



Running from depression: the antidepressant-like potential of prenatal and pre-pubertal exercise in adolescent FSL rats exposed to an early-life stressor

Original Article

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Abstract

Objective: We aimed to answer the questions of whether early-life (perinatal and/or juvenile) exercise can induce antidepressant-like effects in a validated rodent model of depression, and whether such early-life intervention could prevent or reverse the adverse effects of early-life stress in their offspring. **Methods:** Male and female Flinders sensitive line rats born to a dam that exercised during gestation, or not, were either maternally separated between PND02 and 16 and weaned on PND17 or not. Half of these animals then underwent a fourteen-day low-intensity exercise regimen from PND22. Baseline depressive-like behaviour was assessed on PND21 and then reassessed on PND36, whereafter hippocampal monoamine levels, redox state markers and metabolic markers relevant to mitochondrial function were measured. **Results:** Pre-pubertal exercise was identified as the largest contributing factor to the observed effects, where it decreased immobility time in the FST by 6%, increased time spent in the open arms of the EPM by 9%. Hippocampal serotonin and norepinephrine levels were also increased by 35% and 26%, respectively, whilst nicotinic acid was significantly decreased. **Conclusion:** These findings suggest that pre-pubertal low-intensity exercise induces beneficial biological alterations that could translate into antidepressant behaviour in genetically susceptible individuals.

Significant outcomes

- Prenatal exercise may alter coping behaviours in adolescent Flinders sensitive line (FSL) rats offspring.
- Prenatal exercise has long-term beneficial effects on hippocampal redox state.
- Pre-pubertal low-intensity exercise reduces depressive-like behaviour in adolescent FSL rats.
- Low intensity exercise alters hippocampal redox state, pointing to mitochondrial involvement.

Limitations

- The metabolic profile sample collection method, specifically the use of the buffer, may have influenced the results and therefore our findings require confirmation and validation.
- We did not measure any neuro- and biochemical markers on PND21, therefore the behavioural findings of the TST require confirmation.
- Pregnant dams exercised for 13 ± 5 days, which may have influenced the results of the prenatal exercise group.
- No stress-related markers were measured in animals born to an exercised dam, which would elaborate on the behavioural interpretations of these pups.

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Introduction

The human brain accounts for only 2% of a person's weight yet consumes 20% of total glucose and oxygen (Rolfe & Brown, 1997; Manji *et al.*, 2012; Pei & Wallace, 2018), explaining the

particular high energy demand of brain neurons (80–90% of the total brain demand). In fact, the human brain uses ~ 5.7 kg of ATP/day at rest, of which the majority is utilised by cortical neurons (Zhu *et al.*, 2012). These energy demands are significantly increased by stress (Bryan, 1990) and sets off structural and functional changes in surrounding cells to match the required behaviour in response to stress (Picard *et al.*, 2014). Early-life trauma has been shown to negatively affect central and peripheral mitochondrial function (Hoffmann & Spengler, 2018; Ruigrok *et al.*, 2021), thereby adversely altering stress-response pathways (Zitkovsky *et al.*, 2021), and contributing to the underlying pathophysiology of depression (Sharma & Akundi, 2019; van Rensburg *et al.*, 2022) and other psychiatric conditions (van Rensburg *et al.*, 2022). Clinical findings have also reported dysfunctional mitochondria in depressed patients (reviewed by Caruso *et al.* (2019)) and the diverse effects of approved psychotropic drugs on mitochondrial function (Emmerzaal *et al.*, 2021).

With this considered, mitochondrial (dys)function may be a promising target to treat depression (Allen *et al.*, 2018; Sharma & Akundi, 2019; Wu *et al.*, 2019) and is of particular importance and value in vulnerable populations, such as pregnant women and pre-pubertal children, in which the currently approved treatment options are limited to selective serotonin reuptake inhibitors (Kimmel *et al.*, 2018; Viswanathan *et al.*, 2020). Moreover, the use of these (and other) antidepressants during these vulnerable developmental periods is often questioned (Bérard *et al.*, 2017; Molenaar *et al.*, 2018; Hengartner, 2020) because of the uncertainty surrounding their long-term effects and overall safety profiles. In fact, all antidepressants require a “black box” warning for increased suicidal behaviour in juvenile patients (U.S. Food & Drug Administration, 2004). It is for these reasons that alternative treatment options, especially in these vulnerable populations, are necessary. One promising non-pharmacological treatment option is exercise. The antidepressant effects of exercise have been widely established (Carter *et al.*, 2016; Kandola *et al.*, 2019), yet the exact mechanisms through which it exerts its antidepressant effects remain unknown (Schuch *et al.*, 2016). Numerous mechanisms have been proposed (Kandola *et al.*, 2019; de Oliveira *et al.*, 2022) and include (but are not limited to) increased monoamine neurotransmission (Lin & Kuo, 2013) and neuroplasticity (El-Sayes *et al.*, 2019) and decreased inflammation and oxidative stress (Eyre & Baune, 2012), all of which have been implicated in the pathophysiology of depression and as alluded to earlier, linked to mitochondrial function (Allen *et al.*, 2018; Sharma & Akundi, 2019; van Rensburg *et al.*, 2022). Additionally, exercise also induces bio-energetic enhancing effects (i.e., improved mitochondrial functioning (Aguilar *et al.*, 2014; Wu *et al.*, 2019)). In contrast, physical inactivity has been linked to adverse health outcomes, including depression (Kandola *et al.*, 2019) and metabolic disorders, such as diabetes and cardiovascular diseases (Katzmarzyk *et al.*, 2022), which are often co-morbid with depression. Exercise during the prenatal period is known to be safe and beneficial to the mother and offspring (Davenport, *et al.*, 2018; Davenport, *et al.*, 2018; Moyer *et al.*, 2016). Also, increased physical activity during juvenile development is inversely associated with depressive symptoms (Biddle *et al.*, 2019; Dale *et al.*, 2019). Therefore, physical exercise may not only be a promising treatment option for depression but also provide additional health benefits, and even reduce the risk to develop depression in children with a genetic predisposition to develop depression.

Early-life development is a sensitive period, characterised by extensive growth and plasticity, that makes the developing brain sensitive to external influences (Heim *et al.*, 2010; Scattolin *et al.*, 2022). For instance, early-life adversity, such as neglect or abuse, can have detrimental developmental effects with long-lasting consequences (Andersen & Teicher, 2008; Obi *et al.*, 2019) to such an extent that one-third of mental disorders, including depression and anxiety can be ascribed to early-life adversity (Kessler *et al.*, 2010; McLaughlin *et al.*, 2019). In the USA, it is estimated that 4.4% and 9.4% of children aged 3–17 years suffer from depression and anxiety, respectively, with 33% of children who suffer from anxiety also experiencing depressive symptoms (Centers for Disease Control and Prevention, 2021). Importantly, neurodevelopment does not only occur during childhood and the adolescent period but begins as early as the embryonic stage (Scattolin *et al.*, 2022) and is therefore also influenced by the perinatal environment (Schoorjans & Kurrasch, 2013). To this extent, it is worth noting that prenatal stress (including maternal depression and family adversity) is not only associated with offspring depression later in life but is also a significant risk factor for childhood trauma (Liu *et al.*, 2022). It is for this reason that maternal health during the perinatal and post-partum period is significant for a developing child. Importantly, maternal depression during the perinatal and or post-partum period significantly increases the offspring's risk to develop depression later in life by as much as 70% (Tirumalaraju *et al.*, 2020) and therefore, by simply being born into a family with a history of depression, increases the risk for developing depression (Führer *et al.*, 2009; Thompson *et al.*, 2018; Tirumalaraju *et al.*, 2020). Still, due to ethical and practical reasons, whether prenatal exercise can protect or even reverse the adverse effects of early-life adversity, remains unexplored. Therefore, to mimic this increased risk of developing depression both through a combination of genetic and environmental influences, the current study applied an early-life stressor (i.e. maternal separation and early weaning; MSEW) to an approved genetic rodent model of depression (i.e. Flinders sensitive line (FSL) rat) (Overstreet & Wegener, 2013), as this strain has been reported to already display depressive-like behaviour during juvenile development (Malkesman & Weller, 2009; Whitney *et al.*, 2023).

Considering the above, we hypothesised that exercise during the prenatal and pre-pubertal periods would induce antidepressant-like behaviour by improving hippocampal mitochondrial function, monoaminergic neurotransmission and redox state in adolescent FSL rats (representing a juvenile patient, predisposed to develop depression). Moreover, we hypothesised that these effects would prevent and/reverse the depressogenic effects of the early-life stressor.

Materials and methods

Study layout

A similar study layout, except for the exercise interventions, was used as before to build on our previous findings (Whitney *et al.*, 2023). Briefly, pregnant FSL dams were either subjected to a chronic low-intensity exercised regimen or not. Next, the offspring of these dams were then further divided into MSEW and non-MSEW groups. On PND21, animals were subjected to the open field (OFT) and tail suspension tests (TST) to determine early-life depressive-like behaviour. Next, 50% of the animals underwent a 14-day low-intensity exercise regimen, whereafter all animals were subjected to the OFT and forced swim tests (FST) on PND36,

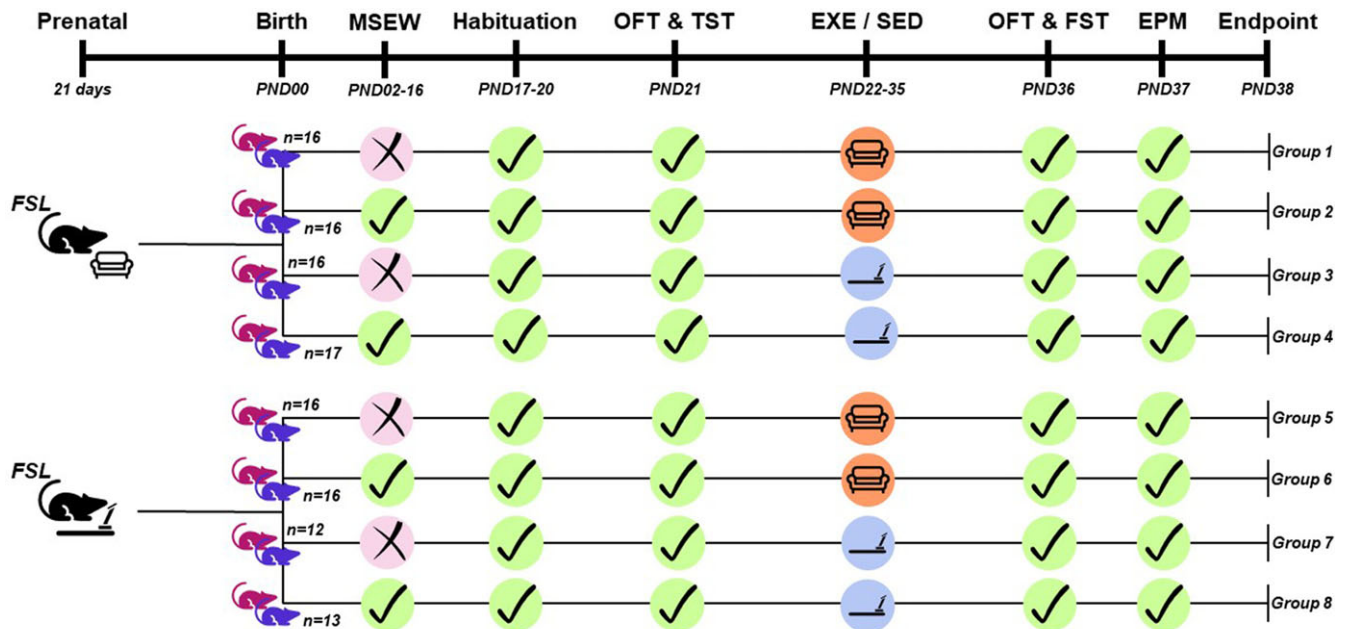


Figure 1. Graphical summary of the study layout. Pregnant FSL dams were either subjected to a prenatal sedentary or low-intensity exercise regimen. Animals were either subjected to early-life stress (MSEW) between PND02 and 17 or not. Early-life behavioural testing took place on PND21 to determine the effects of prenatal exercise. To investigate the bio-behavioural effects of juvenile exercise (with and without prenatal exercise), a 14-day low-intensity exercise (or sedentary) regimen was introduced on PND22, whereafter behavioural testing took place on PND36 and 37, followed by decapitation and brain dissection on PND38. Tissue was frozen at -80°C until neurochemical analyses were performed. Couch icon: sedentary group. Treadmill icon: exercise group. Pink rat icon: female rats. Purple rat icon: male rats. EPM, elevated plus maze; EXE, low-intensity exercise; FRL, flinders resistant line; FSL, flinders sensitive line; FST, forced swim test; MSEW, maternal separation with early weaning; OFT, open field test; PND, postnatal day; SED, sedentary; TST, tail suspension test.

followed by the elevated plus maze test on PND37. The sequence of the behavioural analysis, specifically in terms of performing the forced swim test before the elevated plus maze test, has been carried out by others (Neumann *et al.*, 2011; Rea *et al.*, 2014; Bay-Richter *et al.*, 2019). Moreover, to minimise the risk for potential habitual learning, depressive-like behaviour was analysed by two different behavioural tests (i.e. TST and FST) at different time points. To ensure normal initial foraging and activity of nocturnal animals, testing only commenced one hour after the start of the dark cycle. Tests were carefully spaced to allow 30 min between each test for animals to habituate to the environment. Automated tracking software (Ethovision XT14 Software; Noldus information Technology BV, Wageningen, NLD) was used to track behaviour in the OFT and EPM. TST and FST behaviour was manually scored by a researcher blind to the experimental group details, from recordings of the behavioural tests, recorded with a camera mounted in front of the test apparatus.

