5. Rhee C, Dantes R, Epstein L, *et al.* (2017) Incidence and trends of sepsis in US hospitals using clinical vs claims data, 2009–2014. *JAMA* **318**, 1241–1249.
6. Torio C & Moore B (2016) National inpatient hospital costs: the most expensive conditions by payer, 2013. HCUP statistical brief #204. <https://hcup-us.ahrq.gov/reports/statbriefs/sb204-Most-Expensive-Hospital-Conditions.jsp> (accessed September 2017).
7. Liu V, Escobar GJ, Greene JD, *et al.* (2014) Hospital deaths in patients with sepsis from 2 independent cohorts. *JAMA* **312**, 90–92.
8. Marshall J (2014) Why have clinical trials in sepsis failed? *Trends Mol Med* **20**, 195–203.
9. L'Her E & Sebert P (2001) A global approach to energy metabolism in an experimental model of sepsis. *Am J Respir Crit Care Med* **164**, 1444–1447.
10. Rhodes A, Evans LE, Alhazzani W, *et al.* (2017) Surviving sepsis campaign: international guidelines for the management of sepsis and septic shock: 2016. *Crit Care Med* **45**, 486–552.
11. Manzanares W & Hardy G (2011) Thiamine supplementation in the critically ill. *Curr Opin Clin Nutr Metab Care* **14**, 610–617.
12. Ariaey-Nejad MR, Balaghi M, Baker EM, *et al.* (1970) Thiamin metabolism in man. *Am J Clin Nutr* **23**, 764–778.
13. DiNicolantonio J, Niaz A, Lavie C, *et al.* (2013) Thiamine supplementation for the treatment of heart failure: a review of the literature. *Congest Heart Fail* **19**, 214–222.
14. Krill J (1996) Neuropathology of thiamine deficiency disorders. *Metab Brain Dis* **11**, 9–17.
15. Depient F, Bruce WR, Shangari N, *et al.* (2006) Mitochondrial function and toxicity: role of the B vitamin family on mitochondrial energy metabolism. *Chem Biol Interact* **163**, 94–112.
16. Donnino MW, Carney E, Cocchi MN, *et al.* (2010) Thiamine deficiency in critically ill patients with sepsis. *J Crit Care* **25**, 576–581.
17. Costa NA, Gut AL, de Souza Dorna M, *et al.* (2014) Serum thiamine concentration and oxidative stress as predictors of mortality in patients with septic shock. *J Crit Care* **29**, 249–252.
18. Dizdar OS, Baspinar O, Kocer D, *et al.* (2016) Nutritional risk, micronutrient status and clinical outcomes: a prospective observational study in an infectious disease clinic. *Nutrients* **8**, 124.
19. Goode HF, Cowley HC, Walker BE, *et al.* (1995) Decreased antioxidant status and increased lipid peroxidation in patients with septic shock and secondary organ dysfunction. *Crit Care Med* **23**, 646–651.
20. Lukienko PI, Mel'nichenko NG, Zverinskii IV, *et al.* (2000) Antioxidant properties of thiamine. *Bull Exp Biol Med* **130**, 874–876.
21. De Andrade JAA, Gayer CRM, Nogueira NPA, *et al.* (2014) The effect of thiamine deficiency on inflammation, oxidative stress and cellular migration in an experimental model of sepsis. *J Inflamm* **11**, 11.
22. Cruickshank AM, Telfer AB & Shenkin A (1988) Thiamine deficiency in the critically ill. *Intensive Care Med* **14**, 384–387.
23. Corcoran TB, O'Neill MA, Webb SA, *et al.* (2009) Prevalence of vitamin deficiencies on admission: relationship to hospital mortality in critically ill patients. *Anaesth Intensive Care* **37**, 254–260.
24. Shin BH, Choi SH, Cho EY, *et al.* (2004) Thiamine attenuates hypoxia-induced cell death in cultured neonatal rat cardiomyocytes. *Mol Cells* **18**, 133–140.

