



First seroprevalence survey of bovine anaplasmosis: an emerging tick-borne disease in commercial livestock and dairy farms in Bangladesh

Research Article

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Abstract

Bovine anaplasmosis is an infectious, tick-borne disease caused by *Anaplasma* species, which is accountable for huge economic loss in dairy industry. This study was aimed to determine the seroprevalence of bovine anaplasmosis on randomly selected 61 commercial dairy farms in 3 intensive regions of Bangladesh. A total of 1472 sera were analysed using VMRD *Anaplasma* Antibody Test Kit cELISA v2 for the presence of *Anaplasma*-specific antibodies. The highest regional seroprevalence of *Anaplasma* was 45.93% in individual level and 74.4% in herd level recorded in the southeast region, whereas it was 48.8% in individual level and 83.3% in herd level in Khagrachari and Sherpur districts, indicating an emerging state of the disease. The herd size and type in herd level and regions, districts, sex, age and breed in individual level were significantly ($P \leq 0.05$) associated with anaplasmosis. Multivariate logistic regression analysis showed that cattle aged >1 year had 1.86 times higher odds compared to cattle younger than 1 year. Dairy cows had the highest odds (2.25) of anaplasmosis, followed by dairy heifers (1.68), compared to bulls. Compared to herd sizes of <4, the odds of *Anaplasma* infection were 11.3 and 7.45 times greater in herd sizes of >28 and 4–28. Crossbred cattle had 2.4 times higher odds of anaplasmosis compared to indigenous cattle. This first seroprevalence study signifies the widespread presence and underscores the importance of monitoring and managing anaplasmosis to safeguard cattle health in Bangladesh. Study on the molecular epidemiology and genetic diversity of *Anaplasma* among cattle populations should be prioritized.

Introduction

Bovine anaplasmosis is a highly transmissible tick-borne disease that affects cattle and other ruminants (Wathanadirek *et al.*, 2019), primarily caused by *Anaplasma marginale*. The disease is endemic in tropical and subtropical regions worldwide, causing significant health issues and economic losses in the livestock industry. Other species of *Anaplasma*, such as *A. centrale*, *A. bovis* and *A. phagocytophilum*, also cause various forms of the disease in cattle (Ybañez and Inokuma, 2016). Transmission of *Anaplasma* species occurs through biological vectors (ticks), mechanical vectors (biting flies, fomites) (Radostits and Done, 2007; Kocan *et al.*, 2010; Aubry and Geale, 2011) and rarely through the placenta (Van Loo *et al.*, 2023). Approximately, 20 tick species have been reported as vectors of *A. marginale* globally (Radostits and Done, 2007); however, *Rhipicephalus microplus* was identified as a main natural vector in Bangladesh (Roy *et al.*, 2018). Biological vectors can maintain and propagate *A. marginale* for a significant length of time, making them crucial for disease transmission. However, some strains of *A. marginale* may rely on rapid mechanical transfer due to the limited quantity of the agent transferred (Kocan *et al.*, 2010; Aubry and Geale, 2011).

Anaplasmosis is clinically characterized by showing general weakness, weight loss, fever, severe anaemia, pale mucous membranes, abortion, lethargy, icterus, decreased milk production and often death in animals older than 2 years (Kocan *et al.*, 2015). The severity of the disease depends on some factors, such as the host's immunological state and the presence of other pathogens (Constable *et al.*, 2017). Recovered cattle may develop persistent infection which is considered an important epidemiological factor for bovine anaplasmosis. It has been observed that recovered cattle from acute cases, even those that have been treated with the recommended doses of tetracycline, continue to maintain a microscopically undetectable parasitaemia for their entire lives (Palmer *et al.*, 2000; Radostits and Done, 2007; Kocan *et al.*, 2010; Aubry and Geale, 2011). Persistently infected cattle that are exposed to mechanical and/or biological vectors can act as reservoirs of infection to introduce *A. marginale* into naive cattle populations (de Echaide *et al.*, 2001; Futse *et al.*, 2003).

Bangladesh has around 25.7 million cattle, demonstrating the importance of dairy and meat production in the country (World Bank, 2018). Bovine anaplasmosis has a severe economic

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impact on the dairy industries by reducing weight gain, milk and meat production, abortion, icterus and even death (Hove *et al.*, 2018; Okafor *et al.*, 2018). Several studies have been carried out previously on the subclinical and clinical bovine anaplasmosis in Bangladesh (Samad *et al.*, 1989; Talukder and Karim, 2001). A higher frequency of subclinical anaplasmosis (33%) was detected in the milk vita region of Sirajganj district (Talukder and Karim, 2001). On the other hand, 70% anaplasmosis was detected in cattle with possible clinical signs (Chowdhury *et al.*, 2006), and 1% prevalence of haemoprotozoan parasites was reported in Red Chittagong Cattle of Chattogram district (Siddiki *et al.*, 2010), 22.74% in Sylhet (Akter *et al.*, 2018) and 18.67% in Sirajganj (Islam *et al.*, 2019) based on the microscopic examination of Giemsa-stained blood smear. About 43% prevalence of bovine anaplasmosis was detected in Dhaka (Hassan *et al.*, 2019), 15.75% in Chattogram (Mannan *et al.*, 2022) and 82.86% in Bandarban (Mohanta *et al.*, 2023) through polymerase chain reaction (PCR). Although several epidemiological studies have been performed on bovine anaplasmosis in different regions of Bangladesh based on the microscopic examination of Giemsa-stained blood smears and PCR, the seroprevalence of bovine anaplasmosis has not yet been addressed in Bangladesh. The competitive enzyme-linked immunosorbent assay (cELISA) test is advised for population monitoring and screening, whereas PCR and microscopic examination of blood smears are advised for the investigation of clinical cases, according to diagnostic assays used in veterinary medicine for the detection of *A. marginale* and *A. centrale*. Currently, the prevalence of bovine anaplasmosis and its economic consequences have become a concerned issue in the country. Climate change, vector diversity and diverse geographical areas are making the situation more critical to control disease transmission and prevention. Knowledge regarding the local or regional prevalence of bovine anaplasmosis is required for the effective implementation of control strategies. In order to execute effective management programmes of bovine anaplasmosis in Bangladesh, it is imperative to determine the seroprevalence, which may serve as a lookout for estimating the prevalence of the disease in the study area.