Animals and justification of group sizes

Building on previous findings, where we investigated the effects of MSEW on FSL rats (Whitney *et al.*, 2023), male and female FSL ($n = 122$) rats were divided into eight experimental groups, consisting of sixteen rats (50:50 females:males) per group (Fig. 1). Male and female rats were grouped together as sex differences is not expected in pre-pubertal animals. Still, results were visually inspected to identify any obvious sex differences. Smaller groups were, however, sometimes employed due to the exclusion of non-runners and/or when the breeding programme failed to supply the adequate number of animals. Group sizes were calculated with a predicted effect size F of 0.403 ($\eta^2_p = 0.14$), α error (0.05) and 80% power. Rats were group housed (3–4 rats/cage),

according to sex, with corncob bedding changed weekly and the environmental temperatures maintained at $22 \pm 1^{\circ}\text{C}$ in a relative humidity of $55 \pm 10\%$. A 12 h light/dark cycle was followed with food and water provided *ad libitum*.

Maternal separation and early weaning

As described by George and colleagues (2010), pups in the MSEW groups were left in their home cages, while the dams were relocated to new, clean cages with *ad libitum* access to food and water for 3 h per day from PND02 to 16. In contrast, the non-MSEW pups were left, undisturbed with the dam, with all pups remaining with their littermates. Finally, MSEW animals were also weaned on PND17, opposed to the standard PND21 when the non-MSEW pups were weaned.

Exercise regimen

Forced exercise was implemented using a custom-built, programmable treadmill, comprising of a single running belt six shocking grids installed at the back of the treadmill. The shocking grid delivered an electrical shock of 1 mA (3 Hz), and the shocking intensity was selected to be uncomfortable but not painful or harmful. Sedentary animals were removed from their home cages and placed on a mock (still standing) treadmill. All exercise interventions were performed during the animal's active dark cycle (i.e. from 18:00 in the evening until latest 03:00 in the morning, depending on animal numbers). All animals were familiarised to the treadmill using a 10-min routine to reduce injury risk and identify any 'non-runners'. As described previously (Kregel *et al.*, 2006), animals were classified as 'non-runners' when they were unable to keep up with the speed of the treadmill (i.e. shocked three

Table 1. Summarised protocol for prenatal and juvenile familiarisation and exercise. Adapted from Aksu *et al.*, 2012 and Seo *et al.*, 2013

Intervention period	Protocol
Prenatal exercise	
<i>Familiarisation</i>	5m/min for 10 min/day for 7 days
<i>Pregnancy</i>	Exercise group: 30 min/ day at 2m/min for 5 min followed by 5m/min for 5 min and then 8m/min for 20 min 7days a week until birth
Juvenile exercise	
<i>Familiarisation</i>	All pups (<i>regardless of experimental group</i>) from PND 17-20 were subjected to a daily 10-min routine (<i>except for PND 19 where it is a 9 min routine</i>) with gradual increases in treadmill speed. PND 17: 2m/min for 5 min followed by gradual 0.1m/min increases every 1 min until a final speed of 2.5m/min in the last min. PND 18: 2m/min for 3 min followed by 2.5m/min for 2 min. Then a 1 min rest period. 2.5m/min for 1 min followed by 3m/min for 3 min. PND 19: 2m/min for 3 min followed by 3m/min for 3 min with an increase to 4m/min for 3 min and a final speed of 4.5m/min for 1 min. PND 20: 2.5m/min 3 min, then 3.5m/min for 3 min followed by 4.5m/min for 3 min.
<i>PND22-PND35</i>	Exercise group: 30 min/day at 2m/min for 5 min followed by 5m/min for 5 min and then 12m/min 7 days a week for 14 days. Sedentary group: placed in a mock treadmill for 30 min/day for 7 days a week for 14 days

times within 1 min) during the familiarisation period. Pups identified as 'non-runners' were used as control rats; however, those identified as 'non-runners' during the exercise intervention were removed from the study. We observed a small number of rats (< 10%) that displayed 'non-runner' behaviour. Pregnant FSL dams allocated to the exercised group were familiarised with the treadmill two days after being paired to ensure that dams exercise for the full term of their pregnancy. The protocol approach for the familiarisation and low-intensity exercise differed between prenatal exercise and juvenile exercise and is therefore summarised in Table 1.

Behavioural analyses

Open field test

The OFT was used to measure general locomotor activity, and consisted of a 1 m² test arena, surrounded by opaque black, vertical walls. As previously (Steyn *et al.*, 2020) described, each rat, on the day of testing, was placed in the centre of the arena and allowed to freely explore the arena for 5 min under red light. Total distance moved was interpreted as a measure of general activity.

Tail suspension test

In the current study, the TST was used to screen baseline juvenile depressive-like behaviour on PND21. As before (Castagné *et al.*, 2010; Cryan, *et al.*, 2005), on the day of testing, each rat was suspended by the tail with adhesive tape, positioned three-quarters of the distance from the base of the tail from a suspension hook for 6 min. To avoid injury, the suspension hook went through the

adhesive tape as close as possible to the tail to ensure the animal hangs with its tail in a straight line (Castagné *et al.*, 2010). The total time spent immobile was recorded and interpreted as an indication of depressive-like behaviour.

Forced swim test

The FST was performed on PND36, as previously described in our laboratories (Brand & Harvey, 2017; Steyn *et al.*, 2020), without a pre-conditioning swim trial, 24 hr prior to the testing trial (Overstreet *et al.*, 2005; Overstreet & Wegener, 2013). Briefly, during the dark cycle, animals were placed in an inescapable Perspex[®] cylinder filled with 30 cm of water at a temperature of 25 ± 1°C for 6 min. Behaviour was scored manually by an experimenter blind to the experimental group, with the first minute of the test ignored (Roets *et al.*, 2023). Behaviour scored included immobility (floating with no active movements made, except those necessary to keep the rat's head above water), swimming (horizontal movements throughout the cylinder that included crossing into another quadrant) and struggling (upward-directed movements of the forepaws along the inside of the swim cylinder) (Cryan *et al.*, 2002; Cryan, *et al.*, 2005). Increased immobility was considered an indication of depressive-like behaviour.

Elevated plus maze

The EPM is plus shaped Perspex maze that consists of two closed and two open arms, elevated approximately 50 cm above the floor with a 1 cm transparent Plexiglas border to prevent animals from falling. As described previously (Regenass *et al.*, 2018), rats were placed in the centre zone of the maze, facing the open arm opposite the investigator, and allowed to freely explore the maze for 5 min under red light. Increased time spent in the closed arms was interpreted as anxiety-like behaviour, with entrance into an arm considered when the centre point, as defined by the automated scoring program, entered the arm.

Bio-analyses

Tissue collection and storage

Animals were euthanised by decapitation on PND38, whereafter brain and heart samples were harvested and weighed. Following the decapitation, right and left hippocampi were dissected on an ice-cooled dissection slab and stored separately. The right hippocampi were used for neurochemical analysis via LC-MS and snap-frozen in liquid nitrogen and stored at -80°C. The left hippocampi were removed and immediately placed into an isolation buffer (mannitol 200 mM, sucrose 50 mM, potassium phosphate 5 mM, EGTA 1 mM, 3-(N-morpholino)propanesulfonic acid 5 mM and bovine serum albumin 0.10% pH 7.2) (Kim *et al.*, 2016), whereafter it was also stored at -80°C.

Quantitative analyses of hippocampal monoamines, GSH and GSSG

Quantitative monoaminergic, GSH (glutathione) and GSSG (glutathione disulphide) concentrations were analysed via LC-MS, as before (Whitney *et al.*, 2023). A detailed description of the method is available as Supplementary data.

Metabolic profiling via GC-TOF-MS analysis

Untargeted gas chromatography time-of-flight mass spectrometry (GC-TOF-MS) was performed as previously described (Lindeque *et al.*, 2013; Terburgh *et al.*, 2019) on the left hippocampi of FSL rats (Whitney *et al.*, 2023). Briefly, a stepwise Bligh-Dyer extraction

method (Wu *et al.*, 2008) was performed resulting in biphasic separation. Furthermore, all samples were derivatised via oximation and silylation as previously described (Lindeque *et al.*, 2013; Terburgh *et al.*, 2019) prior to GC-TOF-MS analysis. For data acquisition and extraction, the LECO Corporation ChromaTOF[®] software (v 4.5x) was utilised. The NIST MS search program (v 0.2) using AMDIS (National Institute of Standards and Technology) was used to compare measured spectra to the NIST 11 mass spectral library to identify all the detected components and validate relevant metabolites. A detailed description of the method is available as Supplementary data.

Statistical analyses

Statistical analyses were performed in IBM[®] SPSS[®] Statistics (version 28), assisted by Laerd Statistics[®] (<https://statisticslaerd.com>) and the NWU statistical consultation services. Effect magnitude indicators were calculated in Exploratory Software for Confidence Intervals (Cumming, 2014). All graphical representations were created in GraphPad Prism[®] (version 10) with the initial power analysis performed in G*Power (version 3; Universität Kiel, GER).

The Grubb's test was used to identify outliers and are reported in figure and table legends. Normality of distribution and homogeneity of variances were determined with the Shapiro-Wilk and Levene's tests, respectively. Only instances where these assumptions were not true are reported in the text. As for the metabolomic screening, normality of data was not analysed due to the number of measurements. Instead, data were simply log transformed and analysed with the appropriate statistical tests (Lindeque *et al.*, 2013) and normalised using the MSTUS normalisation method (Warrack *et al.*, 2009). First, normal two-way ANOVAs (analysis of variances) were used for PND21 analysis, with prenatal activity (EXE and SED) and early-life adversity (MSEW and non-MSEW) set as variables. Next, normal three-way ANOVAs were performed on PND36 parameters, with juvenile activity (SED and EXE), prenatal activity (SED and EXE) and early-life adversity (MSEW and non-MSEW) considered. Where locomotor activity was expected to influence results, appropriate ANCOVAs (analysis of co-variances) were performed to correct for this expected influence. A 5% confidence limit for error in all cases was accepted as statistically significant and reported as a Bonferroni-adjusted value. The mean differences between groups are reported with 95% confidence interval. For the GC-MS data, an independent *t*-test was performed in MetaboAnalyst version 5 (www.metaboanalyst.ca).

Statistical analyses were followed up by effect magnitude calculations (Cumming *et al.*, 2007; Lakens, 2013). Partial eta squared (η^2) and the unbiased Cohen's *d* (*d_{unb}*) values (Cumming, 2014) were used to calculate effect magnitude of interactions and intergroup differences, respectively. Large effect sizes were accepted as $\eta^2 \geq 0.14$ (Ellis, 2010) and $d \geq 0.8$ (Sullivan & Feinn, 2012). Importantly, to facilitate the interpretation of these findings, the effect magnitude values of the different behavioural parameters were calculated to identify the largest and statistically non-zero contributing factor (i.e. main effect), which was subsequently used to guide further analyses (see Section 3.1).

Results

Effects of prenatal activity and maternal separation and early weaning

In Fig. 2a, there was no significant interaction between prenatal activity (PRE) and early-life adversity (ELA) ($F_{1,117} = 0.74$,

$p = 0.39$, $\eta_p^2 = 0.006$), nor any significant main effect ($p > 0.05$) for distance moved in the OFT. Nonetheless, this parameter was used as a covariant in the analysis of the TST below.

Locomotor activity did not significantly influence immobility time in the TST ($F_{1,116} = 2.19$, $p = 0.14$, $\eta_p^2 = 0.019$), yet a significant PRE*ELA interaction was identified (Fig. 2b; $F_{1,116} = 5.02$, $p = 0.027$, $\eta_p^2 = 0.041$), with PRE also contributing as an independent factor ($F_{1,116} = 9.53$, $p = 0.003$, $\eta_p^2 = 0.076$). In terms of this effect, pups born to an EXE dam (regardless of ELA) were 27 s [10; 44 s] more immobile than their SED counterparts. More specifically, this effect only reached statistical significance in MSEW animals ($p \leq 0.0005$, $d_{unb} = 1.1$ [0.5; 1.7]).

Effects of pre-pubertal low-intensity exercise

The overall effect of the various factorial interactions and main effects of the behavioural parameters, including those discussed below, are presented in Fig. 3. Based on these results, juvenile activity (JUV) was the largest and statistically non-zero contributing factor that was used to guide further analyses and simplify the interpretation thereof. Still, all other significant findings are available as supplementary data.

FST behaviour on PND36, after correcting for locomotor differences

In Fig. 4a, a significant three-way interaction ($F_{1,114} = 6.89$, $p = 0.01$, $\eta_p^2 = 0.06$; *Supplementary data*), as well as a significant PRE*ELA interaction ($F_{1,114} = 6.77$, $p = 0.01$, $\eta_p^2 = 0.06$), existed for distance moved in the OFT on PND36. Despite narrowly missing significance ($F_{1,114} = 3.74$, $p = 0.056$, $\eta_p^2 = 0.03$), JUV independently trended to influence distance moved, so that pups that exercised (regardless of PRE and ELA) covered 235 cm [6; 476 cm] more than sedentary controls ($d_{unb} = 0.3$ [-0.1; 0.7]). These differences were subsequently used as a covariant in the FST analyses.

Distance moved in the OFT had no significant effect on any of the FST behavioural parameters ($p > 0.05$ in all instances; Table 2). After correcting for distance moved, there were no significant three-way interactions for time spent immobile ($F_{1,113} = 0.239$, $p = 0.63$, $\eta_p^2 = 0.002$), swimming ($F_{1,113} = 0.14$, $p = 0.71$, $\eta_p^2 = 0.001$) or struggling ($F_{1,113} = 0.55$, $p = 0.46$, $\eta_p^2 = 0.005$).