25. Lindenbaum GA, Larrieu AJ, Carroll SF, *et al.* (1989) Effect of cocarboxylase in dogs subjected to experimental septic shock. *Crit Care Med* **17**, 1036–1040.
26. Donnino MW, Andersen LW, Chase M, *et al.* (2016) Randomized, double-blind, placebo-controlled trial of thiamine as a metabolic resuscitator in septic shock: a pilot study. *Crit Care Med* **44**, 360–367.
27. Moskowitz A, Andersen LW, Cocchi MN, *et al.* (2017) Thiamine as a renal protective agent in septic shock: a secondary analysis of a randomized, double-blind, placebo-controlled trial. *Ann Am Thorac Soc* **14**, 737–741.
28. Holmberg M, Moskowitz A, Patel P, *et al.* (2018) Thiamine in septic shock patients with alcohol use disorders: an observational pilot study. *J Crit Care* **43**, 61–64.
29. Marik PE, Khangoora V, Rivera R, *et al.* (2016) Hydrocortisone, vitamin C and thiamine for the treatment of severe sepsis and septic shock: a retrospective before-after study. *Chest* **151**, 1229–1238.
30. Leite HP & de Lima LFP (2016) Metabolic resuscitation in sepsis: a necessary step beyond the hemodynamic? *J Thorac Dis* **8**, E552–E557.
31. Eaton S, Fukumoto L, Stefanutti G, *et al.* (2003) Myocardial carnitine palmitoyltransferase I as a target for oxidative modification in inflammation and sepsis. *Biochem Soc Trans* **31**, 1133–1136.
32. Tanphaichitr V & Broquist HP (1973) Role of lysine and  $\epsilon$ -N-trimethyllysine in carnitine biosynthesis. II. Studies in the rat. *J Biol Chem* **248**, 2176–2181.
33. Tanphaichitr V, Horne DW & Broquist HP (1971) Lysine, a precursor of carnitine in the rat. *J Biol Chem* **246**, 6364–6366.
34. Longo N, di San Filippo CA & Pasquali M (2006) Disorders of carnitine transport and the carnitine cycle. *Am J Med Genet C Semin Med Genet* **142C**, 77–85.
35. Roe CR & Ding J (2001) Mitochondrial fatty acid oxidation disorders. In *The Metabolic Bases of Inherited Disease*, 8th ed., pp. 2297–2326 [CR Scriver, AL Beaudet, WS Sly and D Valle, editors]. New York: McGraw-Hill.
36. Takeyama N, Takagi D, Matsuo N, *et al.* (1989) Effect of L-carnitine on survival. *Am J Physiol* **256**, E31–E38.
37. Gasparetto A, Corbucci GG, de Blasi RA, *et al.* (1991) Influence of acetyl-L-carnitine infusion on haemodynamic parameters and survival of circulatory-shock patients. *Int J Clin Pharm Res* **11**, 83–92.
38. Puskarich M, Kline J, Krabill V, *et al.* (2014) Preliminary safety and efficacy of L-carnitine infusion for the treatment of vasopressor-dependent septic shock: a randomized control trial. *JPEN J Parenter Enteral Nutr* **38**, 736–743.
39. Lewis R, Viele K, Broglio K, *et al.* (2013) An adaptive, phase II, dose-finding clinical trial design to evaluate L-carnitine in the treatment of septic shock based on efficacy and predictive probability of subsequent phase III success. *Crit Care Med* **41**, 1674–1678.
40. Bösl MR, Takaku K, Oshima M, *et al.* (1997) Early embryonic lethality caused by targeted disruption of the mouse selenocysteine tRNA gene (*Trsp*). *Proc Natl Acad Sci U S A* **94**, 5531–5534.