The present study investigated the seroprevalence of bovine anaplasmosis in commercial dairy farms in the northeast, central and southeast regions of Bangladesh using cELISA to provide comprehensive data to the scientific community for future planning to control the disease. The geographical locations of the 3 zones are characterized by plain, hilly and riverine areas. Therefore, the present study was conducted for the first time in Bangladesh to monitor the health status of livestock animals to detect the presence of *Anaplasma* infections by serological assay.

Materials and methods

Study area

The study was conducted in 3 dairy intensive regions of Bangladesh, viz., northeast, central and southeast regions (Fig. 1), from October 2022 to March 2024. The northeast region includes Mymensingh, Sherpur, Jamalpur and Netrakona districts, whereas the southeast region comprises Chattogram and Khagrachari districts and Dhaka, Gazipur and Narsingdi districts belong to the central region, respectively. These selected districts were promising for crossbreed dairy farming because of the growing demand for food derived from animals, the high density of the cattle population, the great potential for productivity enhancement, the agro-ecological conditions that support the production of feed, the accessibility of crop residues and the option of mixed crop-livestock farming (World Bank, 2018). The study area had more than one-third of the cattle farms of Bangladesh combined

(Huque and Khan, 2017) and the list of dairy farms was obtained from sub-district (Upazila) livestock offices posted in the respective district. The list of farms (sampling frame) from these districts of Bangladesh was entered into a spreadsheet (Microsoft Excel 2010). Each farm was assigned an Excel-generated random number using the 'rand' function, and 61 farms were selected. Then the herds were randomly selected from the sampling frame (Islam *et al.*, 2020). The farms with 2 cattle and at least 1 mature cattle were considered as an inclusion criterion for this study. All animals on the farm were included in the study, including weak and emaciated animals, with the exception of calves under 6 months of age and those in advanced pregnancy (>8 months). Geographic coordinates of each selected cattle farm were captured during blood sample collection using a handheld global positioning system reader (Garmin eTrex 10) (Islam *et al.*, 2020). ArcGIS-ArcMap version 10.3 (Environmental System Research Institute, Redlands, CA, USA) was used to visualize the spatial distribution of the cattle farms included in this study (Rahman *et al.*, 2015).

Calculation of sample size and sampling procedure

The sample size was determined by Cochran's sample size formula (Cochran, 1977) for categorical data for an α level *a priori* at 0.05 (error of 5%), $n_0 = (t)^2 * (p)(q)/(d)^2$ [where: n_0 is the sample size; t is the value for the selected α level, e.g. 1.96 for (0.25 in each tail) a 95% confidence level; p (5%) is the expected proportion of an attribute that is present in the population; q is $1-p$; (p) (q) are the estimates of variance; d is the acceptable margin of error for the proportion being estimated, so the confidence interval, in decimals). A total of 1472 blood samples (552 samples from the northeast, 442 from the southeast and 478 samples from the central regions) were obtained from 61 commercial dairy farms in these 3 study regions.

Blood collection

Prior to collection of blood samples, farm owners' oral consent was obtained. From each cattle, 8 mL of blood was withdrawn *via* jugular venipuncture with disposable needles and 6 mL of blood was put into serum collection tubes, labelled and transferred to the laboratory of the Department of Parasitology, BAU, on ice (after clotting) within 12 h. Sera were extracted 1 day later by centrifuging at 3000×g for 30 min, after which blood samples were kept refrigerated (2–8°C) in the laboratory. Each serum sample was labelled with the animal's identification number and stored at –20°C.

Serological study using cELISA

All serum samples (1472) were analysed for the presence of *Anaplasma*-specific antibodies using a commercially available cELISA kit (Veterinary Medical Research and Development Inc., Pullman, WA, USA) according to the manufacturer's instructions and published literature (Parvizi *et al.*, 2020). The wells of the ELISA plates were coated with *Anaplasma* spp. antigen provided with the commercial kits. The commercial kits included both positive and negative control sera for this assay. The optical densities of the samples were measured at 620 nm using an ELISA reader. The inhibition per cent was computed as $I\% = 100 (1 - [\text{sample OD}_{620}/\text{OD}_{620} \text{ of the negative control}])$ to understand the results. Any sample with <30 and $\geq 30\%$ was considered negative and positive, respectively. The manufacturer states that the test has >99% specificity and sensitivity with this cut-off. If a single animal was positive for *Anaplasma* infection, we considered the herd as positive (Islam *et al.*, 2020).

