

Effect of Water Stress on the Growth and Fecundity of Common Waterhemp (*Amaranthus rudis*)

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Common waterhemp is one of the most commonly encountered and troublesome weeds in the midwestern United States. It is well known that water stress adversely affects crop growth and yield; however, the effects of water stress on weed growth and seed production are poorly understood. The objective of this study was to determine the effects of degree and duration of water stress on growth, development, and fecundity of two common waterhemp biotypes in greenhouse experiments conducted at the University of Nebraska–Lincoln. No difference was observed in growth, development, and seed production between two biotypes in response to degree and duration of water stress; therefore, data were combined. The degree of water stress study included five treatments, where the amount of water applied to each pot at 2-d interval was equivalent to 100, 75, 50, 25, and 12.5% of pot (soil) water content. The highest plant height (163 cm), number of leaves (231 plant⁻¹), and growth index (4.4×10^5 cm³) were recorded at 100% of pot water content (no water stress). Similarly, aboveground biomass, total leaf area, and seed production reached their maximum at 100% of pot water content treatment, whereas they were reduced as degree of water stress increased. The study of water stress duration included five treatments, where amount of water applied to each pot at 2-, 4-, 6-, 8-, and 10-d intervals was equivalent to 100% of pot water content. The highest plant height (150 cm), number of leaves (210 plant⁻¹), and growth index (3.8×10^5 cm³) were observed at 2-d interval of water stress, whereas seed production was similar at 2-d (36,549 seeds plant⁻¹) and 4-d (34,176 seeds plant⁻¹) intervals. This study shows that common waterhemp has capacity to survive and reproduce even under a higher degree and duration of water stress.

Nomenclature: Common waterhemp; *Amaranthus rudis* Sauer.

Key words: Aboveground biomass, degree of water stress, duration of water stress, growth index, seed production.

Common waterhemp, a C₄ species, is a summer annual broadleaf weed native to North America (Waselkov and Olsen 2014). It is the most problematic and troublesome weed in row-crop production systems throughout the midwestern United States (Hager et al. 2002; Shoup et al. 2003). In Illinois, season-long infestation of common waterhemp reduced corn (*Zea mays* L.) yield up to 74% (Steckel and Sprague 2004), and soybean (*Glycine max* L.) yield was reduced by 43% when common waterhemp plants were allowed to compete up to 10 wk after soybean unifoliolate expansion (Hager et al. 2002). Changes in cultural practices and weed management strategies have helped to increase the crop

productivity in the midwestern United States, but these changes are also believed to aid in the shifting of the weed flora composition and have resulted in the dominance of small-seeded broadleaf weeds, including common waterhemp (Hausman et al. 2011).

Favorable biological attributes and the rapid evolution of herbicide resistance contributed to the dominance of common waterhemp as a successful weed in corn–soybean production systems. Common waterhemp has a rapid growth habit with a high biomass production potential. A study conducted in Kansas revealed that height of common waterhemp increased 0.11 to 0.16 cm per growing degree day at a relative growth rate of 0.31 g g⁻¹ d⁻¹ (Horak and Loughin 2000). This weed can emerge throughout the growing season, starting from mid-May depending on environmental conditions, making common waterhemp more capable than most weeds of escaping herbicide applications (Hartzler et al. 1999). Moreover, it has the potential to produce over one million seeds per plant under favorable conditions, thus building up a persistent

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seed bank in a relatively short period (Steckel et al. 2003, 2007).

Overreliance on glyphosate as the only method for weed control in glyphosate-resistant crops has created a selective advantage, resulting in the evolution of glyphosate-resistant weeds. Additionally, common waterhemp is a dioecious and wind-pollinated species with a high potential to disseminate herbicide-resistant traits via pollen movement (Liu et al. 2012). The first report of glyphosate-resistant common waterhemp in the United States was from Missouri (Legleiter and Bradley 2008), but as of 2015, it has been confirmed in 17 states (Heap 2015), including Nebraska (Sarangi et al. 2015). In addition, common waterhemp biotypes resistant to herbicides with other modes of action, including acetolactate synthase inhibitors, growth regulators, 4-hydroxyphenylpyruvate dioxygenase inhibitors, and photosystem II-inhibitors have been confirmed in Nebraska (Bernards et al. 2012; Jhala 2015). Therefore, use of alternate herbicide-tolerant crops, and application of PRE and a premix of POST herbicides are now becoming more common among the growers to control herbicide-resistant weeds, including common waterhemp in Nebraska (Aulakh and Jhala 2015; Chahal and Jhala 2015; Chahal et al. 2014; Ganie et al. 2015; Kaur et al. 2014).

Weeds compete with commodity crops for a variety of environmental resources, including radiation, nutrients, and water. Among them, water is the most limiting factor for the optimum crop production in the Great Plains and midwestern United States (Benjamin and Nielsen 2006). In early and mid-2000s and recently in 2012, many midwestern states, including Nebraska, experienced a severe drought that had an adverse effect on crop yields and the economy (Wu et al. 2013). Water deficit can adversely affect growth and productivity of the crops and associated weed species, though the outcomes of the competition for water depend on the abilities of the crop and weed species to survive under water stress conditions (Begg and Turner 1976; Patterson 1995). The C_4 plants, including common waterhemp, usually have higher water use efficiency and seed production potential that allows them to grow successfully in a wide range of climatic conditions (Long 1999; Lovelli et al. 2010). For example, common waterhemp can be found in places ranging from the arid regions of Texas to the humid/subhumid areas of Maine (Costea et al. 2005; Nordby et al. 2007).

Environmental stresses such as water deficiency prevent plants from achieving the maximum growth

potential set by their genotypes (Patterson 1995). The differences in responses to water stress for different plant species are due to their diverse phenological and physiological processes, and response also depends upon climatic conditions, soil, degree and duration of water stress, and management practices (Irmak et al. 2000). Significant reductions in growth and seed production in some weed species, including Benghal dayflower (*Commelina benghalensis* L.), itchgrass [*Rottboellia cochinchinensis* (Lour.) W. D. Clayton], and junglerice [*Echinochloa colona* (L.) Link] have been reported under different degrees and durations of water stress (Chauhan 2013; Chauhan and Johnson 2010; Webster and Grey 2008). However, availability of limited scientific literature about the water use efficiency of *Amaranthus* species (Liu and Stützel 2002a; 2002b) was the basis of this study. The objective of this study was to determine the effect of degree and duration of water stress on the growth and fecundity of common waterhemp.