41. Institute of Medicine (US) Panel on Dietary Antioxidants and Related Compounds (2000) *Dietary Reference Intakes for Vitamin C, Vitamin E, Selenium, and Carotenoids*, Section 7, Selenium. Washington, DC: National Academies Press. <https://www.ncbi.nlm.nih.gov/books/NBK225470/> (accessed June 2018).
42. Thompson JN, Erdody P & Smith DC (1975) Selenium content of food consumed by Canadians. *J Nutr* **10**, 274–277.
43. Burk RF & Hill KE (2005) Selenoprotein P: an extracellular protein with unique physical characteristics and a role in selenium homeostasis. *Annu Rev Nutr* **25**, 215–235.
44. Hill KE, Zhou J, McMahan WJ, *et al.* (2003) Deletion of selenoprotein P alters distribution of selenium in the mouse. *J Biol Chem* **278**, 13640–13646.
45. Epp O, Ladenstein R & Wendel A (1983) The refined structure of the selenoenzyme glutathione peroxidase at 0.2-nm resolution. *Eur J Biochem* **133**, 51–69.
46. Lei XG, Cheng WH & McClung JP (2007) Metabolic regulation and function of glutathione peroxidase-1. *Annu Rev Nutr* **27**, 41–61.
47. Huang Z, Rose AH & Hoffmann PR (2012) The role of selenium in inflammation and immunity: from molecular mechanisms to therapeutic opportunities. *Antioxid Redox Signal* **16**, 705–743.
48. Sakr Y, Reinhart K, Bloos F, *et al.* (2007) Time course and relationship between plasma selenium concentrations, systemic inflammatory response, sepsis, and multiorgan failure. *Br J Anaesth* **98**, 775–784.
49. Renko K, Hofmann PJ, Stoedter M, *et al.* (2009) Down-regulation of the hepatic selenoprotein biosynthesis machinery impairs selenium metabolism during the acute phase response in mice. *FASEB J* **23**, 1758–1765.
50. Forceville X, Vitoux D, Gauzit R, *et al.* (1998) Selenium, systemic immune response syndrome, sepsis, and outcome in critically ill patients. *Crit Care Med* **26**, 1536–1544.
51. Schomburg L & Schweizer U (2009) Hierarchical regulation of selenoprotein expression and sex-specific effects of selenium. *Biochim Biophys Acta* **1790**, 1453–1462.
52. Zolali E, Hamishehkar H, Maleki-Dizaji N, *et al.* (2014) Selenium effect on oxidative stress factors in septic rats. *Adv Pharm Bull* **4**, 289–293.
53. Angstwurm MW, Engelmann L, Zimmermann T, *et al.* (2007) Selenium in Intensive Care (SIC): results of a prospective randomized, placebo-controlled, multiple-center study in patients with severe systemic inflammatory response syndrome, sepsis, and septic shock. *Crit Care Med* **35**, 118–126.
54. Bloos F, Trips E, Nierhaus A, *et al.* (2016) Effect of sodium selenite administration and procalcitonin-guided therapy on mortality in patients with severe sepsis or septic shock: a randomized clinical trial. *JAMA Intern Med* **176**, 1266–1276.
55. Manzanares W, Lemieux M, Elke G, *et al.* (2016) High-dose intravenous selenium does not improve clinical outcomes in the critically ill: a systematic review and meta-analysis. *Crit Care* **20**, 356.
56. Boyera N, Galey I & Bernard BA (1998) Effect of vitamin C and its derivatives on collagen synthesis and cross-linking by normal human fibroblasts. *Int J Cosmet Sci* **20**, 151–158.
57. Hulse JD, Ellis S & Henderson LM (1978) Carnitine biosynthesis:  $\beta$  hydroxylation of trimethyllysine by an  $\alpha$ -keto glutarate dependent mitochondrial dioxygenase. *J Biol Chem* **253**, 1654–1659.
58. Harrison F & May J (2009) Vitamin C function in the brain: vital role of the ascorbate transporter (SVCT2). *Free Radic Biol Med* **46**, 719–730.