In addition to the significant PRE*ELA interaction (*Supplementary data*; $F_{1,113} = 4.85$, $p = 0.03$, $\eta_p^2 = 0.04$), JUV independently also influenced time spent immobile in the FST (Fig. 4b; $F_{1,113} = 5.94$, $p = 0.02$, $\eta_p^2 = 0.05$). Pups that exercised (regardless of PRE and ELA) were 13 s [2; 24 s] less immobile than their sedentary controls ($d_{unb} = 0.5$ [0.1; 0.9]).

For time spent swimming (Fig. 4c), three significant two-way interactions were identified (*Supplementary data*), with PRE*ELA considered the largest ($F_{1,113} = 18.64$, $p \leq 0.0005$, $\eta_p^2 = 0.14$). However, after correcting for distance moved, JUV, as independent factor, did not influence time spent swimming in the FST ($F_{1,113} = 2.19$, $p = 0.14$, $\eta_p^2 = 0.02$).

Only JUV influenced struggling behaviour (Fig. 4d) in the FST ($F_{1,113} = 4.20$, $p = 0.04$, $\eta_p^2 = 0.04$), so that pups that exercised (regardless of PRE or ELA) struggled 8.6 s [0.3; 17 s] longer than sedentary controls ($d_{unb} = 0.4$ [0.1; 0.8]).

EPM behaviour on PND36

In Fig. 5, there was no significant three-way interaction ($F_{1,109} = 0.89$, $p = 0.35$, $\eta_p^2 = 0.01$), nor any two-way interactions

Figure 2. PND21 effects of prenatal exercise on FSL offspring either exposed to early-life adversity or not. (a) distance moved (over 5 min) in the OFT^{a,b} and (b) time spent immobile in the TST on PND21. Data points represent the mean \pm 95% CI, with male and female indicated in blue and pink, respectively. Statistical analyses are reported in the text. ^{a)} not all data sets were normally distributed. ^{b)} outlier identified and excluded from analysis. EXE, pre-natal low-intensity exercise; MSEW, maternal separation and early weaning; SED, sedentary. TST: tail suspension test.

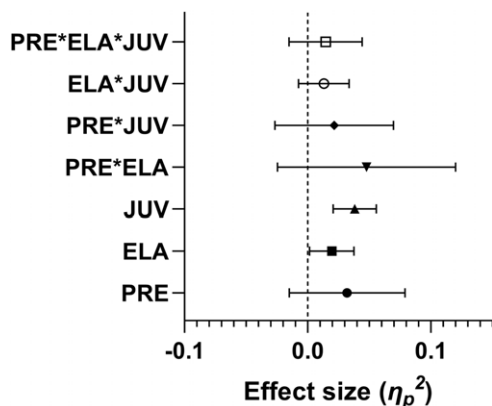
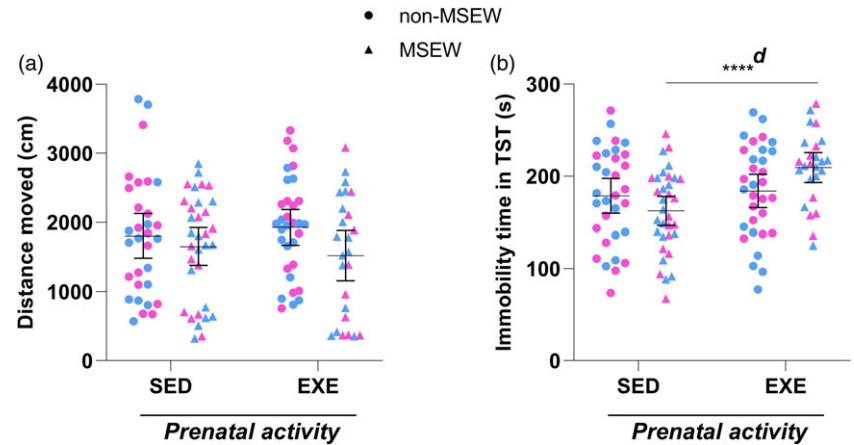


Figure 3. Forest plot of the overall behavioural effects of the contributing factors. ELA, early-life adversity; JUV, juvenile activity; PRE, prenatal activity.

identified for percentage time spent in the open arm of the EPM. However, JUV, independently influenced time spent in the open arms ($F_{1,109} = 6.22$, $p = 0.01$, $\eta_p^2 = 0.05$), so that pups that exercised (regardless of PRE and ELA) spent 9% [2; 15%] more time in the open arms, compared to sedentary controls ($d_{unb} = 0.5$ [0.1; 0.8]).

Anatomical markers

For whole brain (Fig. 6a; $F_{1,107} = 0.62$, $p = 0.43$, $\eta_p^2 = 0.006$) and heart (Fig. 6b; $F_{1,109} = 0.26$, $p = 0.62$, $\eta_p^2 = 0.002$) weight, there were no significant three-way interactions, nor any two-way interactions. However, PRE, ELA (Supplementary data) and JUV independently influenced whole brain ($F_{1,107} = 5.23$, $p = 0.02$, $\eta_p^2 = 0.02$) and heart ($F_{1,109} = 6.237$, $p = 0.014$, $\eta_p^2 = 0.05$) weights, so that the brains and hearts of pups that exercised (regardless of PRE and ELA), respectively, weighed 0.08% [0.02; 0.1%] ($d_{unb} = 0.3$ [-0.02; 0.7]) and 0.03% [0.01; 0.06%] ($d_{unb} = 0.5$ [0.1; 0.9]) more than that of their sedentary controls.

Hippocampal monoamine levels and redox state

In addition to the significant three-way interaction ($F_{1,102} = 18.033$, $p \leq 0.0005$, $\eta_p^2 = 0.15$; Supplementary data), hippocampal norepinephrine levels were also influenced by JUV (Fig. 7a; $F_{1,102} = 12.97$, $p \leq 0.0005$, $\eta_p^2 = 0.11$), independently, so that the levels of pups that exercised (regardless of PRE and ELA) were

233.75 ng/g [105; 363 ng/g] higher than that of sedentary controls ($d_{unb} = 0.4$ [0.0; 0.8]).

Hippocampal serotonin levels were also significantly influenced by a three-way interaction ($F_{1,106} = 13.05$, $p \leq 0.0005$, $\eta_p^2 = 0.110$; Supplementary data) and independently by JUV (Table 3; $F_{1,106} = 5.13$, $p = 0.026$, $\eta_p^2 = 0.046$). In line with the latter, pups that exercised (regardless of PRE and ELA) had 9.86 ng/g [4; 16 ng/g] more serotonin than sedentary controls ($d_{unb} = 0.3$ [-0.1; 0.7]). Despite these differences, hippocampal serotonin turnover (Fig. 7b) was comparable across groups.

In terms of hippocampal redox state, there was no significant three-way interaction ($F_{1,106} = 1.66$, $p = 0.20$, $\eta_p^2 = 0.015$), nor any significant two-way interactions ($p > 0.05$ in all instances; Supplementary data). However, JUV independently affected GSH/GSSG values (Fig. 7c; $F_{1,106} = 35.25$, $p \leq 0.0005$, $\eta_p^2 = 0.25$), so that this ratio (regardless of PRE and ELA) was 4.52 [3; 6] higher in pups that exercised, compared to sedentary controls ($d_{unb} = 0.9$ [0.5; 1.3]).

Metabolic markers relating to mitochondrial function

Summarised in Table 3, the effects of the largest identified behavioural influencing factor (i.e. juvenile activity; JUV) were significant in the following metabolic markers: palmitic acid (or hexadecenoic acid), stearic acid (or octadecanoic acid), oleic acid, 1-monopalmitin, and 1-monostearin and nicotinic acid (or niacin). Specifically, JUV (regardless of PRE and ELA) significantly ($p < 0.05$) decreased all these markers, relative to pups that did not exercise during pre-pubertal development (i.e. SED).

Discussion

In this work, we investigated the interaction between prenatal activity, an early-life stressor in the form of chronic MSEW, and pre-pubertal low-intensity exercise on the behavioural profile of an approved rodent model for depression. Importantly, the characteristic behavioural profile of the juvenile FSL rat was investigated and reported elsewhere (Whitney et al., 2023). Briefly, juvenile FSL rats (regardless of sex) displayed increased immobility and decreased escape-directed behaviour in the FST on PND36, together with increased hippocampal norepinephrine and serotonin turnover (5-HIAA/5-HT), and decreased GSH/GSSG values. In terms of the effect of an early-life stressor, we previously found that MSEW induced lasting behavioural deficits, as measured in the FST (Whitney et al., 2023). Here, both FSL and FRL rats (regardless of

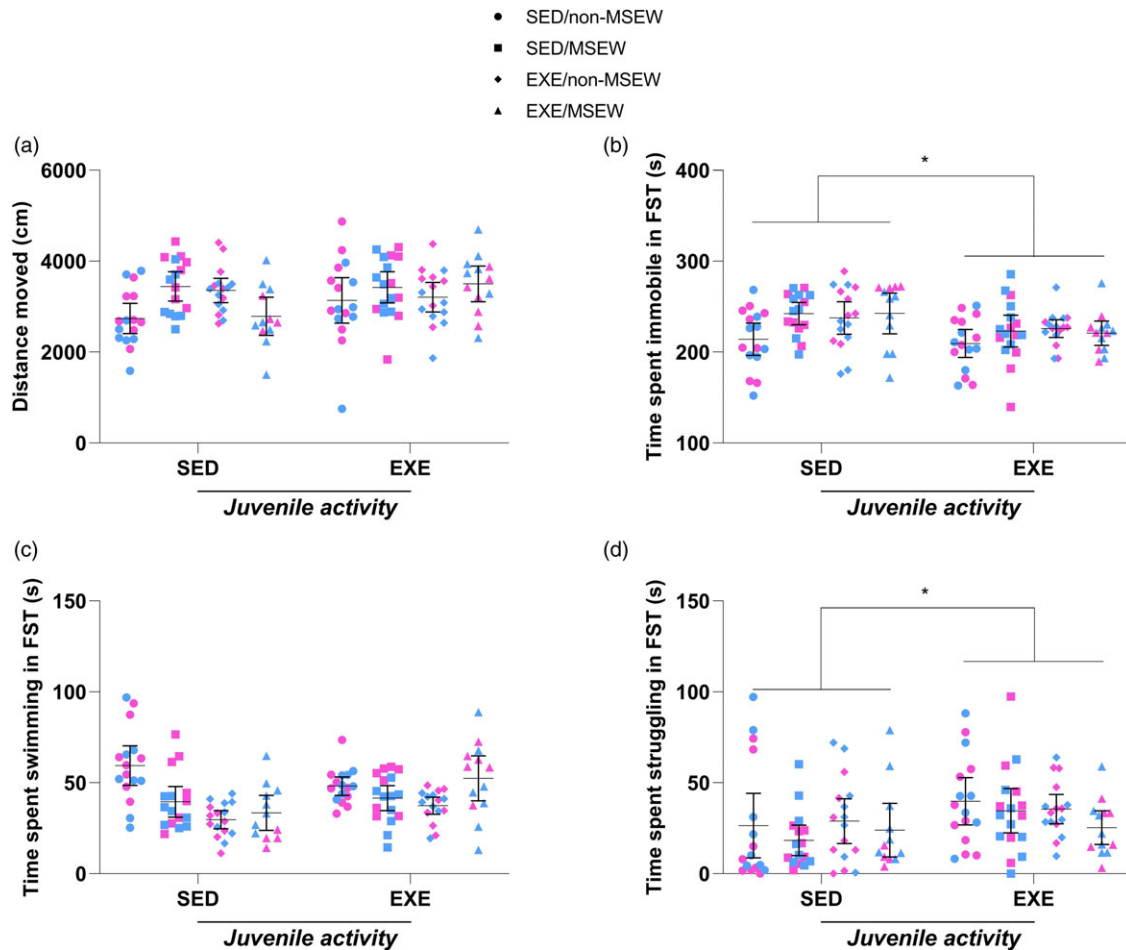


Figure 4. Behavioural effects on PND36. (a) Distance moved in the OFT. (b) Time spent immobile^{a,b,c}, (c) swimming^{a,b,c} and (d) struggling^{a,b,c} in the FST. Data points represent the mean \pm 95% CI, with male and female indicated in blue and pink, respectively. Statistical analyses are reported in the text. ^{a)} Not all data sets were normally distributed. ^{b)} Outlier identified but not excluded. ^{c)} Heterogeneity of variances. FST, forced swim test; EXE, juvenile low-intensity exercise; MSEW, maternal separation and early weaning; SED, sedentary.

sex) exposed to MSEW displayed increased depressive-like behaviour at PND36. Based on these findings, we could investigate whether prenatal and/or pre-pubertal low-intensity exercise could reverse or at least prevent the adverse effects caused by MSEW in a subject with a predisposed susceptibility to develop depression.

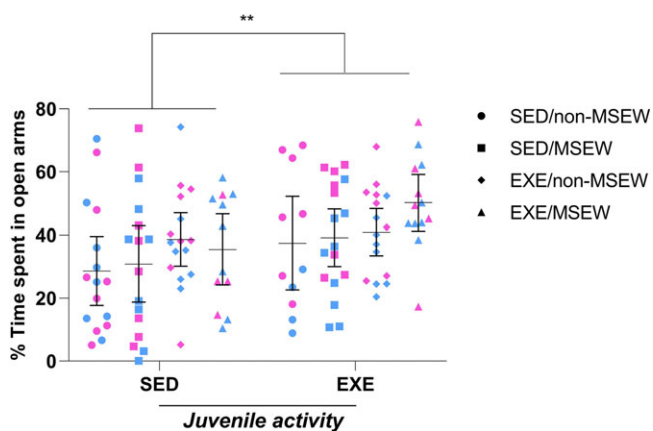
According to recent meta-analyses, exercise interventions induce an overall moderate beneficial effect in terms of childhood and adolescent depressive symptoms (Hu *et al.*, 2020; Wegner *et al.*, 2020; Axelsdottir *et al.*, 2021). Although the exact mechanism of action remains unknown (Schuch *et al.*, 2016), exercise has been linked to improved neuroplasticity (Gourgouvelis *et al.*, 2017; El-Sayes *et al.*, 2019), decreased neuro-inflammation (Eyre & Baune, 2012; Kandola *et al.*, 2019) and oxidative stress damage (Eyre & Baune, 2012; Schuch *et al.*, 2014; Lu *et al.*, 2021), enhanced monoaminergic neurotransmission (Lin & Kuo, 2013; da Costa Daniele *et al.*, 2017) and improved mitochondrial function (Aguiar *et al.*, 2014). Contrastingly, traumatic experiences during early-life development can negatively affect these pathways, eventually leading to impaired mood and/or increased anxiety levels (Palmier-Claus *et al.*, 2016; LeMoult *et al.*, 2020). We therefore aimed to determine whether prenatal low-intensity exercise could induce protective mechanisms against the adverse effects of early-life stress, and whether pre-pubertal exercise, with or without prenatal exercise, could induce antidepressant effects during juvenile development.