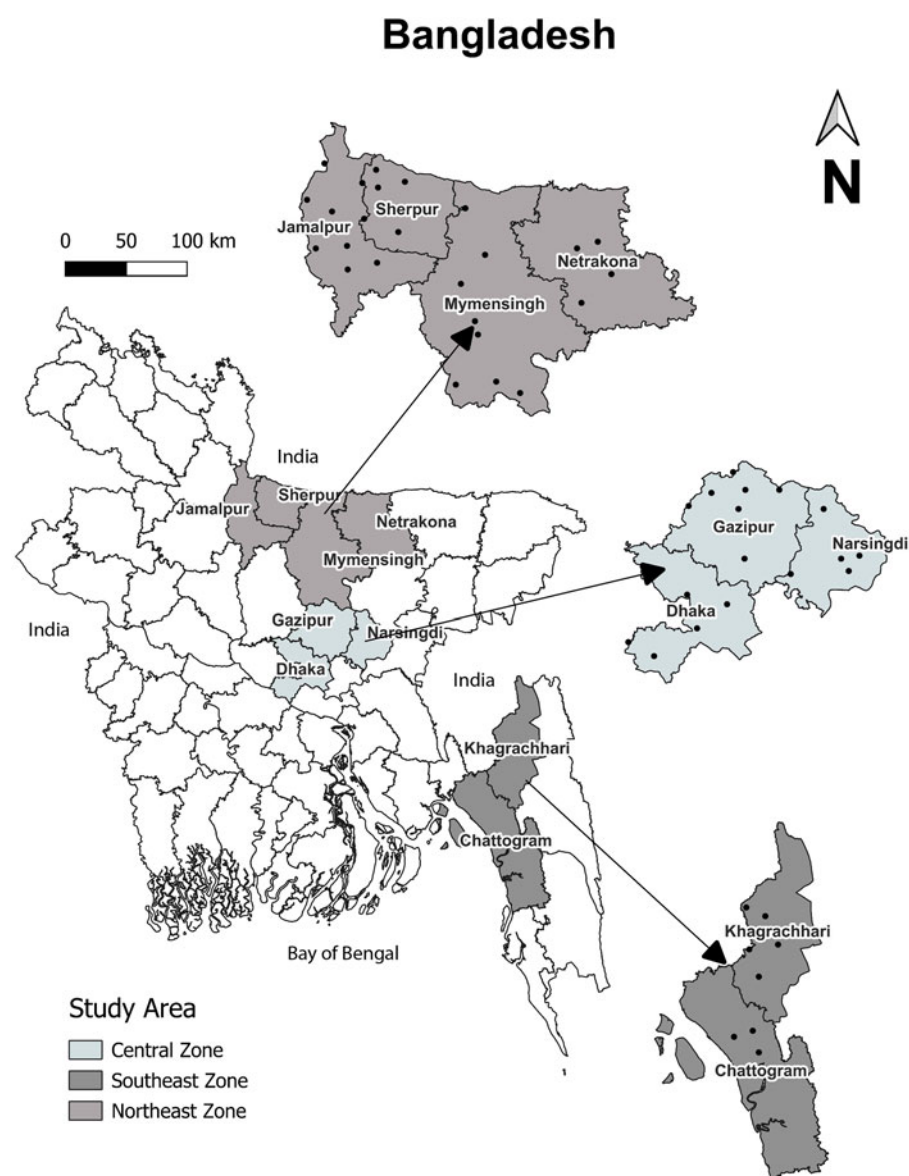


Figure 1. Map of the study districts of Bangladesh. A total of 61 cattle farms of 9 districts were surveyed; black circles are the GIS coordinates of selected farm.

Data management and analysis

Animal, farm-level data and laboratory findings were entered into a spreadsheet (Microsoft Excel 2010). The dataset was coded, checked, validated for integrity and exported to SPSS Statistics software[®], which was used to analyse the data (IBM Corp., Armonk, NY, USA, version 25). We calculated the mean and standard deviation (s.d.) for continuous variables and calculated proportions and frequency distributions for categorical variables. All continuous predictor variables [herd size, age of the animal, sex, breed and cattle raised for various purposes (calf, dairy heifer, beef heifer, bull and dairy cows)] were categorized prior to logistic regression analysis. Initially, univariable mixed-effects logistic regression analyses were performed to find out the effect of individual risk factors on *Anaplasma* infection. The variables that were statistically significant ($P < 0.05$) in the univariate analysis were selected as potential candidates for the multivariable analysis to find out the interaction of different variables. A backward stepwise elimination approach was applied in the multiple logistic regression. Variables with a P value < 0.05 were retained in the final mixed-effects logistic regression model. Collinearity among explanatory variables was assessed by Cramer's phi-prime statistic and a pair of variables was considered collinear if Cramer's phi-prime statistic was > 0.70 (Rahman *et al.*, 2017).

Results

Descriptive epidemiology

A total of 1472 dairy cattle from 61 randomly selected dairy farms, with a herd size of 126 (interquartile range, IQR), across 3 regions were sampled in the investigation. Overall, 42.93% of the cattle were recorded as seropositive in the study. The majority of the sampled cattle (37.5%) were from the northeast region, followed by the central region (32.5%) with the southeast region contributing the smallest proportion (30%) (Table 1). The highest number of seropositive animals was found in the northeast region ($n = 221$). In this study, the majority of cattle were sampled from Mymensingh (21%), followed by Chattogram (18.5%), Gazipur (13.7%), Khagrachhari (11.5%), Dhaka (11%), Jamalpur (7.6%), Narsingdi (7.4%), Netrakona (5%) and Sherpur (3.7%). Mymensingh district recorded the highest number of seropositive animals ($n = 138$). Two-thirds (73.4%) of the sampled cattle were female, with females showing higher seropositivity, accounting for 482 of the seropositive cases. The age distribution of the population under study was nearly equal. Cattle older than 1 year exhibited a higher number of seropositive cases ($n = 348$). Calves, which comprised 48.2% of the total population, had the highest number of seropositive cases ($n = 284$) among all animals. Among adult cattle, dairy cows showed the highest seropositivity ($n = 202$). The majority of the sampled cattle were

Table 1. Overall status of seroprevalence on different animal-level parameters in cattle ($N=1472$) in the study areas of Bangladesh

Parameters	Total animals N (%)	Seropositive animals n	Overall seroprevalence (%)	95% confidence interval
Regions				
Northeast	552 (37.5)	221	15.01	13.19–16.83
Central	478 (32.5)	208	14.13	12.35–15.91
Southeast	442 (30.0)	203	13.79	12.03–15.55
Districts				
Dhaka	167 (11)	63	4.28	3.25–5.31
Gazipur	202 (13.7)	97	6.59	5.32–7.86
Narsingdi	109 (7.4)	48	3.26	2.365–4.17
Mymensingh	310 (21)	138	9.38	7.89–10.87
Jamalpur	112 (7.6)	38	2.58	1.77–3.39
Sherpur	55 (3.7)	22	1.49	0.87–2.11
Netrakona	75 (5)	23	1.56	0.93–2.19
Chattogram	272 (18.5)	120	8.15	6.75–9.55
Khagrachari	170 (11.5)	83	5.64	4.47–6.83
Sex				
Male	392 (26.6)	150	10.19	8.64–11.74
Female	1080 (73.4)	482	32.74	30.34–35.14
Age				
<1 year	710 (48.2)	284	19.29	17.27–21.31
>1 year	762 (51.8)	348	23.64	21.47–25.81
Herd type				
Calf	710 (48.2)	284	19.29	8.64–11.74
Dairy heifer	222 (15.1)	97	6.58	5.31–7.85
Beef heifer	26 (1.8)	8	0.54	0.17–0.91
Bull	126 (8.6)	41	2.79	1.95–3.63
Dairy cow	388 (26.3)	202	13.72	11.96–15.48
Breed				
Crossbred	1116 (75.82)	507	34.44	32.01–36.87
Local	356 (24.18)	125	8.49	7.07–9.91
Overall				
Total	1472 (100)	632	42.93	40.40–45.46

crossbred (75.82%), with a total of 507 seropositive cases in this group (Table 1).