Materials and Methods

Plant Materials. Seedheads of two different common waterhemp plants were collected from two soybean fields located at Clay County, and Lancaster County, NE, and placed separately in two paper bags. Seeds were cleaned thoroughly using a seed blower (South Dakota Seed Blower, Seedburo Equipment Co., 1022 W. Jackson Blvd., Chicago, IL) and germinated using the procedure described by Sarangi et al. (2015). Seedlings were transplanted to 72-celled germination trays containing potting mix (Berger BM1 All-Purpose Mix, Berger Peat Moss Ltd., Saint-Modeste, Quebec, Canada) allowing one common waterhemp seedling per cell. When seedlings reached 8 cm in height, they were then transplanted into round, free-draining black plastic pots (20-cm diam and 30-cm ht) containing finely ground soil. Plants were kept in a greenhouse maintained at a 28/24 C day/night temperature and supplied with adequate water and 24–8–16 commercial plant fertilizer (Miracle-Gro Water Soluble All Purpose Plant Food, Scotts Miracle-Gro Products Inc., 14111 Scottslawn Road, Marysville, OH 43041) until the experiment commenced. Metal halide lamps with 600 $\mu\text{mol photon m}^{-2} \text{s}^{-1}$ light intensity provided supplemental light in greenhouse to ensure a 16-h photoperiod.

Pot (Soil) Water Content. Soil used in this study was collected from a field near Lincoln, NE, with no history of residual herbicides applied at least in

the last 5 yr. Air-dried soil was passed through 3-mm sieve to acquire a uniform consistency. The soil texture was silt-loam with a pH of 6.1, 22% sand, 54% silt, 24% clay, 2.8% organic matter, and bulk density of 1.4 g cm^{-3} . Each pot was filled with 10 kg of dry soil and pot (soil) water content was determined by modifying the method described by Steadman et al. (2004). First, the weight of the pots containing dry soil was measured, then the pots were watered to saturation and covered with shiny paper sheets to minimize the evaporation. They were allowed to freely drain for 36 h, and reweighed to calculate the pot water content using the following equation:

$$WC = [(W_w - W_d)/d] \quad [1]$$

where W_w is the wet weight of the soil plus pot, W_d is the dry weight of the soil plus pot, and d is the density of water (i.e., 1 g cm^{-3}).

Experimental Setup. A preliminary study was conducted in the greenhouse under the same growing conditions as described above to determine an effective interval for adding water to the common waterhemp plants. The study included five treatments at 1-, 2-, 3-, 4-, and 5-d intervals of water stress in a randomized complete block design with six replications. In each treatment, water was applied at 100% of pot water content. Plant height, leaves plant^{-1} , and aboveground biomass were measured 45 d after transplanting (DAT). Results showed that plants treated with 100% of pot water content at the 2-d interval resulted in the highest plant height, leaves plant^{-1} , and aboveground biomass compared to other water stress intervals (data not shown); therefore, a 2-d interval was selected as no water stress treatment for degree and duration of water stress study.

Two separate experiments were conducted for both common waterhemp biotypes in the greenhouse at the University of Nebraska–Lincoln. The treatments were selected based on findings of the preliminary study and by modifying the treatments from available literature on water stress (Chauhan 2013; Chauhan and Johnson 2010, Webster and Grey 2008). For this study, water stress treatments were initiated at 10 DAT and continued until plant harvest at 90 DAT.

Degree of Water Stress. Degree of water stress experiment included five water stress treatments, where the amount of water applied to each pot at

2-d interval was equivalent to 100, 75, 50, 25, and 12.5% of pot water content, simulating different degrees of water stress: no, light, moderate, high, and severe water stress, respectively.

Duration of Water Stress. Duration of water stress experiment included treatments of different durations of water stress at 2-, 4-, 6-, 8-, and 10-d intervals. In each treatment, amount of water applied was equivalent to 100% of pot water content.

Pots from both experiments (degree and duration of water stress) were arranged in a randomized complete block design with six replications and experiments were repeated under similar greenhouse environments.

Data Collection. In both experiments, plant height, leaves plant^{-1} , and growth index were determined at 10-d intervals until common waterhemp reached maturity. Growth index is the quantitative indicator for plant growth rate, and was calculated using the following equation (Dhir and Harkess 2011; Irmak et al. 2004):

$$GI(\text{cm}^3) = \pi \times (w/2)^2 \times h \quad [2]$$

where w is the width of the plant calculated as an average of two widths, one measured at the widest point and another at 90° to the first; and h is the plant height measured from soil surface to the last stem-node at the top.

All the leaves from each individual plant were separated from the stem and total leaf area was measured at maturity (90 DAT) using a leaf area meter (LI-3100C Area Meter, LI-COR, Inc. Lincoln, NE). Moreover, aboveground biomass (shoots and leaves) of each common waterhemp plant was bagged separately at maturity (90 DAT) and the roots were washed under a gentle flow of water to remove soil particles. Plant parts were oven-dried at 65 C for 7 d. Aboveground biomass, root biomass, and root:shoot ratio were recorded based on the dry weight of the plant parts. Seeds collected from female common waterhemp plants were threshed and cleaned in the greenhouse using the method described by Steckel et al. (2003). The average weight of five samples of 200 seeds from each plant was recorded and total number of seeds plant^{-1} was calculated. Additionally, germination percentage of common waterhemp seeds obtained from this study was calculated by modifying the method described by Gallagher and Cardina (1998) and Steckel et al. (2003). Two hundred seeds from each

female plant were placed on a piece of moist Whatman No. 4 filter paper (GE Healthcare UK Limited, Amersham Place Little Chalfont, Buckinghamshire, HP7 9NA, U.K.). Petri dishes were kept in the greenhouse with lids closed to prevent microbial contamination and to minimize the water loss through evaporation. The cumulative germination of common waterhemp seeds were counted at 15-d interval up to 45 d after starting the germination study. The percentage of germination was calculated based on seeds germinated vs. number of seeds planted.

Statistical Analysis. Data were subjected to ANOVA using the PROC GLIMMIX procedure in SAS (SAS Institute Inc, Cary, NC). Years (experimental runs) and blocks (nested within year) were considered random effects, whereas biotypes (from Clay County and Lancaster County, NE) and water stress treatments were considered fixed effects in the model. A four-parameter log-logistic sigmoid growth function (Equation 3) was regressed on plant height, leaves plant⁻¹, and growth index using software R (R statistical software, R Foundation for Statistical Computing, Vienna, Austria) (Knezevic et al. 2007):

$$Y = c + \{d - c / 1 + \exp[b(\log x - \log e)]\}. \quad [3]$$

In this model, Y is plant height, leaves plant⁻¹, or growth index at time x (DAT); c is the lower limit considered as 0; d is the estimated maximum plant height or leaf number or growth index; and e is the time taken to reach 50% of final height, leaf number, or growth index. The parameter b is relative slope around parameter e . For the data of total leaf area, biomass, root:shoot ratio, seed production, and percentage of germination, treatment means were separated at $P \leq 0.05$ using Fisher's protected LSD test and plots were generated by using SigmaPlot (SigmaPlot 12.0, Systat Software Inc., San Jose, CA).