The early-life effects of prenatal activity and an early-life stressor (i.e. MSEW)

The beneficial effects of prenatal exercise are well established and proven to be safe for both the mother and foetus (Davenport *et al.*, 2018; Davenport *et al.*, 2018; Moyer *et al.*, 2016). Several groups have reported on the beneficial metabolic effects of maternal exercise in the rodent offspring (reviewed by Kusuyama *et al.* (2020)), with others also observing cardiovascular (May *et al.*, 2014) and even neuro-behavioural benefits in new-born babies (Clapp *et al.*, 1999). Here, pregnant FSL dams were subjected to a low intensity, treadmill exercise regimen, for an average of 13 ± 5 days. Pups born to these dams (regardless of sex and ELA) were more immobile in the TST on PND21, compared to those born to a sedentary dam. Interestingly, this effect was more prominent in animals exposed to MSEW (Fig. 2b). Others have suggested prenatal exercise to improve cognitive function and induce anxiolytic effects in rodent offspring, in part by increasing hippocampal neuroplasticity (Aksu *et al.*, 2012; Ji *et al.*, 2020). However, a limitation of the current study is that no neurochemical markers were measured at this age (i.e. PND21), leading to any explanation of these behavioural differences to be speculative. Still, that prenatal exercise can influence the hypothalamic-adrenal axis (Carlberg *et al.*, 1996) suggests that prenatal exercise could alter

Table 2. Original and ANCOVA-adjusted parameters of the TST and FST

PRE	ELA	JUV	TST immobility (s)		FST immobility (s)		
			Original mean \pm SD (n)	Adjusted mean \pm SEM (n)	Original mean \pm SD (n)	Adjusted mean \pm SEM (n)	
Sedentary	Normal	SED	178.69 \pm 52.28 (32)	179.52 \pm 8.37 (32)	214.18 \pm 33.42 (16)	211.86 \pm 7.54 (16)	
		EXE			209.48 \pm 28.84 (16)	209.13 \pm 7.30 (16)	
	MSEW	SED	162.35 \pm 44.43 (33)	161.68 \pm 8.23 (33)	242.35 \pm 23.07 (16)	243.53 \pm 7.35 (16)	
		EXE			223.13 \pm 33.95 (17)	224.19 \pm 7.13 (17)	
	Exercise	Normal	SED	183.93 \pm 50.34 (32)	184.92 \pm 9.38 (32)	237.51 \pm 33.61 (16)	238.24 \pm 7.31 (16)
			EXE			225.88 \pm 18.42 (16)	225.89 \pm 7.29 (16)
MSEW		SED	209.39 \pm 39.29 (25)	207.93 \pm 9.52 (25)	242.53 \pm 35.27 (12)	240.45 \pm 8.59 (12)	
		EXE			220.76 \pm 22.21 (13)	222.21 \pm 8.17 (13)	
PRE		ELA	JUV	FST swimming (s)		FST struggling (s)	
				Original mean \pm SD (n)	Adjusted mean \pm SEM (n)	Original mean \pm SD (n)	Adjusted mean \pm SEM (n)
Sedentary	Normal	SED	59.44 \pm 20.43 (16)	60.16 \pm 3.77 (16)	26.43 \pm 33.24 (16)	28.07 \pm 5.85 (16)	
		EXE	48.15 \pm 9.57 (16)	48.26 \pm 3.65 (16)	39.82 \pm 24.48 (16)	40.06 \pm 5.66 (16)	
	MSEW	SED	39.51 \pm 15.74 (16)	39.14 \pm 3.68 (16)	18.29 \pm 15.78 (16)	17.46 \pm 5.71 (16)	
		EXE	41.55 \pm 13.28 (17)	41.22 \pm 3.57 (17)	34.62 \pm 23.69 (17)	33.87 \pm 5.53 (17)	
	Exercise	Normal	SED	29.66 \pm 9.36 (16)	29.43 \pm 3.66 (16)	28.92 \pm 23.09 (16)	28.41 \pm 5.68 (16)
			EXE	37.38 \pm 8.77 (16)	37.38 \pm 3.65 (16)	35.51 \pm 15.17 (16)	35.50 \pm 5.66 (16)
MSEW		SED	33.44 \pm 15.13 (12)	34.08 \pm 4.30 (12)	23.94 \pm 23.27 (12)	25.40 \pm 6.67 (12)	
		EXE	52.42 \pm 20.51 (13)	51.97 \pm 4.09 (13)	25.29 \pm 15.30 (13)	24.27 \pm 6.35 (13)	

**Figure 5.** Percentage time spent in the open arm of the EPM. Data points represent the mean \pm 95% CI, with male and female indicated in blue and pink, respectively. Statistical analyses are reported in the text. EXE: juvenile low-intensity exercise. MSEW: maternal separation and early weaning. SED: sedentary.

behaviour and stress responses. Consequently, the behaviour observed in the TST on PND21 would benefit from corticosterone analyses as a recent meta-analysis concluded that although cortisol release is blunted in children and adolescents who suffered ELA, these effects were more prominent in adults (Bunea *et al.*, 2017). Conversely, an earlier report found that prenatal exercise increased foetal corticosterone levels (Carlberg *et al.*, 1996), while another found that offspring of women who

regularly exercised during pregnancy scored better on the Bayley psychomotor scale (Clapp *et al.*, 1998). Considered together, although on face value, the TST behaviour may represent improved depressive-like behaviour, and further neurochemical analyses are required to confirm such interpretation. Later in life, pups born to an exercised dam also displayed increased immobility time and decreased swimming behaviour (Figure 4 and Supplementary data) in the FST, compared to those born to a sedentary dam. These behaviours again point towards a depressogenic effect yet considering the reduced hippocampal serotonin turnover, improved redox state, and increased brain weight on PND38 (Figure 7 and Supplementary data), it may be that prenatal exercise may actually induce a more resilient behaviour. In this regard, although the FST is an accepted screening tool for antidepressant-like effects, this simplified interpretation of time spent immobile as an indicator of depressive-like behaviour has been challenged (Boccia *et al.*, 2007; Commons *et al.*, 2017). Therefore, pending confirmation, our results may suggest that prenatal exercise does indeed induce significant behavioural changes in adolescent FSL rats, and that these effects may (or may not) resemble improved coping behaviour.

As to whether prenatal exercise could prevent the adverse effects of MSEW, it is first worth noting that we have previously shown that MSEW worsens depressive-like behaviour in the Flinders line rat, with more prominent effects in the resistant (i.e. FRL) strain (Whitney *et al.*, 2023). Similarly, in the current study, MSEW also increased time spent immobile in the FST on PND36 and decreased the whole brain weight of FSL rats (Supplementary data), thereby supporting a depressogenic effect, caused by an

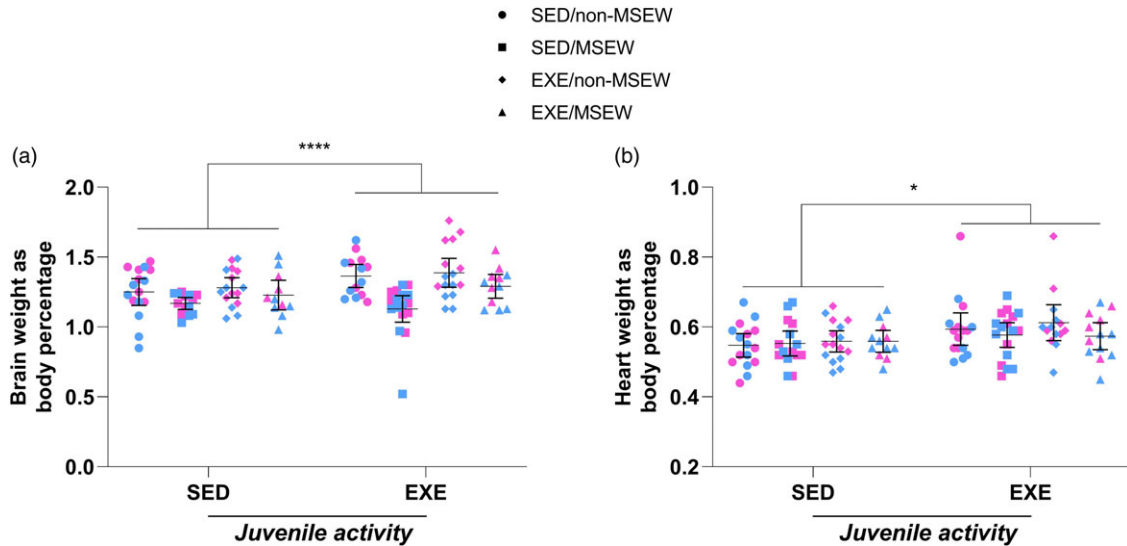


Figure 6. Heart and whole brain weight of male and female FSL rats. (a) Brain^{a,b} and (b) heart^{b,c} weight of FSL rats, expressed as a percentage of body weight. Data points represent the mean ± 95% CI, with male and female indicated in blue and pink, respectively. Statistical analyses are reported in the text. ^{a)} Not all data-sets were normally distributed. ^{b)} Outlier identified and excluded. ^{c)} Outliers identified but not excluded. EXE, juvenile low-intensity exercise; MSEW, maternal separation and early weaning; SED, sedentary.

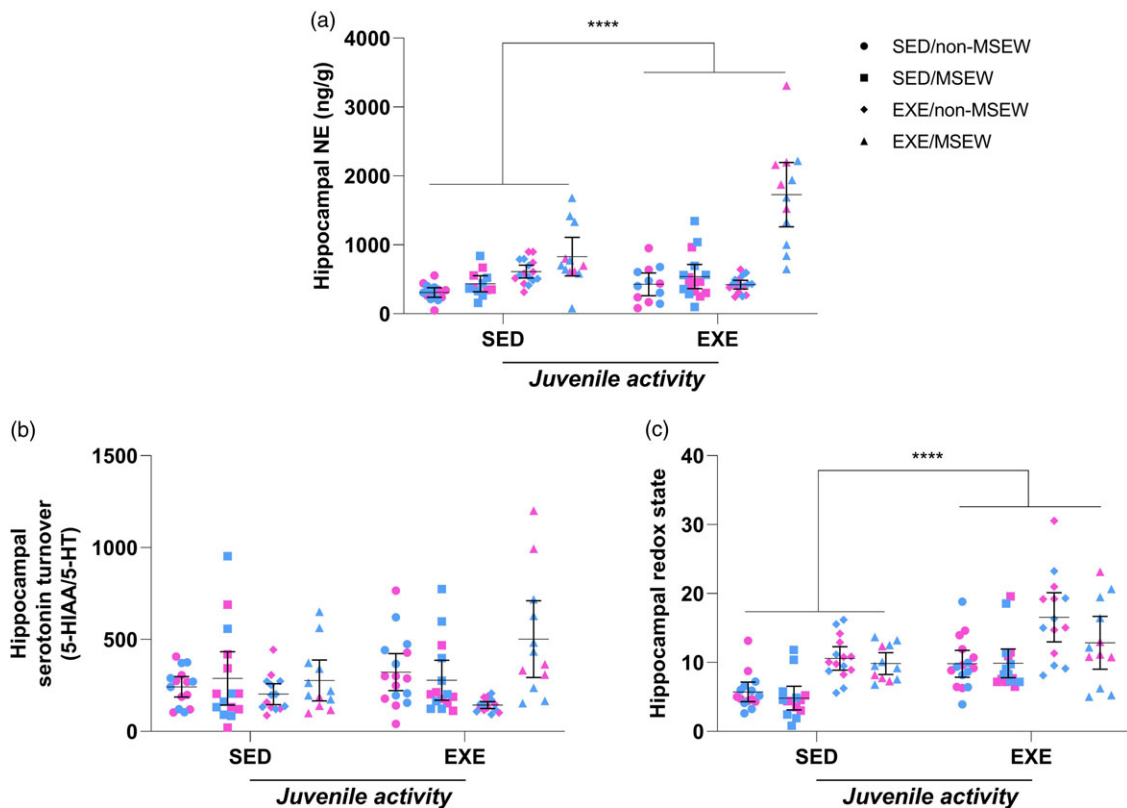


Figure 7. Hippocampal monoamine levels and redox state markers. (a) Norepinephrine levels^{a,c}, (b) serotonin turnover (5-HIAA/5-HT)^{a,b,c}, (c) redox state (GSH/GSSG)^{a,b,c} on PND38. Data points represent the mean ± 95% CI, with male and female indicated in blue and pink, respectively. Statistical analyses are reported in the text. ^{a)} Outliers identified and excluded. ^{b)} Not all data-sets are normally distributed. ^{c)} Heterogeneity of variances. EXE, juvenile low-intensity exercise; MSEW, maternal separation and early weaning; SED, sedentary.

early-life stressor. However, none of the parameters where PRE and MSEW interacted to influence the outcome, showed any statistical evidence to accurately answer this question. Still, that prenatal activity (regardless of ELA and JUV) beneficially altered

hippocampal redox state and serotonin turnover during pubertal onset, at least hints towards a protective effect that was not inhibited nor prevented by an early-life stressor (*Supplementary data*). Further investigation into this aspect is however required.