Herd- and individual-level seroprevalence of anaplasmosis

Regional seroprevalence

In the study, an overall herd-level seroprevalence of 70.6% ($n=89$) was observed, while the individual cattle-level seroprevalence was 42.93% ($n=692$). Among the 3 regions, the highest herd-level seroprevalence of anaplasmosis was observed in the southeast region (74.4%, $n=29$), followed by the central (69.8%, $n=30$) and northeast (68.2%, $n=30$) regions (Table 2). At the individual cattle level, the southeast region also had the highest seroprevalence (45.93%, $n=203$), followed by the central (43.51%, $n=208$) and northeast (40.04%, $n=221$) regions (Table 2). The results indicated that the cattle populations in the southeast, central and northeast regions of Bangladesh

exhibited a high seroprevalence of *Anaplasma* infections. This widespread presence underscores the importance of monitoring and managing *Anaplasma* infections to safeguard cattle health in these regions.

District-level seroprevalence

Among the districts, Sherpur district had the highest herd-level seroprevalence (83.3%, $n=5$), followed by Gazipur district (81.8%, $n=9$), despite the small herd sizes in both cases. Beyond these, Chattogram (80.8%, $n=21$) exhibited the highest seroprevalence, followed by Mymensingh (75%, $n=18$), Dhaka (73.7%, $n=14$), Jamalpur (57.14%, $n=4$) and Netrakona (50%, $n=3$) districts and Khagrachari (61.5%, $n=8$) exhibited the lowest herd-level seroprevalence, respectively (Table 3). Additionally, at individual cattle level, the highest percentages of seropositive cattle were recorded in Khagrachari (48.8%, $n=83$), followed by Gazipur (48%, $n=97$), Mymensingh (44.5%, $n=138$),

Table 2. Seroprevalence of *Anaplasma* infections among different regions of Bangladesh diagnosed by competitive enzyme-linked immunosorbent assay (cELISA)

Regions	Total number of farms	Herd levels		Individual animal levels	
		Positivity % n/N	95% CI	Positivity % n/N	95% CI
Northeast	17	68.2 (30/44)	64.8–71.3	40.04 (221/552)	35.92–44.30
Central	26	69.8 (30/43)	63.3–68.7	43.51 (208/478)	39.02–48.10
Southeast	18	74.4 (29/39)	69.9–78.1	45.93 (203/442)	40.46–45.60
Total	61	70.6 (89/126)	62.7–78.6	42.93 (632/1472)	40.40–45.46

CI, confidence interval; n/N, number of positive/number of examined.

Chattogram (44.4%, $n = 120$), Narsingdi (44%, $n = 48$), Sherpur (40%, $n = 22$), Dhaka (37.7%, $n = 63$), Jamalpur (33.9%, $n = 38$) and Netrakona (30.7%, $n = 23$) districts, respectively (Table 3).

Bovine anaplasmosis risk factors in individual cattle level

At the individual cattle level, regions, districts, sex, age, herd type, herd size and breed were all significantly ($P \leq 0.05$) associated with anaplasmosis (Table 4). The regional- and district-level seroprevalence is described in the previous sections of the study. The southeast region and Khagrachari district had the highest seroprevalence, with both being significantly associated ($P = 0.05$) with higher rates of infection (Table 4). Additionally, the univariable analysis revealed that female animals had a significantly ($P = 0.04$) higher prevalence of *Anaplasma* infections (44.6%, $n = 482$) compared to male animals (38.3%, $n = 150$). A higher prevalence of *Anaplasma* infections was recorded in cattle older than 1 year (45.7%, $n = 348$), while a lower prevalence was observed in cattle younger than 1 year (40%, $n = 284$) (Table 4). The present study was carried out on different cattle herd types, i.e. calves, dairy heifers, beef heifers, bulls and dairy cows among the cattle population in the study areas. The results of univariate analysis indicated that *Anaplasma* infections were significantly ($P = 0.0001$) more prevalent in dairy cows (52.1%, $n = 202$) and dairy heifers (43.7%, $n = 97$), followed by calves (40%, $n = 284$), bulls (32.5%, $n = 41$) and beef heifers (30.8%, $n = 8$), respectively (Table 4). The herd size of animal farms was significantly ($P \leq 0.05$) associated with anaplasmosis, as revealed by univariable logistic regression analysis. The seroprevalence of *Anaplasma* infection was significantly ($P =$

0.002) higher in herds with more than 28 animals (81.1%, $n = 30$) compared to herds with fewer than 4 animals. It was also significantly ($P = 0.01$) higher in herds with 4–28 animals (72.6%, $n = 53$). The study also revealed that crossbred animals had a significantly ($P = 0.001$) higher seroprevalence of *Anaplasma* infection (45.4%, $n = 507$) compared to indigenous cattle breeds (35.4%, $n = 125$) (Table 4).

Regions and districts with age groups and herd types with sex groups were collinear (Cramer's phi-prime statistic > 0.70). Therefore, regions, districts and sex were excluded from the multivariable logistic regression analysis. The odds ratio (OR) of anaplasmosis was significantly ($P = 0.01$) higher in cattle aged > 1 year, with an OR of 1.86 (95% CI 1.4–3.1), compared to cattle aged < 1 year (Table 5). For herd type, significantly ($P = 0.001$) dairy cows had the highest odds of *Anaplasma* infection (2.25 times, 95% CI 1.48–3.44) followed by dairy heifer (1.68 times, 95% CI 1.02–2.54), compared to bulls. Compared with a herd size of < 4 , the odds of *Anaplasma* infection were significantly ($P \leq 0.001$) 11.3 (95% CI 7.9–28.2) and 7.45 times (95% CI 4.6–21.56) greater in herd sizes of > 28 and 4–28, respectively. Crossbred cattle had significantly ($P = 0.001$) higher odds of *Anaplasma* infection, increasing the risk by 2.4 times (95% CI 1.68–3.94) compared to indigenous (*Bos indicus*) cattle (Table 5).