Model Goodness of Fit. Root mean square error (RMSE) and modelling efficiency coefficient (EF) were calculated to test the goodness of fit for the model. They are the commonly used to estimate model quality (Werle et al. 2014b; 2014c). The RMSE was calculated based on an equation (Roman et al. 2000):

$$RMSE = \left[\frac{1}{n} \sum_{i=1}^n (P_i - O_i)^2 \right]^{1/2} \quad [4]$$

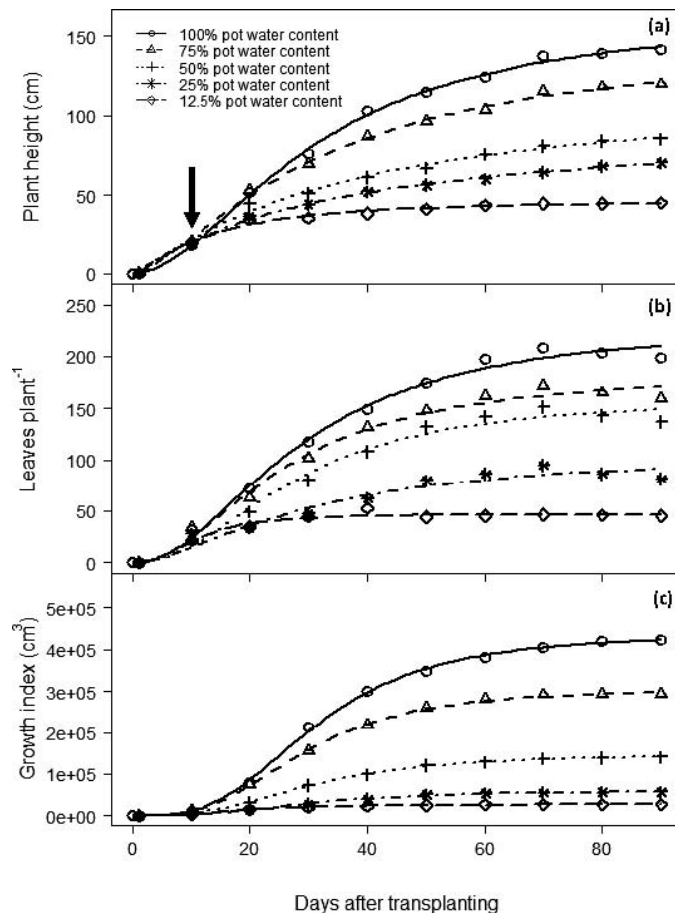


Figure 1. Effect of degree of water stress on (a) height, (b) leaves plant⁻¹, and (c) growth index of common waterhemp in a greenhouse study conducted in Nebraska, where 100, 75, 50, 25, and 12.5% pot water content treatments were considered as no, light, moderate, high, and severe water stress, respectively. The arrow at 10 d after transplanting (DAT) denotes the first day when water stress treatments were imposed.

where P_i is the predicted value, O_i is the observed value, and n is the total number of observations. Smaller RMSE value means better fit to the model due to closer observed and predicted values. The evaluation of R^2 is an inadequate measure for nonlinear models such as Equation 3, as it is extremely biased to highly parametrized models (Spiess and Neumeier 2010); therefore EF, which is different from R^2 by having a lower bound, was calculated (Mayer and Butler 1993):

$$EF = 1 - \left[\frac{\sum_{i=1}^n (O_i - P_i)^2}{\sum_{i=1}^n (O_i - \bar{O}_i)^2} \right] \quad [5]$$

where O_i is the observed value, P_i is the predicted value, \bar{O}_i is the mean observed value, and n is the total number of observations. Generally,

Table 1. Parameter estimates and the goodness of fit (RMSE, and EF)^a of the four-parameter log-logistic function^b fitted to common waterhemp plant height, leaves plant⁻¹, and growth index under different degree of water stress treatments in a greenhouse experiment conducted in Nebraska.

Pot water content (%)	$d^{c,d}$	e (days) ^c	b^c	RMSE	EF
Plant height					
100 (no water stress)	163 ± 11	31 ± 3	-1.8 ± 0.2	16.5	0.90
75 (light water stress)	146 ± 17	31 ± 5	-1.5 ± 0.3	18.4	0.82
50 (moderate water stress)	115 ± 23	35 ± 4	-1.2 ± 0.3	13.8	0.79
25 (high water stress)	93 ± 14	33 ± 4	-1.1 ± 0.2	7.8	0.89
12.5 (severe water stress)	47 ± 3	12 ± 2	-1.4 ± 0.4	7.2	0.78
Leaves plant ⁻¹					
100 (no water stress)	231 ± 15	29 ± 3	-2.0 ± 0.3	31.5	0.84
75 (light water stress)	185 ± 12	26 ± 2	-2.0 ± 0.3	27.8	0.81
50 (moderate water stress)	161 ± 8	28 ± 2	-2.2 ± 0.3	18.7	0.88
25 (high water stress)	105 ± 11	30 ± 5	-1.7 ± 0.4	17.2	0.75
12.5 (severe water stress)	47 ± 3	11 ± 2	-2.4 ± 1.2	21.4	0.32
Growth index ^{e,f}					
100 (no water stress)	4.4 ± 0.2	31 ± 2	-3.1 ± 0.5	0.6	0.87
75 (light water stress)	3.1 ± 0.3	29 ± 3	-3.1 ± 0.9	0.9	0.57
50 (moderate water stress)	1.5 ± 0.1	31 ± 3	-2.8 ± 0.7	0.3	0.69
25 (high water stress)	0.6 ± 0.06	31 ± 4	-2.9 ± 1.1	0.2	0.51
12.5 (severe water stress)	0.3 ± 0.02	19 ± 3	-2.6 ± 0.9	0.1	0.43

^a Abbreviations: RMSE, root mean square error; EF, modelling efficiency coefficient.

^b $Y = c + \{d - c / [1 + \exp\{b(\log x - \log e)\}]\}$, where Y is the plant height, leaves plant⁻¹, or growth index at time x (days after transplanting); c is the lower limit considered as 0; d is the estimated maximum plant height or leaf number or growth index; e is the time taken to reach 50% of final height, leaf number, or growth index; and b is the relative slope around the parameter e .

^c Values are mean ± SEM.

^d The unit of the parameter d is cm, no. plant⁻¹, cm³ for the plant height, leaves plant⁻¹, and growth index, respectively.

^e Growth index = $\pi \times (w/2)^2 \times h$, where w is the width of the plant and h is the plant height.

^f Values presented for d and RMSE are divided by 10⁵.

EF value ranges between $-\infty$ and 1; values closer to 1 means more accurate predictions.

Results and Discussion

Common waterhemp biotypes from two Nebraska counties responded similarly ($P > 0.05$) to degree and duration of water stress. Treatment-by-experiment interaction was not significant in either study; therefore, data from both biotypes and years were combined.