Table 3. Metabolic markers in the hippocampus of FSL rats that relate to mitochondrial function

PRE	ELA	JUV	Palmitic acid mean ± SD (n)	Stearic acid mean ± SD (n)	Oleic acid mean ± SD (n)
Sedentary	Normal	SED	4.40 ± 0.76 (3)	12.31 ± 3.71 (3)	2.54 ± 1.21 (3)
		EXE	1.92 ± 0.74 (10)	8.09 ± 2.15 (10)	1.71 ± 0.62 (10)
	MSEW	SED	3.50 ± 0.63 (7)	10.23 ± 0.79 (7)	1.69 ± 0.20 (7)
		EXE	2.45 ± 0.84 (9)	8.15 ± 2.18 (9)	1.46 ± 0.37 (9)
Exercise	Normal	SED	2.45 ± 0.51 (10 ^a)	7.54 ± 1.79 (10 ^a)	1.50 ± 0.47 (11)
		EXE	2.19 ± 0.70 (9)	7.09 ± 1.89 (9)	1.24 ± 0.31 (9)
	MSEW	SED	2.72 ± 0.65 (9)	8.68 ± 1.14 (9)	1.40 ± 0.14 (9)
		EXE	2.90 ± 0.66 (8)	9.05 ± 2.17 (8)	1.34 ± 0.27 (8)
PRE	ELA	JUV	1-monopalmitin mean ± SD (n)	1-monostearin mean ± SD (n)	Nicotinic acid mean ± SD (n)
Sedentary	Normal	SED	0.64 ± 0.07 (3)	1.34 ± 0.001 (2 ^a)	101.75 ± 1.81 (2 ^a)
		EXE	0.27 ± 0.13 (10)	0.59 ± 0.19 (10)	12.52 ± 13.10 (10)
	MSEW	SED	0.47 ± 0.14 (7)	0.86 ± 0.20 (7)	77.00 ± 46.57 (7)
		EXE	0.31 ± 0.12 (9)	0.63 ± 0.19 (9)	30.99 ± 18.56 (9)
Exercise	Normal	SED	0.33 ± 0.14 (11)	0.69 ± 0.21 (10)	33.67 ± 18.69 (11)
		EXE	0.29 ± 0.08 (9)	0.51 ± 0.11 (9)	26.62 ± 9.05 (5 ^a)
	MSEW	SED	0.30 ± 0.03 (8 ^a)	0.57 ± 0.09 (9)	34.62 ± 15.54 (8)
		EXE	0.29 ± 0.05 (8)	0.57 ± 0.12 (8)	55.39 ± 46.05 (8)

ELA, early-life adversity; JUV, juvenile activity; PRE, prenatal activity.

^aOutlier identified and removed from analysis.

The values presented here are all log transformed and therefore contain no SI unit. Group sizes differ from behavioural analyses and could be explained by the storage buffer used. Because of the significant influence of main effects, all statistical findings are reported in text.

Table 4. Hippocampal serotonergic and redox state markers

PRE	ELA	JUV	5-HT (ng/g) mean ± SD (n)	5-HIAA (ng/g) mean ± SD (n)	5-HIAA/5-HT mean ± SD (n)
Sedentary	Normal	SED	19.8 ± 7.9 (15)	242.5 ± 101.3 (15)	13.9 ± 6.5 (15)
		EXE	23.3 ± 15.4 (16)	332.0 ± 188.5 (16)	19.7 ± 17.8 (16)
	MSEW	SED	19.0 ± 7.5 (15)	288.1 ± 262.3 (15)	16.3 ± 13.2 (15)
		EXE	20.8 ± 9.6 (15)	278.1 ± 195.9 (15)	14.2 ± 6.9 (15)
Exercise	Normal	SED	23.2 ± 10.7 (14)	202.6 ± 97.9 (14)	10.0 ± 5.2 (14)
		EXE	13.6 ± 3.8 (15)	143.4 ± 34.2 (15)	11.1 ± 3.4 (15)
	MSEW	SED	24.6 ± 13.8 (12)	277.0 ± 175.6 (12)	13.0 ± 7.0 (12)
		EXE	54.8 ± 36.1 (12)	501.3 ± 328.6 (12)	10.8 ± 6.5 (12)
Redox state markers					
PRE	ELA	JUV	GSH (µg/g) mean ± SD (n)	GSSG (µg/g) mean ± SD (n)	GSH/GSSG mean ± SD (n)
Sedentary	Normal	SED	42.3 ± 7.3 (15)	8.4 ± 2.8 (15)	5.7 ± 2.6 (15)
		EXE	58.8 ± 25.1 (16)	6.3 ± 2.4 (16)	9.8 ± 3.7 (16)
	MSEW	SED	41.8 ± 21.8 (14)	9.7 ± 4.4 (14)	4.8 ± 3.0 (14)
		EXE	56.5 ± 11.5 (16)	6.2 ± 1.8 (16)	9.9 ± 3.9 (16)
Exercise	Normal	SED	50.5 ± 11.4 (15)	5.1 ± 1.5 (15)	10.6 ± 3.1 (15)
		EXE	33.0 ± 3.9 (14)	2.2 ± 0.8 (14)	16.6 ± 6.2 (14)
	MSEW	SED	77.2 ± 31.5 (12)	7.9 ± 2.4 (12)	9.8 ± 2.5 (12)
		EXE	138.9 ± 55.5 (12)	12.7 ± 7.0 (12)	12.8 ± 6.0 (12)

The mean values of the specific markers are presented here to promote transparency and were used to calculate the serotonin turnover and redox state reported in the results section. Because of the significant influence of main effects, all statistical findings are reported in text.

The influence of pre-pubertal low-intensity exercise

As mentioned earlier, exercise is a recognised non-pharmacological treatment strategy for depression (Carter *et al.*, 2016; Kandola *et al.*, 2019) and importantly, can be implemented across all ages, making it a promising treatment option for childhood depression (Hu *et al.*, 2020; Wegner *et al.*, 2020; Axelsdottir *et al.*, 2021). Therefore, the current study investigated whether a 14-day low-intensity exercise regimen during pre-pubertal development could attenuate the depressive-like phenotype of the adolescent FSL rat. It must however be noted here that although all three factors (early-life adversity, and prenatal and juvenile exercise) were considered and controlled for in the study design, the delayed effect findings are interpreted and discussed only in terms of the most robust and statistically non-zero factor (Fig. 3), that is, pre-pubertal low-intensity exercise. All other statistical findings are available as supplementary data.

On PND36, animals (regardless of sex) that were exposed to low-intensity exercise during pre-pubertal development (PND22 to 35) displayed decreased depressive-like behaviour (i.e. time spent immobile) in the FST, compared to their sedentary controls (Fig. 4b), irrespective of PRE and ELA. Moreover, this behaviour was also accompanied by increased time spent struggling (Fig. 4d) – indicative of antidepressant-like and/or increased coping behaviour (Lucki, 1997). Our findings of exercise exerting antidepressant-like effects are in line with pre-clinical (Steyn *et al.*, 2020; Gruhn *et al.*, 2021; de Oliveira *et al.*, 2022; Sohroforouzani *et al.*, 2022) and clinical (Oberste *et al.*, 2020) findings. Although the FSL rat is not known to display increased anxiety-like behaviour (Overstreet & Wegener, 2013), pre-pubertal low-intensity exercise appeared to induce anxiolytic-like effects in FSL rats on PND37 (Fig. 5). It must be mentioned that this effect was only relative to sedentary FSL, and not FRL controls, and therefore requires confirmational studies. Regardless, clinical (Stubbs *et al.*, 2017) and pre-clinical (Cevik *et al.*, 2018) studies that have also reported on the anxiolytic effect of exercise further validates our findings and supports the efficacy of low-intensity exercise as a treatment option for childhood depression.

To confirm our behavioural findings and shed further light on the probable mechanisms involved, we considered anatomical markers, and measured hippocampal monoamine levels, together with markers of oxidative stress and mitochondrial function. First, pre-pubertal low-intensity exercise (regardless of PRE and ELA) increased the brain and heart weights of male and female FSL rats, relative to sedentary control groups (Fig. 6a, b). This is a noteworthy finding, as we have previously reported, that the adolescent FSL rat has lower whole brain and heart weights than its age matched FRL counterpart (Whitney *et al.*, 2023). Therefore, that chronic low-intensity exercise increased brain weight, suggests neuroplasticity mechanisms to be at play (Gourgouvelis *et al.*, 2017; El-Sayes *et al.*, 2019). These findings however warrant confirmation by means of analysing appropriate markers, such as brain-derived neurotrophic factor – a neurotrophin known to be decreased in depressed patients (Brunoni *et al.*, 2008) and increased by exercise (Luo *et al.*, 2019; Naghibi *et al.*, 2021). Exercise further directly affects autonomic outflow, benefitting cardiovascular functioning (Gademan *et al.*, 2007), and although no cardiac tissue biomarkers were measured, our finding of exercise-induced hypertrophy is at least supported by clinical studies (Xiang *et al.*, 2020). That autonomic dysfunction has been shown to be altered in depressed patients (Hartmann *et al.*, 2019; Herbsleb *et al.*, 2020), further emphasises the value of our findings in a genetic rodent model of depression.

As an indirect indicator of mitochondrial function, pre-pubertal low-intensity exercise also beneficially influenced the hippocampal redox state (GSH/GSSG; Fig. 7c), suggesting antioxidant defences to be increased. The GSH/GSSG ratio is a valuable biomarker of cellular redox state (Enns & Cowan, 2017), with lower levels indicating increased oxidative stress (Chai *et al.*, 1994). Our findings are in agreement with others (Higashi, 2016) showing that regular low- to moderate-intensity exercise induces beneficial effects. To this point, that a dysfunctional redox state has previously been observed in the juvenile FSL strain (relative to FRL controls; Whitney *et al.*, 2023), allow us to conclude that pre-pubertal exercise can reverse this deficit. That this improved redox state was observed together with increased brain weight (Fig. 6a), suggests that oxidative stress damage, specifically in the hippocampus was mitigated, potentially via improvement of mitochondrial function (Memme *et al.*, 2021) and enhanced neuroplasticity and neurogenesis (Park *et al.*, 2018; Park *et al.*, 2019), thereby supporting increasing evidence describing depression as a bio-energetic disorder. To this point, that pre-pubertal low-intensity exercise decreased hexadecanoic acid (palmitic acid), octadecanoic acid (stearic acid), oleic acid (octadecenoic acid), 1-monopalmitin (a monoacylglycerol with hexadecanoic acid), and 1-monostearin, which points towards improved mitochondrial function. These metabolic markers are generally observed in patients with metabolic syndrome and insulin resistance, both conditions strongly associated with mitochondrial dysfunction (Pari & Venkateswaran, 2004; Zeng *et al.*, 2009; Ma *et al.*, 2015). In these patients, a skewed ATP:ADP ratio stimulates lipolysis, which leads to the breakdown of triacylglycerols into monoacylglycerol and free fatty acids (especially hexadecanoic acid and octadecanoic acid). That these levels were decreased in the hippocampi of animals that exercised during pre-pubertal development in the current study, which is likely indicative of increased energy metabolism (and consequently mitochondrial function). Moreover, that these animals also had lower nicotinic acid levels than their sedentary counterparts, which may suggest a lower breakdown of nicotinamide adenine dinucleotide (NAD⁺) and/or better utilisation of nicotinic acid in the formation of NAD⁺. Briefly, nicotinic acid is a precursor of NAD⁺, which acts as an electron carrier in the electron transport chain, where it regulates the redox state of the mitochondria and contributes to ATP production (Crowley *et al.*, 2000; Sauve, 2008). Although clinical and pre-clinical research differs in terms of the effect of exercise on NAD⁺ levels (White & Schenk, 2012), decreased levels are generally associated with age-associated pathologies (reviewed by (Imai & Guarente, 2014)). However, considered with the behavioural and neurochemical alterations reported here, and the known mitochondrial enhancing effects of exercise (Memme *et al.*, 2021), pre-pubertal low-intensity exercise may have increased the nicotinic acid to NAD⁺ conversion, thereby decreasing the available nicotinic levels and potentially increasing the NAD⁺/NADH ratio. This is at least partly supported by our previous finding that pubertal FSL rats have higher hippocampal nicotinic concentrations than FRL controls (Whitney *et al.*, 2023). Still, we invite confirmatory investigations.

Finally, pre-pubertal low-intensity exercise (regardless of PRE and ELA) also increased hippocampal norepinephrine and serotonin, relative to sedentary controls (Fig. 7a and Table 4), without affecting serotonin turnover (Fig. 7b). These increases support the decrease in depressive-like behaviour and the increase in escape-directed behaviour observed in the FST. As mentioned earlier, one of the mechanisms through which exercise exerts its antidepressant-like effects is by increasing

monoamine neurotransmission and considered together with the improved hippocampal redox state and mitochondrial markers, our findings reaffirm this effect and again show that exercise can mimic currently approved pharmacological treatment options. Further research is, however, needed into whether the apparent antidepressant-like effects of pre-pubertal exercise are indeed unrelated to ELA and PRE.

Conclusion

The current study explored three factors influencing depressive-like behaviour in an approved genetic rodent model of depression to demonstrate how environmental and genetic influences can alter this behaviour. Our findings show that pre-natal exercise induces beneficial long-term neurochemical alterations that is unaffected by an early-life stressor. Pre-pubertal low intensity was effective in reducing depressive-like behaviour and oxidative stress in a rodent model of depression, whilst also increasing monoaminergic levels, and in doing so, implicating improved mitochondrial function. Taken together, our findings highlight the need to further investigate the role of mitochondrial function in depression and support the use of pre-pubertal low-intensity exercise as an effective treatment strategy for childhood depression.