Discussion

Anaplasma is a tick-borne pathogen that can cause disease in cattle, leading to economic losses in the livestock industry (Rodríguez *et al.*, 2009). In developing countries like Bangladesh, where there may be limited resources for tick control

Table 3. Seroprevalence of *Anaplasma* infections among different districts diagnosed by competitive enzyme-linked immunosorbent assay (cELISA)

Factors	Districts	Total number of farms	Herd level		Individual animal level	
			Positivity % n/N	95% CI	Positivity % n/N	95% CI
Zones						
Central	Dhaka	6	73.7 (14/19)	69.3–74.7	37.7 (63/167)	30.35–45.05
	Gazipur	6	81.8 (9/11)	79.6–84.4	48.0 (97/202)	41.11–54.89
	Narsingdi	6	50.0 (7/14)	48.4–51.6	44.0 (48/109)	34.68–53.32
Northeast	Mymensingh	9	75.0 (18/24)	70.6–79.4	44.5 (138/310)	38.97–50.03
	Jamalpur	7	57.14 (4/7)	51.1–62.9	33.9 (38/112)	25.13–42.67
	Sherpur	6	83.3 (5/6)	74.2–91.8	40.0 (22/55)	27.05–52.95
	Netrakona	6	50.0 (3/6)	46.8–53.2	30.7 (23/75)	20.26–41.14
Southeast	Chattogram	9	80.8 (21/26)	76.8–85.2	44.4 (120/272)	38.50–50.30
	Khagrachari	6	61.5 (8/13)	59.3–64.7	48.8 (83/170)	41.29–56.31
Total		61	70.6 (89/126)	62.7–78.6	42.93 (632/1472)	40.40–45.46

CI, confidence interval; n/N, number of positive/number of examined.

Table 4. Univariate logistic regression analysis between demographic characteristics and *Anaplasma* seroprevalence among cattle in different selected dairy farms in Bangladesh

Variables	Total number of animals (N)	Seroprevalence (%) positive No. (n)	Odd ratio (95% CI)	P value
Regions				
Northeast	552	40.04 (221)	Reference	–
Central	478	43.51 (208)	1.16 (0.9–1.48)	0.28
Southeast	442	45.93 (203)	1.29 (1–1.65)	0.05*
Districts				
Dhaka	167	37.7 (63)	Reference	–
Gazipur	202	48.0 (97)	0.99 (0.63–1.54)	1
Narsingdi	109	44.0 (48)	1.3 (0.79–2.12)	0.35
Mymensingh	310	44.5 (138)	1.29 (0.88–1.89)	0.23
Jalpur	112	33.9 (38)	0.84 (0.6–1.40)	0.6
Sherpur	55	40.0 (22)	1.8 (0.64–2.2)	0.7
Netrokona	75	30.7 (23)	0.78 (0.43–1.38)	0.47
Chattogram	272	44.4 (120)	1.3 (0.9–1.96)	0.2
Khagrachari	170	48.8 (83)	1.6 (1.02–2.43)	0.05*
Sex				
Male	392	38.3 (150)	Reference	–
Female	1080	44.6 (482)	1.29 (1.01–1.62)	0.04*
Age				
<1 year	710	40.0 (284)	Reference	–
>1 year	762	45.7 (348)	1.26 (1.03–1.55)	0.03*
Herd type				
Calf	710	40.0 (284)	1.22 (0.94–1.61)	0.13
Dairy heifer	222	43.7 (97)	1.34 (1.00–1.79)	0.04*
Beef heifer	26	30.8 (8)	0.94 (0.50–1.77)	0.86
Bull	126	32.5 (41)	Reference	–
Dairy cow	388	52.1 (202)	1.59 (1.22–2.09)	0.0001*
Herd size				
Small herd (<4)	16	37.5 (6)	Reference	–
Medium herd (4–28)	73	72.6 (53)	4.42 (1.42–13.75)	0.01*
Large herd (>28)	37	81.1 (30)	7.14 (1.94–26.32)	0.002*
Breed				
Indigenous	356	35.4 (125)	References	–
Crossbred	1116	45.4 (507)	1.54 (1.2–1.97)	0.001*

CI, confidence interval.

*Significant at $P \leq 0.05$ level.

and veterinary care, bovine anaplasmosis becomes a major problem. This is the first seroprevalence report of *Anaplasma* infections in the cattle population of Bangladesh. In this study, we estimated seroprevalence of *Anaplasma* based on the herd and cattle level in the 9 intensive dairy rearing districts of Bangladesh and identified risk factors for *Anaplasma* infection in cattle.

The study revealed that the seroprevalence of anaplasmosis at regional level varied from 40 to 46% in individual cattle level and 66 to 74% at herd level and the highest seropositivity was found in the southeast region for both cases. In addition, seropositivity was between 32 and 49% at the individual cattle level, while it was 50 and 83% at the herd level in 9 study districts. The study revealed that cattle from Khagrachari, Gazipur, Chattogram and Mymensingh had high seropositivity. This suggests that these animals have had

previous or ongoing infections with *Anaplasma* spp. The seroprevalence of the *Anaplasma* infection in the present study was consistent with those reported previously from the neighbouring country, India, particularly in southern Rajasthan, India, where the seropositivity was 42.28% in cattle and 48.72% in organized cattle herds, respectively (Sharma *et al.*, 2015; Sarangi *et al.*, 2021). Another seroprevalence study reported 34 and 46% seropositivity for bovine anaplasmosis in India and globally (Paramanandham *et al.*, 2019). However, the present study findings conflicted with those reports, where seropositivity of *Anaplasma* infection was 15.02% in Texas (Hairgrove *et al.*, 2014) and 18.5% in Egypt (Parvizi *et al.*, 2020), respectively.