Degree of Water Stress. Degree of water stress influenced growth and fecundity of common waterhemp. With reduced degrees of water stress, common waterhemp plant height, leaves plant⁻¹, and growth index increased following a log-logistic sigmoid growth function (Figure 1). Maximum plant height (d) estimated by the model was 163 cm when amount of water was equivalent to 100% of pot water content (no water stress), which was reduced to 146 and 115 cm with 75% (light water stress) and 50% (moderate water stress) of pot water content, respectively (Table 1). Compared with

100% of pot water content (no water stress), estimated maximum plant height was reduced by 43 and 71% when amount of water added to the plants were 25% (high water stress) and 12.5% (severe water stress) of pot water content, respectively. Chauhan (2013) reported that plant height of itchgrass, another C₄ weed species, was reduced by 49 and 63% at 25 and 12.5% of pot water content, respectively. Common waterhemp plants took 31 d (e) to reach 50% of the estimated maximum plant height with the treatments of 100% (no water stress) and 75% (light water stress) of pot water content as compared with 12 d for 12.5% of pot water content (severe water stress) (Table 1). This is because the plants under severe water stress did not survive 30 DAT, resulting in a flat curve for plant height (Figure 1a).

The highest number of leaves (231 leaves plant⁻¹) was recorded with 100% of pot water content treatment (no water stress), whereas increasing level of water stress decreased the number of leaves plant⁻¹ (Figure 1b). Compared to 100% of pot water content (no water stress), estimated maximum number of leaves plant⁻¹ were reduced by > 30% when

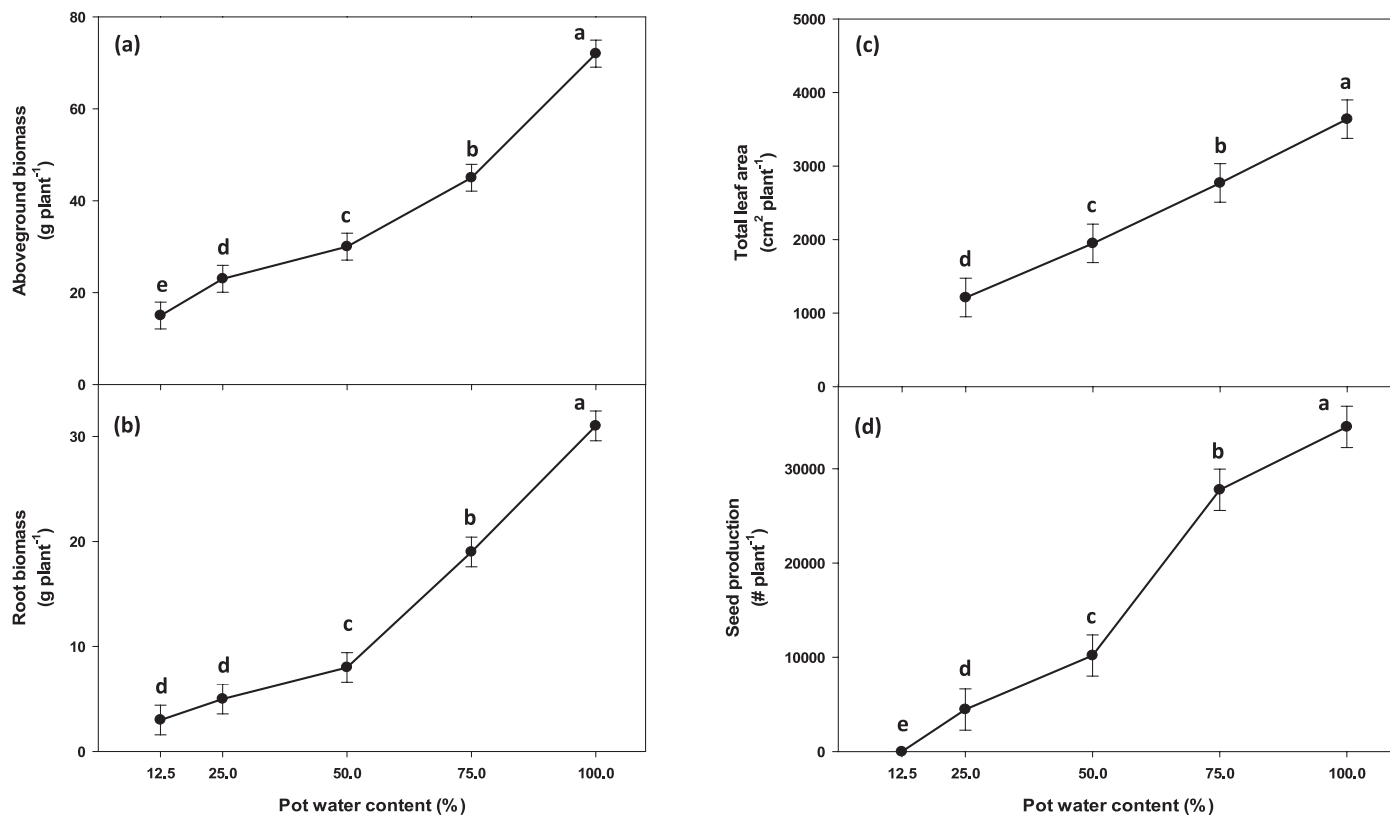


Figure 2. Effect of degree of water stress on (a) aboveground biomass, (b) root biomass, (c) total leaf area, and (d) seed production of common waterhemp in a greenhouse study conducted in Nebraska.

applied water was equivalent to $\leq 50\%$ of pot water content (moderate to severe water stress) (Table 1). Maximum numbers of leaves estimated by the model were 105 and 47 leaves plant^{-1} with 25% (high water stress) and 12.5% (severe water stress) of pot water content, respectively. The model estimated that 50% of maximum leaves were reached within 26 to 30 d at 25 to 100% of pot water content (high to no water stress) (Table 1).

Growth index is the cumulative effect of plant width and plant height (Equation 2). Therefore, the growth index followed a similar pattern as plant height under water stress conditions. Model-estimated highest growth index ($4.4 \times 10^5 \text{ cm}^3 \text{ plant}^{-1}$) was observed with the 100% of pot water content (no water stress) treatment, whereas comparatively lower growth index ($\leq 3.1 \times 10^5 \text{ cm}^3 \text{ plant}^{-1}$) was observed when water was added at 75% of pot water content (light water stress) or less (Figure 1c; Table 1). Compared to the treatment of 100% of pot water content, growth index was reduced by 30, 66, 86, and 93% when amount of applied water was equivalent to 75% (light water stress), 50% (moderate water stress), 25% (high water stress), and 12.5% (severe water stress) of pot water content. Based on the estimation, 50% of maximum growth index was achieved at ≥ 29

DAT under 25 to 100% of pot water content (high to no water stress) treatments compared to only 19 d under 12.5% of pot water content (severe water stress) (Table 1).

The RMSE values for plant height and number of leaves plant^{-1} ranged from 7.2 to 31.5 (Table 1), indicating a good fit of the model. Roman et al. (2000) reported an RMSE value of 6.5 to 37.1 during validation of a model to predict emergence of common lambsquarters (*Chenopodium album* L.). Most of the EF values for plant height and leaves plant^{-1} ranged from 0.75 to 0.90 (Table 1); indicating the good fit of the model. The EF value for leaves plant^{-1} under 12.5% of pot water content (severe water stress) was lower (0.32) compared to other treatments because of more variation in the data set and flat curves after 30 DAT. The RMSE values for growth index were higher, ranging from 0.1×10^5 to 0.9×10^5 (Table 1). Growth index is an interaction between plant height and plant width and it may lead to the higher values and variations for the observed data set. However EF values, ranging from 0.43 to 0.87, showed the goodness of fit for the predicted model.