59. Mandl J, Szarka A & Banhegyi G (2009) Vitamin C: update on physiology and pharmacology. *Br J Pharmacol* **157**, 1097–1110.
60. Carr A & Maggini S (2017) Vitamin C and immune function. *Nutrients* **9**, E1211.
61. Lane D & Richardson D (2014) The active role of vitamin C in mammalian iron metabolism: much more than just enhanced iron absorption! *Free Radic Biol Med* **75**, 69–83.



62. Naidu KA (2003) Vitamin C in human health and disease is still a mystery? An overview. *Nutr J* **2**, 7.
63. Tsukaguchi H, Tokui T, Mackenzie B, *et al.* (1999) A family of mammalian Na<sup>+</sup>-dependent l-ascorbic acid transporters. *Nature* **399**, 70–75.
64. Vera J, Rivas C, Velásquez F, *et al.* (1995) Resolution of the facilitated transport of dehydroascorbic acid from its intracellular accumulation as ascorbic acid. *J Biol Chem* **270**, 23706–23712.
65. Fisher BJ, Seropian IM, Kraskauskas D, *et al.* (2011) Ascorbic acid attenuates lipopolysaccharide-induced acute lung injury. *Crit Care Med* **39**, 1454–1460.
66. Maeda N, Hagihara H, Nakata Y, *et al.* (2000) Aortic wall damage in mice unable to synthesize ascorbic acid. *Proc Natl Acad Sci U S A* **97**, 841–846.
67. Rodemeister S & Biesalski HK (2014) There's life in the old dog yet: vitamin C as a therapeutic option in endothelial dysfunction. *Crit Care* **18**, 461–462.
68. Secor D, Swarbreck S, Ellis CG, *et al.* (2013) Ascorbate reduces mouse platelet aggregation and surface P-selectin expression in an *ex vivo* model of sepsis. *Microcirculation* **20**, 502–510.
69. Chang CY, Chen JY, Chen SH, *et al.* (2016) Therapeutic treatment with ascorbate rescues mice from heat stroke-induced death by attenuating systemic inflammatory response and hypothalamic neuronal damage. *Free Radic Biol Med* **93**, 84–93.
70. Gao YL, Lu B, Zhai JH, *et al.* (2017) The parenteral vitamin C improves sepsis and sepsis-induced multiple organ dysfunction syndrome via preventing cellular immunosuppression. *Mediators Inflamm* **2017**, 4024672.
71. Koekkoek W & van Zanten A (2016) Antioxidant vitamins and trace elements in critical illness. *Nutr Clin Pract* **31**, 457–474.
72. Carr AC, Shaw GM, Fowler AA, *et al.* (2015) Ascorbate-dependent vasopressor synthesis: a rationale for vitamin C administration in severe sepsis and septic shock? *Crit Care* **19**, 418–425.
73. Wilson J (2009) Mechanism of action of vitamin C in sepsis: ascorbate modulates redox signaling in endothelium. *Biofactors* **35**, 5–13.
74. Seno T, Inoue N, Matsui K, *et al.* (2004) Functional expression of sodium-dependent vitamin C transporter 2 in human endothelial cells. *J Vasc Res* **41**, 345–351.
75. Schorah CJ, Downing C, Piriipitsi A, *et al.* (1996) Total vitamin C, ascorbic acid, and dehydroascorbic acid concentrations in plasma of critically ill patients. *Am J Clin Nutr* **96**, 760–765.
76. Berger MM (2009) Vitamin C requirements in parenteral nutrition. *Gastroenterology* **137**, Suppl. 5, S70–S78.
77. Metnitz PG, Bartens C, Fischer M, *et al.* (1999) Antioxidant status in patients with acute respiratory distress syndrome. *Intensive Care Med* **25**, 180–185.