At the individual cattle level, regions, districts, sex, age and breed were identified as potential risk variables for *Anaplasma*

Table 5. Multivariate logistic regression analysis of important variables ($P < 0.05$) after collinearity checking associated with *Anaplasma* seroprevalence among cattle in different selected dairy farms in Bangladesh

Variables	Total seropositive animals (n)	Odds ratio (95% CI)	P value
Age			
<1 year	284	Reference	
>1 year	348	1.86 (1.4–3.1)	0.01*
Herd type			
Bull	41	Reference	–
Dairy heifer	97	1.68 (1.02–2.54)	0.02*
Dairy cow	202	2.25 (1.48–3.44)	0.001*
Herd size			
Small herd (<4)	06	References	
Medium herd (4–28)	53	7.45 (4.6–21.56)	<0.001*
Large herd (>28)	30	11.3 (7.9–28.2)	<0.001*
Breed			
Indigenous	507	References	
Crossbred	125	2.4 (1.68–3.94)	0.001*

CI, confidence interval.

*Significant at $P \leq 0.05$ level.

infection test-positivity, while herd type and herd size were identified as risk variables at the herd level. In the present study, age was determined to be one of the potential risk variables for bovine anaplasmosis. The seroprevalence of *Anaplasma* infections in cattle aged >1 year had around 2 times higher odds of bovine anaplasmosis compared to that of cattle aged <1 year. The findings were in line with other previous published reports where *Anaplasma* infections increase significantly with age and have the highest prevalence in adults more than 1 year old (Chowdhury *et al.*, 2006; Kocan *et al.*, 2010; Alim *et al.*, 2012; Atif *et al.*, 2012). This higher seropositivity in adults compared to young animals might be due to a higher chance to pick up the *Anaplasma* infection as they stay on the farm longer than male cattle. However, these findings conflict with those reports where anaplasmosis was more common in young animals than in adult cattle (Nazar *et al.*, 2018; Khan *et al.*, 2019).

In this study, breed was also identified as a potential risk for the occurrence of *Anaplasma* infections in cattle. Crossbred cattle are 2.4 times more prone to anaplasmosis compared to local/indigenous cattle. The present finding was consistent with the previous reports on anaplasmosis, highlighting the vulnerability of crossbred cattle to *Anaplasma* infections (Ananda *et al.*, 2009; Siddiki *et al.*, 2010). In addition, previous reports observed a higher prevalence of infection in exotic breeds and their crosses compared to local breeds of cattle (Chowdhury *et al.*, 2006; Atif *et al.*, 2012; Farooqi *et al.*, 2018; Khan *et al.*, 2019; Shoaib *et al.*, 2021). This is attributed to the fact that exotic breeds and their crosses are more susceptible to tick infestation. The lower frequency in indigenous cattle could be due to constant exposure to diseases, leading to the development of immunity against *Anaplasma* infections. Conversely, the emphasis on the management of crossbred cattle may offer fewer opportunities for pre-exposure to vectors and may result in limited or no immunity,

thereby leading to a higher prevalence of the disease (Bock *et al.*, 1997).

At herd level, herd type also emerged as a potential risk factor for bovine *Anaplasma* infections where cattle were raised for various purposes, viz., calf, dairy heifer, beef heifer, bull and dairy cow. Dairy cows had more than twice the odds of getting *Anaplasma* infection compared to calves, and between dairy heifers and beef heifers, dairy heifers were found to be more susceptible to *Anaplasma* infection while bull had lower odds than dairy cows. Calves are susceptible to anaplasmosis due to transplacental transmission of the disease and may acquire the infection from infected dams through vertical transmission or through exposure to ticks in calving areas or pastures (Radostits *et al.*, 2000; Kocan *et al.*, 2010; Aubry and Geale, 2011; Van Loo *et al.*, 2023). The findings were consistent with the reports where *A. marginale* in dairy animals was higher than bulls and calves (Rajput *et al.*, 2005). The greater prevalence of *A. marginale* in female cattle may be related to lactation in high-producing animals (Kocan *et al.*, 2010) and probably because they are kept longer for breeding and milk production, with diets insufficient to meet their high demands. Additionally, the frequent use of contaminated needles to inject medications for milk let-down may contribute to the increased occurrence of tick-borne diseases in dairy animals. Another report suggested that exposure to *A. marginale* is common in dairy herds (Oliveira *et al.*, 2011).

Size of herds has been reported as a risk factor for anaplasmosis (Okafor *et al.*, 2019; Spare *et al.*, 2020). In our study, herd sizes of >28 and 4–28 had higher odds compared to herd sizes of <4, and the findings were consistent with the previous reports (Okafor *et al.*, 2019; Spare *et al.*, 2020). However, another explanation might be the study design in which more cattle were tested in larger herds, which increases the herd-level sensitivity in larger herds. In conclusion, a substantial proportion of cattle and herds tested positive for *Anaplasma* infection, with herd size and type, age of individuals, sex and breed status significantly associated with the infection in cattle of these selected districts in Bangladesh. The study further suggests that regular health examinations for *Anaplasma* infection in larger herds, especially targeting older cattle, should be done within the context of Bangladesh.

Data availability statement. The data will be available.

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Competing interests. None.