The highest aboveground biomass (72 g plant^{-1}) and root biomass (31 g plant^{-1}) were recorded in plants receiving 100% of pot water content (no

water stress); whereas biomass production was reduced with increasing degrees of water stress (Figures 2a and 2b), similar to responses reported for itchgrass and junglerice (Chauhan 2013, Chauhan and Johnson 2010). Compared with 100% of pot water content (no water stress), the aboveground biomass was reduced by 68 and 79% with 25% (high water stress) and 12.5% (severe water stress) of pot water content, respectively. A higher root : shoot ratio (≥ 0.42) was observed with the treatments of 100% (no water stress) and 75% (light water stress) of pot water content, whereas the root : shoot ratio was the lowest (≤ 0.22) with high (25% of pot water content) to severe (12.5% of pot water content) water stress (data not shown). Similarly, Moore and Franklin (2011) reported that root : shoot ratio of Palmer amaranth (*Amaranthus palmeri* S. Wats.), a closely related species of common waterhemp, was reduced under water stress compared with drained and flooded conditions. Total leaf area is dependent on plant growth and the total number of leaves plant⁻¹. At 90 DAT, common waterhemp plants produced the highest leaf area (3,638 cm² plant⁻¹) with 100% of pot water content (no water stress) treatment, whereas total leaf area decreased with increasing water stress (Figure 2c). Compared to the 100% of pot water content (no water stress) treatment, total leaf area was reduced by 46 and 67% at 50% (moderate water stress) and 25% (severe water stress) of pot water content, respectively. Plants under severe water stress treatment did not survive after 30 DAT.

Seed production declined with increasing degree of water stress (Figure 2d). The highest number of seeds (34,450 seeds plant⁻¹) was produced with 100% of pot water content (no water stress) as compared with 27,775 seeds plant⁻¹ at 75% of pot water content (light water stress). Surprisingly, plants receiving moderate and high water stress were able to produce 10,194 and 4,469 seeds plant⁻¹, respectively. Reduction in seed production has been reported with increased water stress in itchgrass and junglerice (Chauhan 2013, Chauhan and Johnson 2010). As expected, no seeds were produced by the plants under severe water stress, since these plants did not survive more than 30 DAT.

Duration of Water Stress. Duration of water stress had a significant effect on growth and fecundity of common waterhemp. Similar to the degree of water stress study, a sigmoidal log-logistic response was observed for common waterhemp plant height, leaves plant⁻¹, and growth index under different

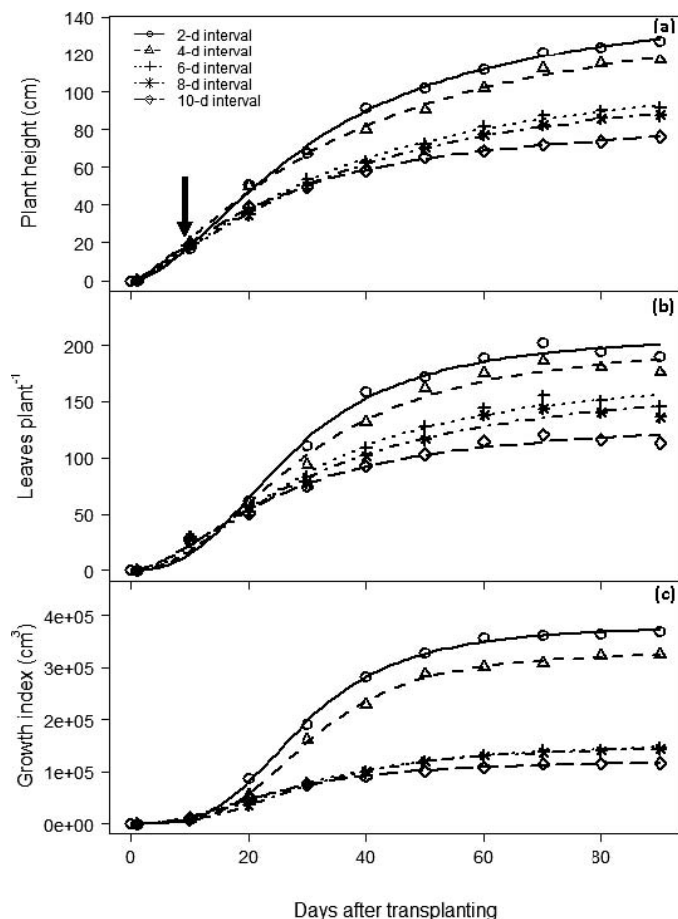


Figure 3. Effect of duration of water stress on (a) height, (b) leaves plant⁻¹, and (c) growth index of common waterhemp in a greenhouse study conducted in Nebraska. The arrow at 10 d after transplanting (DAT) denotes the first day when water stress treatments were imposed.

intervals of water stress (Figure 3). The estimated maximum plant height (d) from the model was similar (150 cm) for 2- and 4-d intervals of water stress, whereas it was reduced to 118 cm for the 6-d water stress interval (Table 2). Similar responses were observed by Chauhan (2013) in itchgrass, where 1- and 3-d intervals of water stress resulted in similar estimated maximum plant height, and increasing duration of water stress decreased plant height. Compared to the 2-d water stress interval, maximum plant height was reduced by 25 and 41% at 8- and 10-d intervals of water stress, respectively. Based on the estimation, common waterhemp plants required 32 d (e) to reach 50% of the maximum plant height at a 2-d water stress interval and with 4-, 6-, and 8-d intervals, they required 35 d (Figure 3a). The number of leaves was highest (210 leaves plant⁻¹) at the 2-d interval of water stress, whereas at 4- and 6-d intervals, plants produced 204 and 174 leaves plant⁻¹, respectively (Table 2). In contrast with the 2-d water stress interval, estimated maximum leaves

Table 2. Parameter estimates and the goodness of fit (RMSE, and EF)^a of the four-parameter log-logistic function^b fitted to common waterhemp plant height, leaves plant⁻¹, and growth index under different duration of water stress treatments in a greenhouse experiment conducted in Nebraska.