Supplementary material. The supplementary material for this article can be found at <https://doi.org/10.1017/neu.2023.52>.

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Author contribution. SFS conceptualised and designed the layout of the study. AJW performed all the experimental work and analysed and interpreted the data with SFS. AJW and SFS also wrote the original draft of the manuscript, with ZL and RK collating and finalising the paper for submission. All authors contributed to the various sections of this paper. The authors would like to acknowledge Dr Laneke Luies for her assistance and support with the GC-TOM-MS system.

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Animal welfare. All animal procedures were approved by the Animal Care, Health and Safety Research Ethics Committee of the North-West University (NWU-AnimCareREC; ethics approval number: NWU-00419-21-A5) and in accordance with the relevant code of ethics. All procedures complied with national legislation that pertains to experimental animal welfare (including the Department of Health's Ethics in Health Research: Principles, Processes and Structures and the South African National Standard: The Care and Use of Animals for Scientific Purposes (SANS 10,386:2008)). This work also complied with the ARRIVE guidelines ensuring that all experimental data are reproducible, transparent, accurate comprehensive and logically ordered to promote well written manuscripts.

Ethical standards. The authors assert that all procedures contributing to this work comply with the ethical standards of the South African National Standards and institutional guides on the care and use of laboratory animals.

References

- Aguiar AS, Stragier E, da Luz Scheffer D, Remor AP, Oliveira PA, Prediger RD, Latini A, Raisman-Vozari R, Mongeau R and Lanfumey L (2014) Effects of exercise on mitochondrial function, neuroplasticity and anxiety-depressive behavior of mice. *Neuroscience* 271, 56–63. DOI: [10.1016/j.neuroscience.2014.04.027](https://doi.org/10.1016/j.neuroscience.2014.04.027).
- Aksu I, Baykara B, Ozbal S, Cetin F, Sisman AR, Dayi A, Gencoglu C, Tas A, Büyük E, Gonenc-Arda S and Uysal N (2012) Maternal treadmill exercise during pregnancy decreases anxiety and increases prefrontal cortex VEGF and BDNF levels of rat pups in early and late periods of life. *Neuroscience Letters* 516(2), 221–225. DOI: [10.1016/j.neulet.2012.03.091](https://doi.org/10.1016/j.neulet.2012.03.091).
- Allen J, Romay-Tallon R, Brymer KJ, Caruncho HJ and Kalynchuk LE (2018) Mitochondria and mood: mitochondrial dysfunction as a key player in the manifestation of depression. *Frontiers in Neuroscience* 12, 386. DOI: [10.3389/fnins.2018.00386](https://doi.org/10.3389/fnins.2018.00386).
- Andersen SL and Teicher MH (2008) Stress, sensitive periods and maturational events in adolescent depression. *Trends in Neurosciences* 31(4), 183–191. DOI: [10.1016/j.tins.2008.01.004](https://doi.org/10.1016/j.tins.2008.01.004).
- Axelsdottir B, Biedilä S, Sagatun Å., Nordheim LV and Larun L (2021) Exercise for depression in children and adolescents – a systematic review and meta-analysis. *Child and Adolescent Mental Health* 26(4), 347–356. DOI: [10.1111/camh.12438](https://doi.org/10.1111/camh.12438).
- Bay-Richter C, Petersen E, Liebenberg N, Elfving B and Wegener G (2019) Latent toxoplasmosis aggravates anxiety- and depressive-like behaviour and suggest a role of gene-environment interactions in the behavioural response to the parasite. *Behavioural Brain Research* 364, 133–139. DOI: [10.1016/j.bbr.2019.02.018](https://doi.org/10.1016/j.bbr.2019.02.018).
- Bérard A, Zhao J-P and Sheehy O (2017) Antidepressant use during pregnancy and the risk of major congenital malformations in a cohort of depressed pregnant women: an updated analysis of the quebec pregnancy cohort. *BMJ Open* 7(1), e013372. DOI: [10.1136/bmjopen-2016-013372](https://doi.org/10.1136/bmjopen-2016-013372).
- Biddle SJ, Ciacconi S, Thomas G and Vergeer I (2019) Physical activity and mental health in children and adolescents: an updated review of reviews and an analysis of causality. *Psychology of Sport and Exercise* 42, 146–155. DOI: [10.1016/j.psychsport.2018.08.011](https://doi.org/10.1016/j.psychsport.2018.08.011).
- Boccia ML, Razzoli M, Vadlamudi SP, Trumbull W, Caleffie C and Pedersen CA (2007) Repeated long separations from pups produce depression-like behavior in rat mothers. *Psychoneuroendocrinology* 32(1), 65–71. DOI: [10.1016/j.psyneuen.2006.10.004](https://doi.org/10.1016/j.psyneuen.2006.10.004).
- Brand SJ and Harvey BH (2017) Exploring a post-traumatic stress disorder paradigm in Flinders sensitive line rats to model treatment-resistant depression I: bio-behavioural validation and response to imipramine. *Acta Neuropsychiatrica* 29(4), 193–206. DOI: [10.1017/neu.2016.44](https://doi.org/10.1017/neu.2016.44).
- Brunoni AR, Lopes M and Fregni F (2008) A systematic review and meta-analysis of clinical studies on major depression and BDNF levels: implications for the role of neuroplasticity in depression. *The International Journal of Neuropsychopharmacology* 11(8), 1169–1180. DOI: [10.1017/S1461145708009309](https://doi.org/10.1017/S1461145708009309).
- Bryan RJr (1990) Cerebral blood flow and energy metabolism during stress. *American Journal of Physiology - Heart and Circulatory Physiology* 259(2), H269–H280. DOI: [10.1152/ajpheart.1990.259.2.H269](https://doi.org/10.1152/ajpheart.1990.259.2.H269).
- Bunea IM, Szentágotai-Táttar A and Miu AC (2017) Early-life adversity and cortisol response to social stress: a meta-analysis. *Translational Psychiatry* 7(12), 1–8. DOI: [10.1038/s41398-017-0032-3](https://doi.org/10.1038/s41398-017-0032-3).
- Carlberg KA, Alvin BL and Gwosdow AR (1996) Exercise during pregnancy and maternal and fetal plasma corticosterone and androstenedione in rats. *American Journal of Physiology-endocrinology and Metabolism* 271(5), E896–E902. DOI: [10.1152/ajpendo.1996.271.5.E896](https://doi.org/10.1152/ajpendo.1996.271.5.E896).
- Carter T, Morres ID, Meade O and Callaghan P (2016) The effect of exercise on depressive symptoms in adolescents: a systematic review and meta-analysis. *Journal of the American Academy of Child & Adolescent Psychiatry* 55(7), 580–590. DOI: [10.1016/j.jaac.2016.04.016](https://doi.org/10.1016/j.jaac.2016.04.016).
- Caruso G, Benatti C, Blom J, Caraci F and Tascadda F (2019) The many faces of mitochondrial dysfunction in depression: from pathology to treatment. *Frontiers in Pharmacology* 10, 995. DOI: [10.3389/fphar.2019.00995](https://doi.org/10.3389/fphar.2019.00995).
- Castagné V, Moser P, Roux S and Porsolt RD (2010) Rodent models of depression: forced swim and tail suspension behavioral despair tests in rats

- and mice. *Current Protocols in Pharmacology* 49(1), 5 8 1–5 8 14. DOI: [10.1002/0471141755.ph0508s49](https://doi.org/10.1002/0471141755.ph0508s49).
- Centers for Disease Control and Prevention.** 2021. *Data and statistics on children's mental health* <https://www.cdc.gov/childrensmentalhealth/data.html>. Accessed: April 19, 2021.
- Cevik OS, Sahin L and Tamer L** (2018) Long term treadmill exercise performed to chronic social isolated rats regulate anxiety behavior without improving learning. *Life Sciences* 200, 126–133. DOI: [10.1016/j.lfs.2018.03.029](https://doi.org/10.1016/j.lfs.2018.03.029).
- Chai Y-C, Ashraf SS, Rokutan K, Johnston RB and Thomas JA** (1994) S-thiolation of individual human neutrophil proteins including actin by stimulation of the respiratory burst: evidence against a role for glutathione disulfide. *Archives of Biochemistry and Biophysics* 310(1), 273–281. DOI: [10.1006/abbi.1994.1167](https://doi.org/10.1006/abbi.1994.1167).
- Clapp JF III, Lopez B and Harcar-Sevcik R** (1999) Neonatal behavioral profile of the offspring of women who continued to exercise regularly throughout pregnancy. *American Journal of Obstetrics and Gynecology* 180(1), 91–94. DOI: [10.1016/S0002-9378\(99\)70155-9](https://doi.org/10.1016/S0002-9378(99)70155-9).
- Clapp JF III, Simonian S, Lopez B, Appleby-Wineberg S and Harcar-Sevcik R** (1998) The one-year morphometric and neurodevelopmental outcome of the offspring of women who continued to exercise regularly throughout pregnancy. *American Journal of Obstetrics and Gynecology* 178(3), 594–599. DOI: [10.1016/S0002-9378\(98\)70444-2](https://doi.org/10.1016/S0002-9378(98)70444-2).
- Commons KG, Cholanians AB, Babb JA and Ehlinger DG** (2017) The rodent forced swim test measures stress-coping strategy, not depression-like behavior. *ACS Chemical Neuroscience* 8(5), 955–960. DOI: [10.1021/acschemneuro.7b00042](https://doi.org/10.1021/acschemneuro.7b00042).
- Crowley C, Payne C, Bernstein H, Bernstein C and Roe D** (2000) The NAD⁺ precursors, nicotinic acid and nicotinamide protect cells against apoptosis induced by a multiple stress inducer, deoxycholate. *Cell Death & Differentiation* 7(3), 314–326. DOI: [10.1038/sj.cdd.4400658](https://doi.org/10.1038/sj.cdd.4400658).
- Cryan JF, Markou A and Lucki I** (2002) Assessing antidepressant activity in rodents: recent developments and future needs. *Trends in Pharmacological Sciences* 23(5), 238–245. DOI: [10.1016/s0165-6147\(02\)02017-5](https://doi.org/10.1016/s0165-6147(02)02017-5).
- Cryan JF, Valentino RJ and Lucki I** (2005) Assessing substrates underlying the behavioral effects of antidepressants using the modified rat forced swimming test. *Neuroscience & Biobehavioral Reviews* 29(4–5), 547–569. DOI: [10.1016/j.neubiorev.2005.03.008](https://doi.org/10.1016/j.neubiorev.2005.03.008).
- Cryan JF, Mombereau C and Vassout A** (2005) The tail suspension test as a model for assessing antidepressant activity: review of pharmacological and genetic studies in mice. *Neuroscience & Biobehavioral Reviews* 29(4–5), 571–625. DOI: [10.1016/j.neubiorev.2005.03.009](https://doi.org/10.1016/j.neubiorev.2005.03.009).
- Cumming G** (2014) The new statistics: why and how. *Psychological Science* 25(1), 7–29. DOI: [10.1177/0956797613504966](https://doi.org/10.1177/0956797613504966).
- Cumming G, Fidler F, Leonard M, Kalinowski P, Christiansen A, Kleinig A and Wilson S** (2007) Statistical reform in psychology: is anything changing? *Psychological Science* 18(3), 230–232. DOI: [10.1111/j.1467-9280.2007.01881.x](https://doi.org/10.1111/j.1467-9280.2007.01881.x).
- da Costa Daniele TM, de Bruin PFC, Rios ERV and de Bruin VMS** (2017) Effects of exercise on depressive behavior and striatal levels of norepinephrine, serotonin and their metabolites in sleep-deprived mice. *Behavioural Brain Research* 332, 16–22. DOI: [10.1016/j.bbr.2017.05.062](https://doi.org/10.1016/j.bbr.2017.05.062).
- Dale LP, Vanderloo L, Moore S and Faulkner G** (2019) Physical activity and depression, anxiety, and self-esteem in children and youth: an umbrella systematic review. *Mental Health and Physical Activity* 16, 66–79. DOI: [10.1016/j.mhpa.2018.12.001](https://doi.org/10.1016/j.mhpa.2018.12.001).
- Davenport MH, Meah VL, Ruchat S-M, Davies GA, Skow RJ, Barrowman N and Garcia AJ** (2018) Impact of prenatal exercise on neonatal and childhood outcomes: a systematic review and meta-analysis. *British Journal of Sports Medicine* 52(21), 1386–1396. DOI: [10.1136/bjsports-2018-099836](https://doi.org/10.1136/bjsports-2018-099836).
- Davenport MH, Ruchat S-M, Poitras VJ, Garcia AJ, Gray CE, Barrowman N and Sobierajski F** (2018) Prenatal exercise for the prevention of gestational diabetes mellitus and hypertensive disorders of pregnancy: a systematic review and meta-analysis. *British Journal of Sports Medicine* 52(21), 1367–1375. DOI: [10.1136/bjsports-2018-099355](https://doi.org/10.1136/bjsports-2018-099355).
- de Oliveira LRS, Machado FSM, Rocha-Dias I, De Sousa RAL and Cassilhas RC** (2022) An overview of the molecular and physiological antidepressant mechanisms of physical exercise in animal models of depression. *Molecular Biology Reports* 49(6), 1–11. DOI: [10.1007/s11033-022-07156-z](https://doi.org/10.1007/s11033-022-07156-z).
- El-Sayes J, Harasym D, Turco CV, Locke MB and Nelson AJ** (2019) Exercise-induced neuroplasticity: a mechanistic model and prospects for promoting plasticity. *The Neuroscientist* 25(1), 65–85. DOI: [10.1177/1073858418771538](https://doi.org/10.1177/1073858418771538).
- Ellis PD** (2010) *The essential guide to effect sizes: statistical power, meta-analysis, and the interpretation of research results*. United Kingdom: Cambridge University Press.
- Emmerzaal TL, Nijkamp G, Veldic M, Rahman S, Andreatza AC, Morava E and Kozicz T** (2021) Effect of neuropsychiatric medications on mitochondrial function: for better or for worse. *Neuroscience & Biobehavioral Reviews* 127, 555–571. DOI: [10.1016/j.neubiorev.2021.05.001](https://doi.org/10.1016/j.neubiorev.2021.05.001).
- Enns GM and Cowan TM** (2017) Glutathione as a redox biomarker in mitochondrial disease—Implications for therapy. *Journal of Clinical Medicine* 6(5), 50. DOI: [10.3390/jcm6050050](https://doi.org/10.3390/jcm6050050).
- Eyre H and Baune BT** (2012) Neuroimmunological effects of physical exercise in depression. *Brain, Behavior, and Immunity* 26(2), 251–266.
- Fihler I, McMahon CA and Taylor AJ** (2009) The impact of postnatal and concurrent maternal depression on child behaviour during the early school years. *Journal of Affective Disorders* 119(1–3), 116–123. DOI: [10.1016/j.jad.2009.03.001](https://doi.org/10.1016/j.jad.2009.03.001).
- Gademan MG, Swenne CA, Verwey HF, Van Der Laarse A, Maan AC, Van De Vooren H and Cleuren GV** (2007) Effect of exercise training on autonomic derangement and neurohumoral activation in chronic heart failure. *Journal of Cardiac Failure* 13(4), 294–303. DOI: [10.1016/j.cardfail.2006.12.006](https://doi.org/10.1016/j.cardfail.2006.12.006).
- George ED, Bordner KA, Elwafi HM and Simen AA** (2010) Maternal separation with early weaning: a novel mouse model of early life neglect. *BMC Neuroscience* 11(1), 1–14. DOI: [10.1186/1471-2202-11-123](https://doi.org/10.1186/1471-2202-11-123).
- Gourgouvelis J, Yelder P and Murphy B** (2017) Exercise promotes neuroplasticity in both healthy and depressed brains: an fMRI pilot study. *Neural Plasticity*, 8305287. DOI: [10.1155/2017/8305287](https://doi.org/10.1155/2017/8305287).
- Gruhn K, Siteneski A, Camargo A, Freitas AE, Olescowicz G, Brocardo PS and Rodrigues ALS** (2021) Physical exercise stimulates hippocampal mTORC1 and FNDC5/irisin signaling pathway in mice: possible implication for its antidepressant effect. *Behavioural Brain Research* 400, 113040. DOI: [10.1016/j.bbr.2020.113040](https://doi.org/10.1016/j.bbr.2020.113040).
- Hartmann R, Schmidt FM, Sander C and Hegerl U** (2019) Heart rate variability as indicator of clinical state in depression. *Frontiers in Psychiatry* 9, 735. DOI: [10.3389/fpsy.2018.00735](https://doi.org/10.3389/fpsy.2018.00735).
- Heim C, Shugart M, Craighead WE and Nemeroff CB** (2010) Neurobiological and psychiatric consequences of child abuse and neglect. *Developmental Psychobiology* 52(7), 671–690. DOI: [10.1002/dev.20494](https://doi.org/10.1002/dev.20494).
- Hengartner MP** (2020) Editorial: antidepressant prescriptions in children and adolescents. *Frontiers in Psychiatry* 11, 600283. DOI: [10.3389/fpsy.2020.600283](https://doi.org/10.3389/fpsy.2020.600283).
- Herbsleb M, Schumann A, Lehmann L, Gabriel HH and Bär K-J** (2020) Cardio-respiratory fitness and autonomic function in patients with major depressive disorder. *Frontiers in Psychiatry* 10, 980. DOI: [10.3389/fpsy.2019.00980](https://doi.org/10.3389/fpsy.2019.00980).
- Higashi Y** (2016) Exercise is a double-edged sword for endothelial function. *Hypertension Research* 39(2), 61–63. DOI: [10.1038/hr.2015.127](https://doi.org/10.1038/hr.2015.127).
- Hoffmann A and Spengler D** (2018) The mitochondrion as potential interface in early-life stress brain programming. *Frontiers in Behavioral Neuroscience* 12, 306. DOI: [10.3389/fnbeh.2018.00306](https://doi.org/10.3389/fnbeh.2018.00306).
- Hu MX, Turner D, Generaal E, Bos D, Ikram MK, Ikram MA and Penninx BW** (2020) Exercise interventions for the prevention of depression: a systematic review of meta-analyses. *BMC Public Health* 20(1), 1–11. DOI: [10.1186/s12889-020-09323-y](https://doi.org/10.1186/s12889-020-09323-y).
- Imai S-i and Guarente L** (2014) NAD⁺ and sirtuins in aging and disease. *Trends in Cell Biology* 24(8), 464–471. DOI: [10.1016/j.tcb.2014.04.002](https://doi.org/10.1016/j.tcb.2014.04.002).
- Ji E-S, Kim Y-M, Ko YJ and Baek S-S** (2020) Treadmill exercise in obese maternal rats during pregnancy improves short-term memory through neurogenesis in the hippocampus of rat pups. *Journal of Exercise Rehabilitation* 16(5), 392–397. DOI: [10.12965%2Fjer.2040618.309](https://doi.org/10.12965%2Fjer.2040618.309).
- Kandola A, Ashdown-Franks G, Hendrikse J, Sabiston CM and Stubbs B** (2019) Physical activity and depression: towards understanding the antidepressant mechanisms of physical activity. *Neuroscience & Biobehavioral Reviews* 107, 525–539. DOI: [10.1016/j.neubiorev.2019.09.040](https://doi.org/10.1016/j.neubiorev.2019.09.040).