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References

- Akter S, Paul S, Syed SUA, Mandal P and Nazneen S (2018) Prevalence and determinants of bovine anaplasmosis in Sylhet district of Bangladesh. *International Journal of Natural Sciences* **8**, 01–06.
- Alim MA, Das S, Roy K, Masuduzzaman, Sikder MS, Hassan MM, Siddiki AZ and Hossain MA (2012) Prevalence of hemoprotozoan diseases in cattle population of Chittagong Division, Bangladesh. *Pakistan Veterinary Journal* **32**, 221–224.
- Ananda KJ, D'souza PE and Puttalakshamma GC (2009) Prevalence of haemoprotozoan diseases in crossbred cattle in Bangalore North. *Veterinary World* **2**, 15–16.
- Atif FA, Khan MS, Iqbal HJ, Arshad GM, Ashraf E and Ullah S (2012) Prevalence of *Anaplasma marginale*, *Babesia bigemina* and *Theileria annulata* infections among cattle in Sargodha District, Pakistan. *African Journal of Agricultural Research* **7**, 3302–3307. doi: 10.5897/AJAR11.2051
- Aubry P and Geale D (2011) A review of bovine anaplasmosis. *Transboundary and Emerging Diseases* **58**, 1–30. doi: 10.1111/j.1865-1682.2010.01173.x
- Bock R, Vos AD, Kingston T and McLellan D (1997) Effect of breed of cattle on innate resistance to infection with *Babesia bovis*, *B. bigemina* and *Anaplasma marginale*. *Australian Veterinary Journal* **75**, 337–340.
- Chowdhury S, Hossain MA, Barua SR and Islam S (2006) Occurrence of common blood parasites of cattle in Sirajgonj sadar area of Bangladesh. *Bangladesh Journal of Veterinary Medicine* **4**, 143–145.
- Cochran WG (1977) *Sampling Techniques*, 3rd Edn, New York: John Wiley & Sons.
- Constable P, Hinchcliff K, Done S and Grunberg W (2017) Diseases of the haemolymphatic and immune systems. In Cathy Patterson, DVM (Ed.), *Veterinary Medicine: A Textbook of the Diseases of Cattle, Horses, Sheep, Pigs, and Goats*, 11th Edn. London, UK: Saunders Limited, pp. 716–844.
- de Echaide, ST, Knowles, DP, McGuire, TC, Palmer, GH, Suarez, CE and McElwain, TF (2001) Detection of cattle naturally infected with *Anaplasma marginale* in a region of endemicity by nested PCR and a competitive enzyme-linked immunosorbent assay using recombinant major surface protein 5. *Journal of Clinical Microbiology* **39**, 1207–1207.
- Farooqi SH, Ijaz M, Rashid MI, Nabi H, Islam S, Aqib AI, Hussain K, Khan A, Rizvi SNB, Mahmood S and Mehmood K (2018) Molecular epidemiology of bovine anaplasmosis in Khyber Pakhtunkhwa, Pakistan. *Tropical Animal Health and Production* **50**, 1591–1598.
- Futse JE, Ueti MW, Knowles Jr, DP and Palmer GH (2003) Transmission of *Anaplasma marginale* by *Boophilus microplus*: retention of vector competence in the absence of vector-pathogen interaction. *Journal of Clinical Microbiology* **41**, 3829–3834. doi: 10.1128/jcm.41.8.3829-3834.2003
- Hairgrove TB, Craig TM, Budke CM, Rodgers SJ and Gill RJ (2014) Seroprevalence of *Anaplasma marginale* in Texas cattle. *Preventive Veterinary Medicine* **116**, 188–192.
- Hassan MZ, Giasuddin M, Rahman MM, Ershaduzzaman M and Hasan M (2019) Identification of vector-borne blood protozoa in cattle and sheep in Bangladesh. *Journal of Virology & Antivirals* **2**, 004.
- Hove P, Khumalo ZT, Chaisi ME, Oosthuizen MC, Brayton KA and Collins NE (2018) Detection and characterisation of *Anaplasma marginale* and *A. centrale* in South Africa. *Veterinary Sciences* **5**, 26.
- Huque K and Khan MYA (2017) Socio-geographic distribution of livestock and poultry in Bangladesh – a review. *Bangladesh Journal of Animal Science* **46**, 65–81.
- Islam MS, Rahman MS, Hossain MZ and Haque MA (2019) Emerging status of anaplasmosis in cattle in Sirajganj district with therapeutic evaluation of traditional treatments. *Asian-Australasian Journal of Bioscience and Biotechnology* **4**, 162–168.
- Islam SKS, Rumi TB, Kabir SML, Van der Zanden AGM, Kapur V, Rahman A, Ward MP, Bakker D, Ross AG and Rahim Z (2020) Bovine tuberculosis prevalence and risk factors in selected districts of Bangladesh. *PLoS ONE* **15**, e0241717.
- Khan NU, Sarwar MS, Ayaz S, Ali H, Ali A, Khan MA, Khan AU, Hussain M and Ali H (2019) Prevalence and risk factors analysis associated with anaplasmosis in symptomatic cattle under field conditions in southern Khyber Pakhtoonkhwa, Pakistan. *Pure and Applied Biology* **8**, 2119–2127. doi: 10.19045/bspab.2019.80156
- Kocan KM, de la Fuente J, Blouin EF, Coetzee JF and Ewing S (2010) The natural history of *Anaplasma marginale*. *Veterinary Parasitology* **167**, 95–107.
- Kocan KM, de la Fuente J and Cabezas-Cruz A (2015) The genus *Anaplasma*: new challenges after reorganization. *In Revue Scientifique et Technique (International Office of Epizootics)* **34**, 577–586.
- Mannan A, Alim MA, Manik M, Mahub K, Rahman U and Alamgir HM (2022) Prevalence of anaplasmosis in cattle from Chattogram Division of Bangladesh. *Bangladesh Journal of Veterinary and Animal Sciences* **9**, 67–73.
- Mohanta UK, Chikufenji B, Galon EM, Ji S, Ma Z, El-Sayed SAES, Amer MM, Do TT and Xuan X (2023) Molecular characterization and phylogeny of *Anaplasma marginale*, *A. phagocytophilum* and *A. bovis* in livestock of Bangladesh. *Parasitology International* **97**, 102790. doi: 10.1016/j.parint.2023.102790
- Nazar M, Khan MA, Shah AA, Rahman SU, Khan I, Ullah A, Khan IU and Shuaib M (2018) Occurrence and transplacental transmission of *Anaplasma marginale* in dairy cattle. *Slovenian Veterinary Research* **55**, 183–191. doi: 10.26873/SVR-499-2018
- Okafor CC, Collins SL, Daniel JA, Harvey B, Sun X, Coetzee JF and Whitlock BK (2018) Factors associated with seroprevalence of *Anaplasma marginale* in Kentucky cattle. *Veterinary Parasitology: Regional Studies and Reports* **13**, 212–219. doi: 10.1016/j.vprsr.2018.07.003
- Okafor CC, Collins SL, Daniel JA, Coetzee JF and Whitlock BK (2019) Seroprevalence of bovine anaplasmosis in Georgia. *Veterinary Parasitology: Regional Studies and Reports* **15**, 100258.
- Oliveira JB, Montoya J, Romero JJ, Urbina A, Soto-Barrientos N, Melo ESP, Ramos CAN and Araújo FR (2011) Epidemiology of bovine anaplasmosis in dairy herds from Costa Rica. *Veterinary Parasitology* **177**, 359–365.
- Palmer GH, Brown WC and Rurangirwa FR (2000) Antigenic variation in the persistence and transmission of the Ehrlichia *Anaplasma marginale*. *Microbes and Infection* **2**, 167–176.
- Paramanandham K, Mohankumar A, Suresh KP, Jacob SS and Roy P (2019) Prevalence of *Anaplasma* species in India and the World in dairy animals: a systematic review and meta-analysis. *Research in Veterinary Science* **123**, 159–170.
- Parvizi O, El-Adawy H, Melzer F, Roesler U, Neubauer H and Mertens-Scholz K (2020) Seroprevalence and molecular detection of bovine anaplasmosis in Egypt. *Pathogens* **9**, 64.
- Radostits OM and Done SH (2007) *Veterinary Medicine: A Textbook of the Diseases of Cattle, Sheep, Pigs, Goats, and Horses*, 10th Edn. New York: Elsevier Saunders, xxii, p. 2156.
- Radostits OM, Blood DC and Gay CC (2000) *Veterinary Medicine: A Text Book of Disease of Cattle, Sheep, Pigs, Goats and Horse*. 9th Edn. London: Baillere Tindall Publication, pp. 1172–1173, 1289–1290.
- Rahman ASMS, Sumon SMMR, Khan MAHNA and Islam MT (2015) Current status of subclinical form of babesiosis and anaplasmosis in cattle at Rangpur district in Bangladesh. *Progressive Agriculture* **26**, 51–59.
- Rahman AKMA, Islam SKS, Talukder MH, Hassan MK, Dhand NK and Ward MP (2017) Fascioliasis risk factors and space-time clusters in domestic ruminants in Bangladesh. *Parasites & Vectors* **10**, 228.
- Rajput ZI, Song-hua HU, Arijo AG, Habib M and Khalid M (2005) Comparative study of *Anaplasma* parasites in tick carrying buffaloes and cattle. *Journal of Zhejiang University-Science B* **6**, 1057–1062.
- Rodríguez SD, Ortiz MÁG, Ocampo RJ and Murguía YCAV (2009) Molecular epidemiology of bovine anaplasmosis with a particular focus in Mexico. *Infection, Genetics and Evolution* **9**, 1092–1101.
- Roy B, Krücken J, Ahmed J, Majumder S, Baumann M, Clausen PH and Nijhof A (2018) Molecular identification of tick-borne pathogens infecting cattle in Mymensingh district of Bangladesh reveals emerging species of *Anaplasma* and *Babesia*. *Transboundary and Emerging Diseases* **65**, e231–e242.
- Samad MA, Bashar SA, Shahidullah M and Ahmed MU (1989) Prevalence of haemoprotozoan parasites in cattle of Bangladesh. *Indian Journal of Veterinary Medicine* **13**, 50–51.
- Sarangi LN, Rana SK, Prasad A, Ponnanna NM and Sharma GK (2021) Prevalence of antibodies to *Anaplasma* in cattle and buffaloes of different organized herds in India. *Journal of Parasitological Disease* **45**, 359–365.
- Sharma DK, Bhatnagar CS, Bhardawaj B and Meena SK (2015) Incidence of haemoprotozoan diseases in cattle in Southern Rajasthan, India. *International Journal of Current Microbiology and Applied Sciences* **4**, 509–514.
- Shoaib M, Rashid MI, Akbar H, Sheikh AA, Farooqi SH, Khan MA, Khan FA and Khan R (2021) Prevalence and molecular characterization of *Anaplasma marginale* in cattle population of Khyber Pakhtunkhwa