Duration of water stress	$d^{c,d}$	e (days) ^c	b^c	RMSE	EF
— Plant height —					
2-d interval	150 ± 9	32 ± 3	-1.7 ± 0.2	11.6	0.93
4-d interval	150 ± 13	35 ± 5	-1.4 ± 0.2	11.1	0.92
6-d interval	118 ± 10	35 ± 5	-1.4 ± 0.2	7.9	0.94
8-d interval	113 ± 7	35 ± 3	-1.4 ± 0.1	5.4	0.97
10-d interval	88 ± 9	24 ± 4	-1.4 ± 0.3	10.9	0.83
— Leaves plant ⁻¹ —					
2-d interval	210 ± 9	27 ± 1	-2.6 ± 0.4	27.3	0.87
4-d interval	204 ± 10	30 ± 2	-2.2 ± 0.3	23.1	0.89
6-d interval	174 ± 14	30 ± 3	-1.9 ± 0.3	25.3	0.81
8-d interval	170 ± 19	31 ± 5	-1.7 ± 0.4	26.9	0.77
10-d interval	133 ± 9	25 ± 3	-1.7 ± 0.3	17.1	0.84
— Growth index ^{e,f} —					
2-d interval	3.8 ± 0.2	29 ± 2	-3.4 ± 0.8	0.9	0.72
4-d interval	3.3 ± 0.2	30 ± 2	-3.5 ± 0.8	0.8	0.73
6-d interval	1.6 ± 0.1	31 ± 2	-2.3 ± 0.3	0.2	0.86
8-d interval	1.5 ± 0.1	31 ± 4	-2.7 ± 0.8	0.4	0.64
10-d interval	1.2 ± 0.1	24 ± 4	-2.3 ± 0.7	0.4	0.56

^a Abbreviations: RMSE, root mean square error; EF, modelling efficiency coefficient.

^b $Y = c + \{d - c / 1 + \exp[b(\log x - \log e)]\}$, where Y is the plant height, leaves plant⁻¹, or growth index at time x (days after transplanting); c is the lower limit considered as 0; d is the estimated maximum plant height or leaf number or growth index, e is the time taken to reach 50% of final height, leaf number, or growth index; and b is the relative slope around the parameter e .

^c Values are mean ± SEM.

^d The unit of the parameter d is cm, no. plant⁻¹, cm³ for the plant height, leaves plant⁻¹, and growth index, respectively.

^e Growth index = $\pi \times (w/2)^2 \times h$, where w is the width of the plant and h is the plant height.

^f Values presented for d and RMSE are divided by 10⁵.

plant⁻¹ were reduced by 19 and 37% at 8- and 10-d intervals of water stress (Figure 3b). The highest growth index (3.8×10^5 cm³ plant⁻¹) was observed at 2-d interval of water stress compared to other treatments, requiring 29 d to reach the 50% of the estimated maximum growth index (Figure 3c). Compared to the 2-d interval of water stress, growth index was reduced by 13, 58, 61, and 68% at 4-, 6-, 8-, and 10-d intervals, respectively.

The RMSE and EF values for the plant height and leaves plant⁻¹ ranged from 5.4 to 27.3, and 0.77 to 0.97 (Table 2), respectively; indicating the good fit of the model. Werle et al. (2014a) predicted the emergence of winter annual weeds and reported the RMSE and EF ranging from 13.4 to 23.1 and 0.63 to 0.85, respectively. The RMSE values for growth index were higher, ranging from 0.2×10^5 to 0.9×10^5 (Table 2), whereas EF values ranged from 0.56 to 0.86. The higher RMSE values for growth index could be due to the higher numbers and more variations among the observed data set.

The aboveground and root biomass decreased with increasing duration of water stress (Figures 4a and 4b). Similarly, Chauhan (2013) reported

reduction in itchgrass biomass with increasing durations of water stress. The highest aboveground biomass (59 g plant⁻¹) was recorded at the 2-d interval of water stress, whereas similar trend was observed in the root biomass (30 g plant⁻¹). Compared to the 2-d interval of water stress, aboveground biomass was reduced by 39 and 51% at 8- and 10-d intervals, respectively (Figure 4a). Common waterhemp root biomass was sharply reduced with increasing duration of water stress (Figure 4b), but it was similar at 6- and 8-d intervals of water stress. Root : shoot ratio was highest (0.51) at a 2-d interval of water stress, whereas it was similar (0.39 to 0.44) at 4- to 8-d intervals (data not shown). At 90 DAT, total leaf area was similar at 2-d (3,913 cm² plant⁻¹) and 4-d (3,265 cm² plant⁻¹) intervals of water stress (Figure 4c). Seed production is the most important characteristic of a weed for reproduction and survival, and increasing duration of water stress usually reduces seed production (Chauhan 2013). Common waterhemp seed production was highest (34,176 to 36,549 seeds plant⁻¹) at 2- and 4-d intervals of water stress (Figure 4d), but was reduced by 42, 51, and