- Katzmarzyk PT, Friedenreich C, Shiroma EJ and Lee I-M (2022) Physical inactivity and non-communicable disease burden in low-income, middle-income and high-income countries. *British journal of Sports Medicine* 56(2), 101–106. DOI: [10.1136/bjsports-2020-103640](https://doi.org/10.1136/bjsports-2020-103640).
- Kessler RC, McLaughlin KA, Green JG, Gruber MJ, Sampson NA, Zaslavsky AM and Angermeyer M (2010) Childhood adversities and adult psychopathology in the WHO world mental health surveys. *The British Journal of Psychiatry* 197(5), 378–385. DOI: [10.1192/bjp.bp.110.080499](https://doi.org/10.1192/bjp.bp.110.080499).
- Kim Y, McGee S, Czechorz J, Walker A, Kale R, Kouzani A and Tye S (2016) Nucleus accumbens deep-brain stimulation efficacy in ACTH-pretreated rats: alterations in mitochondrial function relate to antidepressant-like effects. *Translational Psychiatry* 6(6), e842–e842. DOI: [10.1038/tp.2016.84](https://doi.org/10.1038/tp.2016.84).
- Kimmel MC, Cox E, Schiller C, Gettes E and Meltzer-Brody S (2018) Pharmacologic treatment of perinatal depression. *Obstetrics and Gynecology Clinics* 45(3), 419–440. DOI: [10.1016/j.ogc.2018.04.007](https://doi.org/10.1016/j.ogc.2018.04.007).
- Kregel KC, Allen DL, Booth FW, Fleshner MR, Henriksen EJ, Musch T and Ra'anan A (2006) Resource book for the design of animal exercise protocols. American Physiological Society.
- Kusuyama J, Alves-Wagner AB, Makarewicz NS and Goodyear LJ (2020) Effects of maternal and paternal exercise on offspring metabolism. *Nature Metabolism* 2(9), 858–872. DOI: [10.1038/s42255-020-00274-7](https://doi.org/10.1038/s42255-020-00274-7).
- Lakens D (2013) Calculating and reporting effect sizes to facilitate cumulative science: a practical primer for t-tests and ANOVAs. *Frontiers in Psychology* 4, 863. DOI: [10.3389/fpsyg.2013.00863](https://doi.org/10.3389/fpsyg.2013.00863).
- LeMoult J, Humphreys KL, Tracy A, Hoffmeister J-A, Ip E and Gotlib IH (2020) Meta-analysis: exposure to early life stress and risk for depression in childhood and adolescence. *Journal of the American Academy of Child & Adolescent Psychiatry* 59(7), 842–855. DOI: [10.1016/j.jaac.2019.10.011](https://doi.org/10.1016/j.jaac.2019.10.011).
- Lin T-W and Kuo Y-M (2013) Exercise benefits brain function: the monoamine connection. *Brain Sciences* 3(1), 39–53. DOI: [10.3390/brainsci3010039](https://doi.org/10.3390/brainsci3010039).
- Lindeque JZ, Hidalgo J, Louw R and van der Westhuizen FH (2013) Systemic and organ specific metabolic variation in metallothionein knockout mice challenged with swimming exercise. *Metabolomics* 9(2), 418–432. DOI: [10.1007/s11306-012-0459-8](https://doi.org/10.1007/s11306-012-0459-8).
- Liu Y, Heron J, Hickman M, Zammit S and Wolke D (2022) Prenatal stress and offspring depression in adulthood: the mediating role of childhood trauma. *Journal of Affective Disorders* 297, 45–52. DOI: [10.1016/j.jad.2021.10.019](https://doi.org/10.1016/j.jad.2021.10.019).
- Lu Z, Xu Y, Song Y, Bíró I and Gu Y (2021) A mixed comparisons of different intensities and types of physical exercise in patients with diseases related to oxidative stress: a systematic review and network meta-analysis. *Frontiers in Physiology* 12, 700055. DOI: [10.3389/fphys.2021.700055](https://doi.org/10.3389/fphys.2021.700055).
- Lucki I (1997) The forced swimming test as a model for core and component behavioral effects of antidepressant drugs. *Behavioural Pharmacology* 8(6-7), 523–532.
- Luo L, Li C, Du X, Shi Q, Huang Q, Xu X and Wang Q (2019) Effect of aerobic exercise on BDNF/proBDNF expression in the ischemic hippocampus and depression recovery of rats after stroke. *Behavioural Brain Research* 362, 323–331. DOI: [10.1016/j.bbr.2018.11.037](https://doi.org/10.1016/j.bbr.2018.11.037).
- Ma W, Wu JH, Wang Q, Lemaitre RN, Mukamal KJ, Djousse L and Delaney JA (2015) Prospective association of fatty acids in the de novo lipogenesis pathway with risk of type 2 diabetes: the cardiovascular health study. *The American Journal of Clinical Nutrition* 101(1), 153–163. DOI: [10.3945/ajcn.114.092601](https://doi.org/10.3945/ajcn.114.092601).
- Malkesman O and Weller A (2009) Two different putative genetic animal models of childhood depression - a review. *Progress in Neurobiology* 88(3), 153–169. DOI: [10.1016/j.pneurobio.2009.03.003](https://doi.org/10.1016/j.pneurobio.2009.03.003).
- Manji H, Kato T, Di Prospero NA, Ness S, Beal MF, Krams M and Chen G (2012) Impaired mitochondrial function in psychiatric disorders. *Nature Reviews Neuroscience* 13(5), 293–307. DOI: [10.1038/nrn3229](https://doi.org/10.1038/nrn3229).
- May LE, Scholtz SA, Suminski R and Gustafson KM (2014) Aerobic exercise during pregnancy influences infant heart rate variability at one month of age. *Early Human Development* 90(1), 33–38. DOI: [10.1016/j.earlhumdev.2013.11.001](https://doi.org/10.1016/j.earlhumdev.2013.11.001).
- McLaughlin KA, Weissman D and Bitrán D (2019) Childhood adversity and neural development: a systematic review. *Annual Review of Developmental Psychology* 1(1), 277–312. DOI: [10.1146/annurev-devpsych-121318-084950](https://doi.org/10.1146/annurev-devpsych-121318-084950).
- Memme JM, Erlich AT, Phukan G and Hood DA (2021) Exercise and mitochondrial health. *The Journal of Physiology* 599(3), 803–817. DOI: [10.1113/JP278853](https://doi.org/10.1113/JP278853).
- Molenaar NM, Kamperman AM, Boyce P and Bergink V (2018) Guidelines on treatment of perinatal depression with antidepressants: an international review. *Australian & New Zealand Journal of Psychiatry* 52(4), 320–327. DOI: [10.1177/0004867418762057](https://doi.org/10.1177/0004867418762057).
- Moyer C, Reoyo OR and May L (2016) The influence of prenatal exercise on offspring health: a review. *Clinical Medicine Insights: Women's Health* 9, CMWH.S34670. DOI: [10.4137/CMWH.S34670](https://doi.org/10.4137/CMWH.S34670).
- Naghibi S, Joneydi MS, Barzegari A, Davoodabadi A, Ebrahimi A, Eghdami E and Rostami M (2021) Treadmill exercise sex-dependently alters susceptibility to depression-like behaviour, cytokines and BDNF in the hippocampus and prefrontal cortex of rats with sporadic Alzheimer-like disease. *Physiology & Behavior* 241, 113595. DOI: [10.1016/j.physbeh.2021.113595](https://doi.org/10.1016/j.physbeh.2021.113595).
- Neumann I, Wegener G, Homberg J, Cohen H, Slattery D, Zohar J and Mathé A (2011) Animal models of depression and anxiety: what do they tell us about human condition? *Progress in Neuro-Psychopharmacology and Biological Psychiatry* 35(6), 1357–1375. DOI: [10.1016/j.pnpbp.2010.11.028](https://doi.org/10.1016/j.pnpbp.2010.11.028).
- Oberste M, Medele M, Javelle F, Lioba Wunram H, Walter D, Bloch W and Walzik D (2020) Physical activity for the treatment of adolescent depression: a systematic review and meta-analysis. *Frontiers in Physiology* 11, 185. DOI: [10.3389/fphys.2020.00185](https://doi.org/10.3389/fphys.2020.00185).
- Obi IE, McPherson KC and Pollock JS (2019) Childhood adversity and mechanistic links to hypertension risk in adulthood. *British Journal of Pharmacology* 176(12), 1932–1950. DOI: [10.1111/bph.14576](https://doi.org/10.1111/bph.14576).
- Overstreet DH and Wegener G (2013) The flinders sensitive line rat model of depression - 25 years and still producing. *Pharmacological Reviews* 65(1), 143–155. DOI: [10.1124/pr.111.005397](https://doi.org/10.1124/pr.111.005397).
- Overstreet DH, Friedman E, Mathé AA and Yadid G (2005) The flinders sensitive line rat: a selectively bred putative animal model of depression. *Neuroscience & Biobehavioral Reviews* 29(4-5), 739–759. DOI: [10.1016/j.neubiorev.2005.03.015](https://doi.org/10.1016/j.neubiorev.2005.03.015).
- Palmier-Claus J, Berry K, Bucci S, Mansell W and Varese F (2016) Relationship between childhood adversity and bipolar affective disorder: systematic review and meta-analysis. *British Journal of Psychiatry* 209(6), 454–459. DOI: [10.1192/bjp.bp.115.179655](https://doi.org/10.1192/bjp.bp.115.179655).
- Pari L and Venkateswaran S (2004) Protective role of phaseolus vulgaris on changes in the fatty acid composition in experimental diabetes. *Journal of Medicinal Food* 7(2), 204–209. DOI: [10.1089/1096620041224120](https://doi.org/10.1089/1096620041224120).
- Park H-S, Kim C-J, Kwak H-B, No M-H, Heo J-W and Kim T-W (2018) Physical exercise prevents cognitive impairment by enhancing hippocampal neuroplasticity and mitochondrial function in doxorubicin-induced chemo-brain. *Neuropharmacology* 133, 451–461. DOI: [10.1016/j.neuropharm.2018.02.013](https://doi.org/10.1016/j.neuropharm.2018.02.013).
- Park S-S, Park H-S, Kim C-J, Baek S-S and Kim T-W (2019) Exercise attenuates maternal separation-induced mood disorder-like behaviors by enhancing mitochondrial functions and neuroplasticity in the dorsal raphe. *Behavioural Brain Research* 372, 112049. DOI: [10.1016/j.bbr.2019.112049](https://doi.org/10.1016/j.bbr.2019.112049).
- Pei L and Wallace DC (2018) Mitochondrial etiology of neuropsychiatric disorders. *Biological Psychiatry* 83(9), 722–730. DOI: [10.1016/j.biopsych.2017.11.018](https://doi.org/10.1016/j.biopsych.2017.11.018).
- Picard M, Juster R-P and McEwen BS (2014) Mitochondrial allostatic load puts the 'gluc' back in glucocorticoids. *Nature Reviews Endocrinology* 10(5), 303–310. DOI: [10.1038/nrendo.2014.22](https://doi.org/10.1038/nrendo.2014.22).
- Rea E, Rummel J, Schmidt TT, Hadar R, Heinz A, Mathé AA and Winter C (2014) Anti-anhedonic effect of deep brain stimulation of the prefrontal cortex and the dopaminergic reward system in a genetic rat model of depression: an intracranial self-stimulation paradigm study. *Brain Stimulation* 7(1), 21–28. DOI: [10.1016/j.brs.2013.09.002](https://doi.org/10.1016/j.brs.2013.09.002).
- Regenass W, Möller M and Harvey BH (2018) Studies into the anxiolytic actions of agomelatine in social isolation reared rats: role of corticosterone and sex. *Journal of Psychopharmacology* 32(2), 134–145. DOI: [10.1177/0269881117735769](https://doi.org/10.1177/0269881117735769).
- Roets M, Brand L and Steyn SF (2023) Increased depressive-like behaviour of postpartum flinders sensitive and resistant line rats is reversed by a predictable postpartum stressor. *Behavioural Brain Research* 442, 114321. DOI: [10.1016/j.bbr.2023.114321](https://doi.org/10.1016/j.bbr.2023.114321).