78. Borrelli E, Roux-Lombard P, Grau GE, *et al.* (1996) Plasma concentrations of cytokines, their soluble receptors, and antioxidant vitamins can predict the development of multiple organ failure in patients at risk. *Crit Care Med* **24**, 392–397.
79. De Grooth HM, Spoelstra-de Man AME & Oudemans-van Straaten HM (2014) Early plasma vitamin C concentration, organ dysfunction and ICU mortality. *Intensive Care Med* **40**, Suppl. 1, S199.
80. Gaut JP, Belaaouaj A, Byun J, *et al.* (2006) Vitamin C fails to protect amino acids and lipids from oxidation during acute inflammation. *Free Radic Biol Med* **40**, 1494–1501.
81. Li W, Maeda N & Beck MA (2006) Vitamin C deficiency increases the lung pathology of influenza virus-infected *gulo*<sup>-/-</sup> mice. *J Nutr* **136**, 2611–2616.
82. Tynl K, Li F & Wilson JX (2005) Delayed ascorbate bolus protects against maldistribution of microvascular blood flow in a rat model of sepsis. *Crit Care Med* **33**, 1823–1828.
83. Fisher BJ, Kraskauskas D, Martin EJ, *et al.* (2012) Mechanisms of attenuation of abdominal sepsis induced acute lung injury by ascorbic acid. *Am J Physiol Lung Cell Mol Physiol* **303**, L20–L32.
84. Fisher BJ, Kraskauskas D, Martin EJ, *et al.* (2014) Attenuation of sepsis-induced organ injury in mice by vitamin C. *JPEN J Parenter Enteral Nutr* **38**, 825–839.
85. Leelahavanichkul A, Sompam P, Bootprapan T, *et al.* (2015) High-dose ascorbate with low-dose amphotericin B attenuates severity of disease in a model of the reappearance of candidemia during sepsis in the mouse. *Am J Physiol Regul Integr Comp Physiol* **309**, R223–R234.
86. Galley HF, Davies MJ & Webster NR (1996) Ascorbyl radical formation in patients with sepsis: effect of ascorbate loading. *Free Radic Biol Med* **20**, 139–143.
87. Long CL, Maull KI, Krishnan RS, *et al.* (2003) Ascorbic acid dynamics in the seriously ill and injured. *J Surg Res* **109**, 144–148.
88. Muhlhofer A, Mrosek S, Schlegel B, *et al.* (2004) High-dose intravenous vitamin C is not associated with an increase of pro-oxidative biomarkers. *Eur J Clin Nutr* **58**, 1151–1158.
89. Zabet MH, Mohammadi M, Ramezani M, *et al.* (2016) Effect of high-dose ascorbic acid on vasopressor requirement in septic shock. *J Res Pharm Pract* **5**, 94–100.
90. Fowler AA, Syed AA, Knowlson S, *et al.* (2014) Phase I safety trial of intravenous ascorbic acid in patients with severe sepsis. *J Transl Med* **12**, 32.
91. Bharara A, Grossman C, Grinnan D, *et al.* (2016) Intravenous vitamin C administered as adjunctive therapy for recurrent acute respiratory distress syndrome. *Case Rep Crit Care* **2016**, 8560871.
92. Chesney RW (2010) Vitamin D and The Magic Mountain: the anti-infectious role of the vitamin. *J Pediatr* **156**, 698–703.
93. Wolf G (2004) The discovery of vitamin D: the contribution of Adolf Windaus. *J Nutr* **134**, 1299–1302.
94. Kempker JA, Han JE, Tangpricha V, *et al.* (2012) Vitamin D and sepsis: an emerging relationship. *Dermatoendocrinology* **4**, 101–108.
95. Holick MF (2007) Vitamin D deficiency. *N Engl J Med* **357**, 266–281.
96. Baeke F, Takiishi T, Korf H, *et al.* (2010) Vitamin D: modulator of the immune system. *Curr Opin Pharmacol* **10**, 482–496.