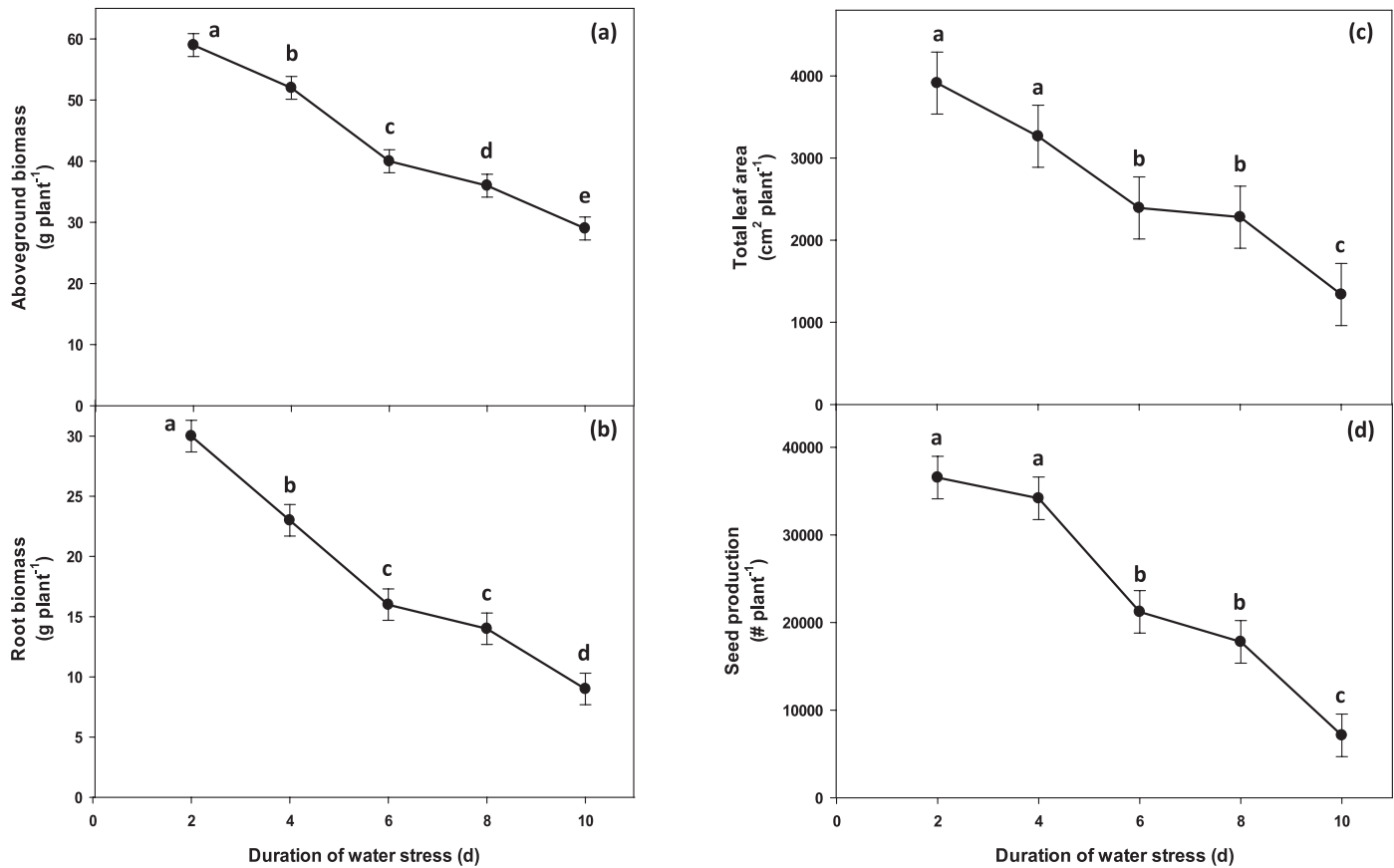


Figure 4. Effect of duration of water stress on (a) aboveground biomass, (b) root biomass, (c) total leaf area, and (d) seed production of common waterhemp in a greenhouse study conducted in Nebraska.

80% at 6-, 8-, and 10-d intervals of water stress, respectively.

This study revealed that water stress can impact growth and seed production of common waterhemp. Similarly, Moran and Showler (2005) reported that water stress can reduce shoot height and fresh weight of Palmer amaranth by 31 and 35%, respectively, when 25% of pot water content water was added at 4-d interval. Reduction in plant root elongation under water stress conditions was reported by Bengough et al. (2011), mainly due to an increase in mechanical impedance in the dry soil. Moreover, Masle and Passioura (1987) and Young et al. (1997) reported that root and shoot growth are correlated and as a result, leaf expansion can be affected by water deficit, supporting the response of common waterhemp to water stress observed in this study. Seed dormancy of certain species plays a key role in developing an effective weed management strategy. Fenner (1991) reported that water stress during seed development can affect the germination of subsequent seeds depending on a species' mechanism of dormancy. In this study, water stress had no effect on germination of common waterhemp seeds (data not shown). Similarly, Chauhan and Johnson (2010)

reported that junglerice seed production was reduced sharply with increasing duration of water stress, but with no effect on seed germination.

Water stress may influence the critical weed-free period for different crop species (Patterson 1995). Weed species that have faster growth rates with high biomass production ability, and preempt the available growth resources, would be considered as a highly competitive weed species over other slow-growing species (Horak and Loughin 2000). Results of this study will provide information about biological attributes of common waterhemp under water stress conditions that can be used to understand and evaluate the effects of environmental stress on the weed-crop interaction by using a mathematical model in the future. This information can also be used for developing climate simulation models to understand the effect of drought on crop and weed species in the future. Additionally, it is known that efficacy of POST herbicides is reduced under water stress situations due to less retention and uptake of herbicides by the target plants (Kudsk and Kristensen 1992). For example, uptake of glyphosate decreased in black nightshade (*Solanum nigrum* L.) when plants were under water stress (Ruiter and

Meinen 1998). Therefore, future research should focus on relative competitiveness of common waterhemp with different crop species and response to POST herbicides under water stress conditions.

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Literature Cited

- Aulakh JS, Jhala AJ (2015) Comparison of glufosinate-based herbicide programs for broad-spectrum weed control in glufosinate-resistant soybean. *Weed Technol* 29:419–430
- Begg JE, Turner NC (1976) Crop water deficits. *Adv Agron* 28:161–217
- Bengough AG, McKenzie BM, Hallett PD, Valentine TA (2011) Root elongation, water stress, and mechanical impedance: a review of limiting stresses and beneficial root tip traits. *J Exp Bot* 62:59–68
- Benjamin JG, Nielsen DC (2006) Water deficit effects on root distribution of soybean, field pea and chickpea. *Field Crops Res* 97:248–253
- Bernards ML, Crespo RJ, Kruger GR, Gaussoin R, Tranel PJ (2012) A waterhemp (*Amaranthus tuberculatus*) population resistant to 2,4-D. *Weed Sci* 60:379–384
- Chahal PS, Jhala AJ (2015) Herbicide programs for control of glyphosate-resistant volunteer corn in glufosinate-resistant soybean. *Weed Technol* 29:431–443
- Chahal PS, Kruger G, Blanco H, Jhala AJ (2014) Efficacy of pre-emergence and post-emergence soybean herbicides for control of glufosinate-, glyphosate-, and imidazolinone-resistant volunteer corn. *J Agric Sci* 6:131–140
- Chauhan BS (2013) Growth response of itchgrass (*Rottboellia cochinchinensis*) to water stress. *Weed Sci* 61:98–103
- Chauhan BS, Johnson DE (2010) Growth and reproduction of junglerice (*Echinochloa colona*) in response to water-stress. *Weed Sci* 58:132–135
- Costea M, Weaver SE, Tardif FJ (2005) The biology of invasive alien plants in Canada. 3. *Amaranthus tuberculatus* (Moq.) Sauer var. *rudis* (Sauer) Costea & Tardif. *Can J Pl Sci* 85:507–522
- Dhir R, Harkess RL (2011) Elevated air temperatures cause foliar bleaching of ivy geranium 'Beach' and 'Butterfly'. *HortScience* 46:411–415
- Fenner M (1991) The effects of the parent environment on seed germinability. *Seed Sci Res* 1:75–84
- Gallagher RS, Cardina J (1998) Phytochrome-mediated *Amaranthus* germination II: development of very low fluence sensitivity. *Weed Sci* 46:53–58
- Ganie ZA, Stratman G, Jhala AJ (2015) Response of selected glyphosate-resistant broadleaved weeds to premix of fluthiacetmethyl and mesotrione (SolsticeTM) applied at two growth stages. *Can J Plant Sci* 95: 861–869
- Hager AG, Wax LM, Stoller EW, Bollero GA (2002) Common waterhemp (*Amaranthus rudis*) interference in soybean. *Weed Sci* 50:607–610
- Hartzler RG, Buhler DD, Stoltenberg DE (1999) Emergence characteristics of four annual weed species. *Weed Sci* 47:578–584
- Hausman NE, Singh S, Tranel PJ, Riechers DE, Kaundun SS, Polge ND, Thomas DA, Hager AG (2011) Resistance to HPPD-inhibiting herbicides in a population of waterhemp (*Amaranthus tuberculatus*) from Illinois, United States. *Pest Manag Sci* 67:258–261
- Heap I (2015) The International Survey of Herbicide Resistant Weeds. Herbicide Resistant Tall Waterhemp Globally. <http://weedsociology.org/Summary/Species.aspx?WeedID=219>. Accessed May 30, 2015
- Horak MJ, Loughin TM (2000) Growth analysis of four *Amaranthus* species. *Weed Sci* 48:347–355
- Irmak S, Haman DZ, Bastug R (2000) Determination of crop water stress index for irrigation timing and yield estimation of corn. *Agron J* 92:1221–1227
- Irmak S, Haman DZ, Irmak A, Jones JW, Campbell KL, Crisman TL (2004) Measurement and analysis of growth and stress parameters of *Viburnum odoratissimum* (Ker-gawl) grown in a multi-plot box system. *HortScience* 39:1445–1455
- Jhala AJ (2015) Herbicide-resistant weeds. Pages 18–19 in Knezevic SZ, Jhala AJ, Klein RN, Kruger GR, Reicher ZJ, Wilson RG, Shea PJ, Ogg CL, eds. 2015 Guide for Weed Management in Nebraska with Insecticide and Fungicide Information. Lincoln, NE: University of Nebraska–Lincoln Extension
- Kaur S, Sandell LD, Lindquist JL, Jhala AJ (2014) Glyphosate-resistant giant ragweed (*Ambrosia trifida*) control in glufosinate-resistant soybean. *Weed Technol* 28:569–577
- Knezevic SZ, Streibig JC, Ritz C (2007) Utilizing R software package for dose-response studies: the concept and data analysis. *Weed Technol* 21:840–848
- Kudsk P, Kristensen JL (1992) Effect of environmental factors on herbicide performance. Pages 173–186 in Proceedings of the First International Weed Control Congress. Melbourne, Australia: Weed Science Society of Victoria Inc
- Legleiter TR, Bradley KW (2008) Glyphosate and multiple herbicide resistance in common waterhemp (*Amaranthus rudis*) populations from Missouri. *Weed Sci* 56:582–587
- Liu F, Stützel H (2002a) Leaf expansion, stomatal conductance, and transpiration of vegetable amaranth (*Amaranthus* sp.) in response to soil drying. *J Am Soc Hortic Sci* 127:878–883
- Liu F, Stützel H (2002b) Leaf water relations of vegetable amaranth (*Amaranthus* spp.) in response to soil drying. *Eur J Agron* 16:137–150
- Liu J, Davis AS, Tranel PJ (2012) Pollen biology and dispersal dynamics in waterhemp (*Amaranthus tuberculatus*). *Weed Sci* 60:416–422
- Long SP (1999) Ecology of C₄ photosynthesis-environmental responses. Pages 215–249 in Sage RF and Monson RK, eds. C₄ Plant Biology. San Diego, CA: Academic Press
- Lovelli S, Perniola M, Ferrara A, Amato M, Tommaso TD (2010) Photosynthetic response to water stress of pigweed (*Amaranthus retroflexus*) in a Southern-Mediterranean area. *Weed Sci* 58:126–131
- Masle J, Passioura JB (1987) The effect of soil strength on the growth of young wheat plants. *Aust J Plant Physiol* 14:643–656
- Mayer DG, Butler DG (1993) Statistical validation. *Ecol Model* 68:21–32