- Rolfe D and Brown GC** (1997) Cellular energy utilization and molecular origin of standard metabolic rate in mammals. *Physiological Reviews* 77(3), 731–758. DOI: [10.1152/physrev.1997.77.3.731](https://doi.org/10.1152/physrev.1997.77.3.731).
- Ruigrok S, Yim K, Emmerzaal T, Geenen B, Stöberl N, den Blaauwen J and Kozicz T** (2021) Effects of early-life stress on peripheral and central mitochondria in male mice across ages. *Psychoneuroendocrinology* 132, 105346. DOI: [10.1016/j.psyneuen.2021.105346](https://doi.org/10.1016/j.psyneuen.2021.105346).
- Sauve AA** (2008) NAD⁺ and vitamin B3: from metabolism to therapies. *Journal of Pharmacology and Experimental Therapeutics* 324(3), 883–893. DOI: [10.1124/jpet.107.120758](https://doi.org/10.1124/jpet.107.120758).
- Scattolin MA, Resegue RM and Rosário MCd** (2022) The impact of the environment on neurodevelopmental disorders in early childhood. *Jornal de Pediatria* 98, 66–72. DOI: [10.1016/j.jped.2021.11.002](https://doi.org/10.1016/j.jped.2021.11.002).
- Schuch FB, Deslandes AC, Stubbs B, Gosmann NP, da Silva CTB and de Almeida Fleck MP** (2016) Neurobiological effects of exercise on major depressive disorder: a systematic review. *Neuroscience & Biobehavioral Reviews* 61, 1–11. DOI: [10.1016/j.neubiorev.2015.11.012](https://doi.org/10.1016/j.neubiorev.2015.11.012).
- Schuch FB, Vasconcelos-Moreno MP, Borowsky C, Zimmermann AB, Wollenhaupt-Aguiar B, Ferrari P and de Almeida Fleck MP** (2014) The effects of exercise on oxidative stress (TBARS) and BDNF in severely depressed inpatients. *European Archives of Psychiatry and Clinical Neuroscience* 264(7), 605–613. DOI: [10.1007/s00406-014-0489-5](https://doi.org/10.1007/s00406-014-0489-5).
- Schuermans C and Kurrasch D** (2013) Neurodevelopmental consequences of maternal distress: what do we really know? *Clinical Genetics* 83(2), 108–117. DOI: [10.1111/cge.12049](https://doi.org/10.1111/cge.12049).
- Seo J-H, Kim T-W, Kim C-J, Sung Y-H and Lee S-J** (2013) Treadmill exercise during pregnancy ameliorates post-traumatic stress disorder-induced anxiety-like responses in maternal rats. *Molecular Medicine Reports* 7(2), 389–398. DOI: [10.3892/mmr.2012.1197](https://doi.org/10.3892/mmr.2012.1197).
- Sharma S and Akundi RS** (2019) Mitochondria: a connecting link in the major depressive disorder jigsaw. *Current Neuropharmacology* 17(6), 550–562. DOI: [10.2174/1570159X16666180302120322](https://doi.org/10.2174/1570159X16666180302120322).
- Sohroforouzani AM, Shakerian S, Ghanbarzadeh M and Alaei H** (2022) Effect of forced treadmill exercise on stimulation of BDNF expression, depression symptoms, tactile memory and working memory in LPS-treated rats. *Behavioural Brain Research* 418, 113645. DOI: [10.1016/j.bbr.2021.113645](https://doi.org/10.1016/j.bbr.2021.113645).
- Steyn SF, Harvey BH and Brink CB** (2020) Pre-pubertal, low-intensity exercise does not require concomitant venlafaxine to induce robust, late-life antidepressant effects in flinders sensitive line rats. *European journal of neuroscience* 52(8), 3979–3994. DOI: [10.1111/ejn.14757](https://doi.org/10.1111/ejn.14757).
- Stubbs B, Vancampfort D, Rosenbaum S, Firth J, Cosco T, Veronese N and Schuch FB** (2017) An examination of the anxiolytic effects of exercise for people with anxiety and stress-related disorders: a meta-analysis. *Psychiatry Research* 249, 102–108. DOI: [10.1016/j.psychres.2016.12.020](https://doi.org/10.1016/j.psychres.2016.12.020).
- Sullivan GM and Feinn R** (2012) Using effect size—or why the P value is not enough. *Journal of Graduate Medical Education* 4(3), 279–282. DOI: [10.4300/JGME-D-12-00156.1](https://doi.org/10.4300/JGME-D-12-00156.1).
- Terburgh K, Lindeque Z, Mason S, Van der Westhuizen F and Louw R** (2019) Metabolomics of Ndufs4^{-/-} skeletal muscle: adaptive mechanisms converge at the ubiquinone-cycle. *Biochimica et Biophysica Acta (BBA) - Molecular Basis of Disease* 1865(1), 98–106. DOI: [10.1016/j.bbadis.2018.10.034](https://doi.org/10.1016/j.bbadis.2018.10.034).
- Thompson SM, Jiang L, Hammen C and Whaley SE** (2018) Association of maternal depressive symptoms and offspring physical health in low-income families. *Maternal and Child Health Journal* 22(6), 874–882. DOI: [10.1007/s10995-018-2462-9](https://doi.org/10.1007/s10995-018-2462-9).
- Tirumalaraju V, Suchting R, Evans J, Goetzl L, Refuerzo J, Neumann A and Cowen PJ** (2020) Risk of depression in the adolescent and adult offspring of mothers with perinatal depression: a systematic review and meta-analysis. *JAMA Network Open* 3(6), e208783–e208783. DOI: [10.1001/jamanetworkopen.2020.8783](https://doi.org/10.1001/jamanetworkopen.2020.8783).
- U.S. Food & Drug Administration**. 2004. *FDA patient safety news: Show #34, December 2004 - “Black box” warning for antidepressants*. <http://www.fda.gov/downloads/safety/fdapatientsafetynews/ucm417804.pdf>. Accessed: December 14, 2004.
- van Rensburg D, Lindeque Z, Harvey BH and Steyn SF** (2022) Reviewing the mitochondrial dysfunction paradigm in rodent models as platforms for neuropsychiatric disease research. *Mitochondrion* 64, 82–102. DOI: [10.1016/j.mito.2022.03.002](https://doi.org/10.1016/j.mito.2022.03.002).
- Viswanathan, M., Kennedy, S.M., McKeeman, J., Christian, R., Coker-Schwimmer, M., Middleton, J.C., ... Forman-Hoffman, V.** 2020. *Treatment of depression in children and adolescents: a systematic review*. https://effectivehealthcare.ahrq.gov/sites/default/files/Evidence%20Summary_0.pdf Accessed October 31, 2020.
- Warrack BM, Hnatyshyn S, Ott K-H, Reily MD, Sanders M, Zhang H and Drexler DM** (2009) Normalization strategies for metabolomic analysis of urine samples. *Journal of Chromatography B* 877(5-6), 547–552. DOI: [10.1016/j.jchromb.2009.01.007](https://doi.org/10.1016/j.jchromb.2009.01.007).
- Wegner M, Amatriain-Fernández S, Kaulitzky A, Murillo-Rodríguez E, Machado S and Budde H** (2020) Systematic review of meta-analyses: exercise effects on depression in children and adolescents. *Frontiers in Psychiatry* 11, 81. DOI: [10.3389/fpsy.2020.00081](https://doi.org/10.3389/fpsy.2020.00081).
- White AT and Schenk S** (2012) NAD⁺/NADH and skeletal muscle mitochondrial adaptations to exercise. *American Journal of Physiology-Endocrinology and Metabolism* 303(3), E308–E321. DOI: [10.1152/ajpendo.00054.2012](https://doi.org/10.1152/ajpendo.00054.2012).
- Whitney A, Lindeque JZ, Kruger R and Steyn SF** (2023) Genetically predisposed and resilient animal models of depression reveal divergent responses to early-life adversity. *Acta Neuropsychiatrica* 1–13. DOI: [10.1017/neu.2023.37](https://doi.org/10.1017/neu.2023.37).
- Wu H, Southam AD, Hines A and Viant MR** (2008) High-throughput tissue extraction protocol for NMR-and MS-based metabolomics. *Analytical Biochemistry* 372(2), 204–212. DOI: [10.1016/j.ab.2007.10.002](https://doi.org/10.1016/j.ab.2007.10.002).
- Wu T, Huang Y, Gong Y, Xu Y, Lu J, Sheng H and Ni X** (2019) Treadmill exercise ameliorates depression-like behavior in the rats with prenatal dexamethasone exposure: the role of hippocampal mitochondria. *Frontiers in Neuroscience* 13, 264. DOI: [10.3389/fnins.2019.00264](https://doi.org/10.3389/fnins.2019.00264).
- Xiang K, Qin Z, Zhang H and Liu X** (2020) Energy metabolism in exercise-induced physiologic cardiac hypertrophy. *Frontiers in Pharmacology* 11, 1133. DOI: [10.3389/fphar.2020.01133](https://doi.org/10.3389/fphar.2020.01133).
- Zeng M, Che Z, Liang Y, Wang B, Chen X, Li H and Zhou Z** (2009) GC-MS based plasma metabolic profiling of type 2 diabetes mellitus. *Chromatographia* 69(9-10), 941–948. DOI: [10.1365/s10337-009-1040-0](https://doi.org/10.1365/s10337-009-1040-0).
- Zhu X-H, Qiao H, Du F, Xiong Q, Liu X, Zhang X and Chen W** (2012) Quantitative imaging of energy expenditure in human brain. *NeuroImage* 60(4), 2107–2117. DOI: [10.1016/j.neuroimage.2012.02.013](https://doi.org/10.1016/j.neuroimage.2012.02.013).
- Zitkovsky EK, Daniels TE and Tyrka AR** (2021) Mitochondria and early-life adversity. *Mitochondrion* 57, 213–221. DOI: [10.1016/j.mito.2021.01.005](https://doi.org/10.1016/j.mito.2021.01.005).