- Province, Pakistan. *International Journal of Agriculture & Biology* 2021, 1560–8530.
- Siddiki AZ, Uddin MB, Hasan MB, Hossain MF, Rahman MM, Das BC, Sarker MS and Hossain MA** (2010) Coproscopic and haematological approaches to determine the prevalence of helminthiasis and protozoan diseases of Red Chittagong Cattle (RCC) breed in Bangladesh. *Pakistan Veterinary Journal* 30, 1–6.
- Spare MR, Hanzlicek GA, Wootten KL, Anderson GA, Thomson DU, Sanderson MW, Ganta RR, Reif KE and Raghavan RK** (2020) Bovine anaplasmosis herd prevalence and management practices as risk-factors associated with herd disease status. *Veterinary Parasitology* 277, 10021.
- Talukder MH and Karim J** (2001) Subclinical *Anaplasma* infection in crossbred cattle in Bangladesh. *Bangladesh Veterinary Journal* 35, 159–160.
- Van Loo H, Pascottini OB, Hooyberghs J, De Bleecker K, Ribbens S, Opsomer G and Pardon B** (2023) Detection of *Anaplasma phagocytophilum* in fetal and placental tissue of bovine abortions and perinatal mortalities. *Veterinary Record* 193, e2880.
- Watthanadirek A, Chawengkirttikul R, Poolsawat N, Junsiri W, Boonmekam D, Reamtong O and Anuracpreeda P** (2019) Recombinant expression and characterization of major surface protein 4 from *Anaplasma marginale*. *Acta Tropica* 197, 105047.
- World Bank** (2018) Combined project information documents/integrated safeguards datasheet (PID/ISDS). Livestock development-based dairy revolution and meat production project (P161246).
- Ybañez AP and Inokuma H** (2016) *Anaplasma* species of veterinary importance in Japan. *Veterinary World* 9, 1190–1196. doi: 10.14202/vetworld.2016.1190-1196