- Moore JE, Franklin SB (2011) Understanding the relative roles of disturbance and species interactions in shaping Mississippi River island plant communities. *Community Ecol* 12:108–116
- Moran PJ, Showler AT (2005) Plant responses to water deficit and shade stresses in pigweed and their influence on feeding and oviposition by the beet armyworm (Lepidoptera: Noctuidae). *Environ Entomol* 34:929–937
- Nordby D, Hartzler B, Bradley K (2007) Biology and Management of Waterhemp. Purdue Extension. GWC-13. 3 p
- Patterson DT (1995) Effects of environmental stress on weed/crop interactions. *Weed Sci* 43:483–490
- Roman ES, Murphy SD, Swanton CJ (2000) Simulation of *Chenopodium album* seedling emergence. *Weed Sci* 48:217–224
- Ruiter HD, Meinen E (1998) Influence of water stress and surfactant on the efficacy, absorption, and translocation of glyphosate. *Weed Sci* 46:289–296
- Saranghi D, Sandell LD, Knezevic SZ, Aulakh JS, Lindquist JL, Irmak S, Jhala AJ (2015) Confirmation and control of glyphosate-resistant common waterhemp (*Amaranthus rudis*) in Nebraska. *Weed Technol* 29:82–92
- Shoup DE, Al-Khatib K, Peterson DE (2003) Common waterhemp (*Amaranthus rudis*) resistance to protoporphyrinogen oxidase-inhibiting herbicides. *Weed Sci* 51:145–150
- Spiess AN, Neumeyer N (2010) An evaluation of R2 as an inadequate measure for nonlinear models in pharmacological and biochemical research: a Monte Carlo approach. *BMC Pharmacol* 10:6
- Steadman KJ, Ellery AJ, Chapman R, Moore A, Turner NC (2004) Maturation temperature and rainfall influence seed dormancy characteristics of annual ryegrass (*Lolium rigidum*). *Aust J Agric Res* 55:1047–1057
- Steckel LE, Sprague CL (2004) Common waterhemp (*Amaranthus rudis*) interference in corn. *Weed Sci* 52:359–364
- Steckel LE, Sprague CL, Hager AG, Simmons FW, Bollero GA (2003) Effects of shading on common waterhemp (*Amaranthus rudis*) growth and development. *Weed Sci* 51:898–903
- Steckel LE, Sprague CL, Stoller EW, Wax LM, Simmons FW (2007) Tillage, cropping system, and soil depth effects on common waterhemp (*Amaranthus rudis*) seed-bank persistence. *Weed Sci* 55:235–239
- Waselkov KE, Olsen KM (2014) Population genetics and origin of the native North American agricultural weed waterhemp (*Amaranthus tuberculatus*; Amaranthaceae). *Am J Bot* 101:1726–1736
- Webster TM, Grey TL (2008) Growth and reproduction of Benghal dayflower (*Commelina benghalensis*) in response to drought stress. *Weed Sci* 56:561–566
- Werle R, Bernards ML, Arkebauer TJ, Lindquist JL (2014a) Environmental triggers of winter annual weed emergence in the midwestern United States. *Weed Sci* 62:83–96
- Werle R, Sandell LD, Buhler DD, Hartzler RG, Lindquist JL (2014b) Predicting emergence of 23 summer annual weed species. *Weed Sci* 62:267–279
- Werle R, Schmidt JJ, Laborde J, Tran A, Creech CF, Lindquist JL (2014c) Shattercane × ALS-tolerant sorghum F1 hybrid and shattercane interference in ALS-tolerant sorghum. *J Agric Sci* 6:159–165
- Wu D, Qu JJ, Hao X, Xiong J (2013) The 2012 agricultural drought assessment in Nebraska using MODIS satellite data. Pages 170–175 in *Proceedings of 2nd International Conference on Agro-Geoinformatics*. Fairfax, VA: Center for Spatial Information Science and Systems
- Young IM, Montagu K, Conroy J, Bengough AG (1997) Mechanical impedance of root growth directly reduces leaf elongation rates of cereals. *New Phytol* 135:613–619

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