97. Provedine DM, Tsoukas CD, Deftos LJ, *et al.* (1983) 1,25-Dihydroxyvitamin D<sub>3</sub> receptors in human leukocytes. *Science* **221**, 1181–1183.
98. Yim S, Dhawan P, Raganath C, *et al.* (2008) Induction of cathelicidin in normal and CF bronchial epithelial cells by 1,25-dihydroxyvitamin D<sub>3</sub>. *J Cyst Fibros* **6**, 403–410.
99. Liu PT, Stenger S, Li H, *et al.* (2006) Toll-like receptor triggering of a vitamin D-mediated human antimicrobial response. *Science* **311**, 1770–1773.
100. Lemire JM (1992) Immunomodulatory role of 1,25-dihydroxyvitamin D<sub>3</sub>. *J Cell Biochem* **49**, 26–31.
101. Boonstra A, Barrat FJ, Crain C, *et al.* (2001) 1 $\alpha$ ,25-Dihydroxyvitamin D<sub>3</sub> has a direct effect on naive CD4<sup>+</sup> T cells to enhance the development of Th2 cells. *J Immunol* **167**, 4974–4980.



102. Sadeghi K, Wessner B, Laggner U, *et al.* (2006) Vitamin D<sub>3</sub> down-regulates monocyte TLR expression and triggers hyporesponsiveness to pathogen-associated molecular patterns. *Eur J Immunol* **36**, 361–370.
103. Jeng L, Yamshchikov AV, Judd SE, *et al.* (2009) Alterations in vitamin D status and anti-microbial peptide levels in patients in the intensive care unit with sepsis. *J Transl Med* **7**, 28.
104. Parekh D, Patel JM, Scott A, *et al.* (2017) Vitamin D deficiency in human and murine sepsis. *Crit Care Med* **45**, 282–289.
105. Ginde AA, Camargo CA & Shapiro NI (2011) Vitamin D insufficiency and sepsis severity in emergency department patients with suspected infection. *Acad Emerg Med* **18**, 551–554.
106. Moromizato T, Litonjua AA, Braun AV, *et al.* (2014) Association of low serum 25-hydroxyvitamin D levels and sepsis in the critically ill. *Crit Care Med* **42**, 97–107.
107. Equils O, Naiki Y, Shapiro AM, *et al.* (2006) 1,25-Dihydroxyvitamin D inhibits lipopolysaccharide-induced immune activation in human endothelial cells. *Clin Exp Immunol* **143**, 58–64.
108. Moller S, Laigaard F, Olgaard K, *et al.* (2007) Effect of 1,25-dihydroxy-vitamin D<sub>3</sub> in experimental sepsis. *Int J Med Sci* **4**, 190–195.
109. Quraishi SA, De Pascale G, Needleman JS, *et al.* (2015) Effect of cholecalciferol supplementation on vitamin D status and cathelicidin levels in sepsis: a randomized, placebo-controlled trial. *Crit Care Med* **43**, 1928–1937.
110. Leaf DE, Raed D, Donnino MW, *et al.* (2014) Randomized controlled trial of calcitriol in severe sepsis. *Am J Respir Crit Care Med* **190**, 533–541.
111. Kearns MD, Alvarez JA, Seidel N, *et al.* (2015) Impact of vitamin D on infectious disease. *Am J Med Sci* **349**, 245–262.
112. Langlois PL, Szwec C, D'Aragnon F, *et al.* (2018) Vitamin D supplementation in the critically ill: a systemic review and meta-analysis. *Clin Nutr* **37**, 1238–1246.
113. Hewison M (2011) Vitamin D and innate and adaptive immunity. *Vitam Horm* **86**, 23–62.
114. Rivers EP, Jaehne AK, Nguyen HB, *et al.* (2013) Early biomarker activity in severe sepsis and septic shock and a contemporary review of immunotherapy trials: not a time to give up, but to give it earlier. *Shock* **39**, 127–137.